Tropical Journal of Pharmaceutical Research March 2024; 23 (3): 491-499 ISSN: 1596-5996 (print); 1596-9827 (electronic) © Pharmacotherapy Group, Faculty of Pharmacy, University of Benin, Benin City, 300001 Nigeria.

> Available online at http://www.tjpr.org http://dx.doi.org/10.4314/tjpr.v23i3.2

**Original Research Article** 

# *In vitro* cytotoxic activity of *Sansevieria trifasciata* against various cancer cell lines

# Sheryar Afzal<sup>1\*</sup>, Yuan Seng Wu<sup>2</sup>, Chan Zelynn<sup>1</sup>, V Appalaraju<sup>2</sup>

<sup>1</sup>Department of Biomedical Sciences, College of Veterinary Medicine, King Faisal University, Al Hufof, Saudi Arabia, <sup>2</sup>School of Medical and Life Sciences, Sunway University, 47500 Subang Jaya, Selangor, Malaysia

\*For correspondence: Email: safzal@kfu.edu.sa, sengwu\_21@yahoo.com; Tel: +966563840308, +60163720821

Sent for review: 16 November 2023

Revised accepted: 27 February 2024

# Abstract

**Purpose:** To determine the in vitro anticancer properties of Sansevieria trifasciata extracts. **Methods:** The cytotoxic activity of n-hexane, ethyl acetate and methanol leaf extracts at different concentrations was assessed against Vero, pancreatic (AsPC-1), lung (A549), esophageal squamous carcinoma (KYSE 150) and rhabdomyosarcoma (RD) cancer cell lines using MTT assay. The apoptosis-inducing potential of selected S. trifasciata leaf extract was determined using a caspase 3/7 assay kit.

**Results:** Ethyl acetate extract of S. trifasciata leaf showed significant anticancer activity against RD cells and methanol extract against KYSE cells (p < 0.05). Ethyl acetate and methanol extract possessed higher toxicity towards RD and KYSE 150 cell lines and exhibited relatively lower toxicity towards Vero normal cells. Ethyl acetate extract-treated RD cells and KYSE 150 cancer cells had more typical apoptotic morphologic features when compared to normal cells.

**Conclusion:** Sansevieria trifasciata possesses anticancer activity and provides new insights for future molecular work on investigating its utilization in cancer treatment.

Keywords: Sansevieria trifasciata, Cancer cell lines, Apoptosis, Chemotherapeutic potential, MTT assay

This is an Open Access article that uses a funding model which does not charge readers or their institutions for access and distributed under the terms of the Creative Commons Attribution License (http://creativecommons.org/licenses/by/4.0) and the Budapest Open Access Initiative (http://www.budapestopenaccessinitiative.org/read), which permit unrestricted use, distribution, and reproduction in any medium, provided the original work is properly credited.

Tropical Journal of Pharmaceutical Research is indexed by Science Citation Index (SciSearch), Scopus, Web of Science, Chemical Abstracts, Embase, Index Copernicus, EBSCO, African Index Medicus, JournalSeek, Journal Citation Reports/Science Edition, Directory of Open Access Journals (DOAJ), African Journal Online, Bioline International, Open-J-Gate and Pharmacy Abstracts

## INTRODUCTION

Worldwide, cancer continues to be the second most common cause of death. The International Agency for Research on Cancer reported 19.3 million new cancer cases and almost 10.0 million deaths worldwide in 2020, according to Global Cancer Observatory (GLOBOCAN) data. Cancer cases are further expected to rise by over 50 %, with 32.2 million cases in 2040 [1]. Even with certain notable advancements in cancer treatment, adverse side effects and medication resistance continue to produce unsatisfactory therapeutic results. Thus, the need for other sources to address the problems stated is a global priority [2].

Cancer chemoprevention agents with natural phytochemical compounds are recognized as an emerging approach to prevent, delay, impede, or control malignancy, especially for advanced metastasized cancer. Many cultures throughout

© 2024 The authors. This work is licensed under the Creative Commons Attribution 4.0 International License

the world still rely on indigenous medicinal plants to alleviate various health problems [3] and 60 to 75 % of FDA-approved anti-infectious and anticancer drugs are proven to be derived from natural products of plant origin [4]. Sansevieria member of trifasciata. а the family Asparagaceae, is used in the treatment of various diseases such as urinary diseases, diabetes mellitus, pharyngitis, earache, skin itch, and also possesses analgesic, antipyretic, antioxidants along with antibacterial properties [5].

Apoptosis, one of the primary types of cell death, is characterized by a variety of unique and biochemical morphological alterations. including chromatin condensation, membrane blebbing, cell shrinkage, and the formation of apoptotic bodies [6]. Because cancer can result from a lack of apoptotic regulation, apoptosis plays a crucial function in cancer treatment [7]. Despite the many therapeutic potentials that S. trifasciata has demonstrated, relatively few studies concerning the cytotoxic activity of S. trifasciata in contrast to various cancer cell lines have been reported. Therefore, this study investigates the potential anticancer activity of S. trifasciata.

## **EXPERIMENTAL**

#### Preparation of plant extract

Fresh leaves of S. trifasciata were harvested from a local field close to the School of Medical and Life Sciences, Sunway University, Selangor, Malavsia and were authenticated bv а taxonomist (Mohd Hafizi Adzmi Hanafi, reference no. KM0016/22, at the Institute of Bioscience, Putra Malaysia (UPM), Universiti Seri Kembangan, Malaysia. The leaves were dried and pulverized. Thereafter, approximately 200 g of the pulverized leaf was extracted sequentially in a Soxhlet apparatus with solvents of different polarities such as n-hexane, ethyl acetate and methanol. The amount of solvent used to wet the plant sample and fill the round bottom flask was set to a quantitative relation of approximately 1:2. in which two parts of solvent were utilized for one part of plant material. The selected solvent (100 mL) was transferred into the round bottom flask, followed by a small amount of boiling chips. The resulting extract was concentrated in a rotating device, filtered through a 0.22 µm filter, and stored at -20 °C in a dark glass bottle. After the extract was dissolved in DMSO, the final DMSO concentration was adjusted to less than 0.1 %. The leaf extraction or percentage yield (Y) was determined using Eq 1.

Y (%) = (A/W)100 .....(1)

where A is the actual yield and W is the weight of plant.

#### Preliminary phytochemical analysis

Leaf extracts of *S. trifasciata* were subjected to preliminary identification of phytochemical constituents using standard procedures [8-9].

#### Cell culture

Pancreatic cancer cells (AsPC-1), lung cancer cells (A549), esophageal squamous carcinoma cells (KYSE 150), and rhabdomyosarcoma cancer cells (RD) were provided by Professor Dr. Poh Chit Laa of Sunway University's School of Medical and Life Sciences' Centre for Virus and Vaccine Research. American Type Culture Collection provided Vero cells. At 37 °C in a humidified atmosphere with 5 % CO<sub>2</sub>, cells were cultured in DMEM (GIBCO, USA) supplemented with 10 % heat-inactivated FBS and 100 units/mL of pen/strep (100 µg/mL) as the antibiotic source.

#### Cell viability assay

S. trifasciata extracts were evaluated for their impact on viability of Vero, AsPC-1, A549, KYSE150, and RD cells. In 96-well plates, cells were cultivated at a density of  $1 \times 10^4$  per well and incubated until 80 % adherence was achieved. Then cells were treated with different concentrations of S. trifasciata extract (62.5, 125, 250 and 500 µg/mL). The cells of control group were incubated with 100 µL of 2 % DMEM. After 24 h, 20 µL of MTT reagents (Macklin, Canada) were added in the dark at 37 °C under 5 % CO<sub>2</sub> for 4 h. Subsequently, 100 µL of 10 % sodium dodecyl sulfate/0.01 M hydrochloric acid was added to each well to solubilize the formazan crystals formed. Using a microplate reader, the absorbance was measured at 570 nm with a reference wavelength of 630 nm. (Infinite® F50 Robotic; Tecan, Männedorf, Switzerland).

#### Morphological analysis

The KYSE 150 and RD cells were treated with various concentrations of ethyl acetate extract (31.25 and 125  $\mu$ g/mL) for 2 h in 96-well plates, using untreated cells as negative control. Following treatment, phosphate-buffered saline (PBS) was used to wash the cells, and the morphological changes of treated KYSE 150 and RD cells were observed under an inverted light microscope.

#### Apoptosis analysis by Caspase 3/7 assay

The KYSE 150 and RD cells were treated with various concentrations of ethyl acetate extract (15.625 and 31.25  $\mu$ g/mL) and untreated cells as negative control. Utilizing 100  $\mu$ L of Caspase-Glo® 3/7 reagent, which was added to each well and allowed to incubate for an hour at room temperature, apoptosis triggered by ethyl acetate extract was identified. The signals produced were read using a microplate reader.

#### Statistical analysis

All tests were conducted in duplicates in two independent experiments. Using GraphPad Prism version 8.0, the data were analyzed using ANOVA and Bonferroni post hoc test. Values were reported as mean  $\pm$  standard deviation (SD). Statistical significance was assigned to values with (p < 0.05).

## RESULTS

Percentage yield of the extraction of S. trifasciata leaf

The yield of the leaf extraction is displayed in Table 1.

# Preliminary phytochemical classes analysis of extract

The results of phytochemical screening are summarized in Table 2.

# In vitro cytotoxic activity of S. trifasciata leaf extract

Table 3 displays the findings of extract's cytotoxic activities. KYSE 150 cell line was the

most sensitive to treatment of hexane extract (IC<sub>50</sub>: 13.8  $\pm$  0 µg/mL), while hexane extract exhibited weak or inactive cytotoxic activity against A549, RD and AsPC-1 cell lines.

Ethyl acetate extract possessed significant cytotoxic activity against RD (IC<sub>50</sub>: 19.09  $\pm$  12.86 µg/mL), and KYSE 150 cells (IC<sub>50</sub>: 20.22  $\pm$  1.65 µg/mL), while insignificant against A549 and AsPC-1 cells. Lastly, KYSE 150 was the most sensitive to the treatment of methanol extract (IC50: 14.80  $\pm$  0.49 µg/mL), while the extract exhibited weak or inactive cytotoxicity against RD, A549 and AsPC-1 cells.

# Assessing the selectivity of *S. trifasciata* leaf extract using normal cell lines

The cytotoxicity of *S. trifasciata* leaf extract was determined in the cancer cell lines as well as the normal cell line. The findings are shown in Table 4. The ethyl acetate and methanol extract have higher toxicity towards RD and KYSE 150 cells and exhibit relatively lower toxicity towards Vero normal cells.

Therefore, RD cells and KYSE 150 cells were selected for further experiments to determine the presence of apoptosis by measuring Caspase 3/7 levels.

#### Apoptosis of RD and KYSE 150 cells

As determined by the MTT assay (Figure 1) and selectivity index (SI), ethyl acetate extract which exhibited noticeable cytotoxic activity and relatively higher selectivity against RD and KYSE 150 cancer cell lines was then employed in further investigations to determine the apoptotic activity.

Table 1: Weight of extraction, percentage of yield, color and solidity of S. trifasciata leaf extract

Extract	Weight of extract (g)	Yield (%)	Colour	Consistency
n-hexane	3.95	1.98	Dark green	Semi-solid
Ethyl acetate	7.63	3.82	Dark green	Semi-solid
Methanol	14.27	7.14	Dark brown	Semi-solid

Table 2: Phytochemical analysis testing of different extracts of Sansevieria trifasciata

Constituent of detection	Types of biochemical test	Results						
constituent of detection	Types of biochemical test	n- hexane	Ethyl acetate	Methanol				
Flavonoid	Alkaline	+	-	-				
Alkaloid	Wagner's reagent	+	-	-				
Phenolic	Ferric chloride	+	+	-				
Terpenoid	Salkowski	-	-	-				
Tannin	Ferric chloride	+	+	-				
Glycoside	Modified Borntrager (C-glycoside)	+	+	+				
Saponin	Foam	-	-	+				

*Note:* –: not present, +: present in low concentration

#### Afzal et al

Table 3: IC<sub>50</sub> values (µg/mL) of *S. trifasciata* leaf extract against cancer and normal cell lines as determined by MTT assay

Extract		Cancer cell lines													Normal cell line		
	RD			KYSE 150			A549			AsPC-1			Vero				
	1	2	Mean ± SD (µg/mL)	1	2	Mean ± SD (µg/mL)	1	2	Mean ± SD (µg/mL)	1	2	Mean±SD (µg/mL)	1	2	Mean±SD (µg/mL)		
Hexane	186.21	190.55	188.38±3.07	13.8	13.8	13.8±0.00	91.2	131.83	111.52±28.73	-	-	-	13.8	14.45	14.13±0.46		
Ethyl acetate	10	28.18	19.09±12.86	19.05	21.38	20.22±1.65	69.18	93.33	81.26±17.08	83.18	109.62	96.4±18.7	144 .54	158.49	151.52±9.86		
Methanol	32.36	33.11	32.74±0.53	15.14	14.45	14.80±0.49	91.2	102.33	96.77±7.87	52.48	52.48	52.48±0	436.52	489.78	463.15±37.66		

**Table 4:** Selectivity index (SI) of *S. trifasciata* leaf extract calculated by the IC<sub>50</sub> against Vero cells and cancer cell lines. Data were obtained from two independent experiments and presented as mean ± standard deviation (SD)

		Cancer cell lines										
Extract	RD			KYSE 150			A549			AsPC-1		
	1	2	Mean ± SD	1	2	Mean ± SD	1	2	Mean± SD	1	2	Mean± SD
Hexane	1.16	1.16	1.16± 0.004	1	1	1±0	0.58	0.54	0.56± 0.03	-	-	-
Ethyl acetate	2.2	1.52	1.86± 0.48	1.72	1.65	1.69± 0.05	1.20	1.12	1.16± 0.06	1.15	1.08	$1.11 \pm 0.05$
Methanol	1.75	1.74	1.74± 0.01	2.24	2.28	$2.26 \pm 0.03$	1.35	1.31	1.33± 0.02	1.53	1.53	1.53±0

Afzal et al



**Figure 1**: Percentage of (A) Vero, (B) RD, (C) KYSE 150, (D) AsPC-1 and (E) A549 cells viability inhibition after treating with different concentrations of hexane, ethyl acetate and methanol extract of *S. trifasciata* against as evaluated by MTT assay. *Note:* \*P < 0.05 versus concentration at 0 µg/mL (Control group); #P < 0.05 between hexane and ethyl acetate extract;  $^{\delta}P < 0.05$ , between hexane and methanol extract;  $^{\phi}P < 0.05$  between ethyl acetate and methanol extract.

# Morphological changes induced by ethyl acetate extract of *S. trifasciata* leaf in KYSE 150 and RD cells

The morphological changes induced are shown in Figure 2. Two hours of treatment of KYSE 150 and RD cells with 31.25  $\mu$ g/mL and 125  $\mu$ g/mL (concentrations causing 50% cell viability inhibition) reduced cell number and cell membrane blebbing due to loss of cellular viability.

The onset of apoptosis was observed which was characterized by shrinkage of cells and nucleus, followed by detachment of cells from the surrounding tissue. The cells became convoluted and formed extensions called budding. The formation of apoptotic bodies with the cellular organelles and fragments of nuclei closely packed inside the cells was also recognized in Figure 3 and Figure 4.

## DISCUSSION

Cancer is a devastating disease that has become one of the world's most pressing health concerns [10]. Several clinically approved cancer treatments are available [11]. However, the of conventional chemotherapeutic agents is often





Concentration (µg/mL)

**Figure 2:** Apoptotic (Caspase 3/7) activity in (A) KYSE 150 (B) RD cells treated with 15.625 and 31.25  $\mu$ g/mL of ethyl acetate extract for 2 h. \**P* < 0.05 versus control.



Figure 3: Changes in morphology caused by *S. trifasciata* ethyl acetate extract in (A) KYSE 150 cells, (B) at 31.25  $\mu$ g/mL and (C) at 125  $\mu$ g/mL after 2 h treatment compared with untreated control. Magnification: 10X



**Figure 4:** Changes in morphology brought upon by the *S. trifasciata* ethyl acetate extract in RD cells at (B) 31.25  $\mu$ g/mL and (C) 125  $\mu$ g/mL after 2 h treatment compared with untreated controls (A). Magnification: 10x

Trop J Pharm Res, March 2024; 23(3): 496

accompanied by harmful side effects and chemo-resistance. Hence, it is crucial to seek a novel anticancer agent with high efficacy and low toxicity.

Sansevieria trifasciata is important in ethnomedicine for the treatment of several ailments [12]. Its anti-inflammatory activities have been demonstrated [13]. However, relatively few studies concerning the cytotoxic activity of S. trifasciata leaf extract against different cancer cell lines have been reported. The phytochemical screening of these extracts indicated the presence of several phytochemical classes that known to possess strong anti-cancer are properties and combat various diseases through specific modes of action [14].

Based on the result obtained, the hexane and methanol extract from the leaves of *S. trifasciata* exhibited a noticeable cytotoxic effect on KYSE 150 cancer cell lines while ethyl acetate extract displayed a higher cytotoxic effect on RD cells and KYSE 150 cancer cell lines. The low cytotoxicity of the plant extract against A549 and AsPC-1 cancer cells may be due to the resistance of the cancer cell lines to the type of secondary metabolites present.

It has been reported that the selectivity index (SI) > 1.0 is favorable as it indicates that treatment with extract has greater efficacy against cancer cells with low toxicity against normal cells [15], which would be a safer anticancer drug candidate for further testing in vivo models. Merely utilizing malignant cell lines to assess the anticancer potential of plant extract without identifying SI is a subpar predictor for subsequent investigations. The ideal anticancer medication should be able to eradicate cancer cells without endangering healthy, normal cells. Because traditional chemotherapeutic medicines lack selectivity and are extremely toxic to both cancer and healthy normal tissues, their usage is frequently followed by chemo-resistance and unpleasant side effects [2,16]. In the result obtained, the SI values of hexane extract on A549 and AsPC-1 cells were less than one, which can be toxic to Vero normal cells and cannot be used as an herbal drug. In contrast, ethyl acetate and methanol extract of S. trifasciata had comparatively higher SI values on RD and KYSE 150 cancer cell lines, which are presumably non-toxic and bioactive. Certain hazardous components are present in the extracts with comparatively low SI (< 1) data. Comparative analysis of MTT assay results suggested that the cytotoxic effect of ethyl acetate and methanol extract of S. trifasciata can be related to secondary metabolites such as polyphenols, flavonoids and alkaloids which were important for anticancer activity, especially saponin [12]. However, the ethyl acetate extract was selected to determine the apoptotic activity against RD and KYSE 150 cancer cells as it contains various phytochemicals as compared to methanol extract.

Known as "programmed cell death," apoptosis is a widely used process for tissue remodeling, cell replacement, and the removal of damaged cells. Anticancer drugs cause apoptosis, which decreases the sensitivity of treatment when it is disrupted [11]. Apoptosis is one of the key underlying mechanisms for cytotoxic antitumor agents, whereas studies have found that some natural compounds present in plants can induce apoptotic pathways that are blocked in cancer cells [17]. Ethyl acetate extract of S. trifasciata at 15.625 and 31.25 µg/mL was selected for cell death detection based on the results of the MTT assay for caspase 3/7. In this study, the apoptotic activity in KYSE 150 cells induced by ethyl acetate extract was the highest at 15.625 µg/mL as compared to 31.25 µg/mL, and this trend was similar to that of RD cell activity.

Under a light microscope, the morphological alterations of RD and KYSE 150 cells were examined to ascertain if the growth inhibitory action of S. trifasciata plant extract was connected to the induction of apoptosis. The American National Cancer Institute's (NCI) protocol states that a crude plant extract's upper limit for its IC<sub>50</sub> is 30 µg/mL [19]. Nevertheless, it was discovered that after being exposed to 125 µg/mL and ethyl acetate extract (31.25) for two hours, the cells began to generate apoptotic Typical apoptotic morphological bodies. observation included cell rounding, membrane blebbing, and formation of several apoptotic bodies were observed, compared to homogeneous nuclear chromatin in the control cells [11]. Taken together, this finding revealed that ethyl acetate extract of S. trifasciata induces apoptosis in RD and KYSE 150 cells. Therefore, the beneficial role of S. trifasciata leaf extract in preventing apoptotic cell death is proposed. Future mechanistic research is necessary to determine whether the lethal impact of S. trifasciata extract on cancer cells is primarily caused by apoptosis induction or cell cycle arrest.

#### CONCLUSION

Ethyl acetate extract of *S. trifasciata* leaf shows potent anticancer activity against RD cells while ethyl acetate and methanol extract have the highest potency against KYSE 150 cells. The present investigation provides evidence that *S. trifasciata* leaf extract may be an effective anticancer drug and justifies its use in ethnomedical settings.

#### DECLARATIONS

#### Acknowledgements

This work was supported by the Deanship of Scientific Research, Vice Presidency for Graduate Studies and Scientific Research, King Faisal University, Saudi Arabia (Grant no. 5,965). The authors are thankful to the Faculty of Pharmacy, Mahsa University and Centre for Virus and Vaccine Research, School of Medical and Life Sciences, Sunway University.

#### Funding

None provided.

#### Ethical approval

None provided.

#### Availability of data and materials

The datasets used and/or analyzed during the current study are available from the corresponding author on reasonable request.

#### **Conflict of Interest**

No conflict of interest associated with this work.

#### **Contribution of Authors**

We declare that this work was done by the authors named in this article and all liabilities pertaining to claims relating to the content of this article will be borne by the authors. Sheryar Afzal and V Appalaraju Study concept and design, manuscript handling and manuscript writing. Chan Zelynn performed experiments, data collection, and analysis, a major contribution to manuscript writing. Yuan Seng Wu supervised and designed the project, analysis and critical revision of the manuscript. All authors read and approved the final draft of the manuscript for publication.

#### **Open Access**

This is an Open Access article that uses a funding model which does not charge readers or their institutions for access and distributed under the terms of the Creative Commons Attribution License (http://creativecommons.org/licenses/by/ 4.0) and the Budapest Open Access Initiative (http://www.budapestopenaccessinitiative.org/rea d), which permit unrestricted use, distribution, and reproduction in any medium, provided the original work is properly credited.

## REFERENCES

- Sung H, Ferlay J, Siegel RL, Laversanne M, Soerjomataram I, Jemal A, Global cancer statistics 2020: Globocan estimates of incidence and mortality worldwide for 36 cancers in 185 countries. CA Cancer J Clin 2021; 71(3): 209-249.
- Nguyen NH, Pham QT, Luong TNH, Phung VT, Duong TH. Anticancer activity of novel plant extract and compounds from Adenosma bacterium (Bonati) in human lung and liver cancer cells. Molecul 2020; 25(12): 2912.
- Ighodaro A, Anegbe B. The phytochemical and chemotherapeutic effect of three indigenous African plants used in asthma therapy. J Pharmacog Phytochem 2015; 3(6): 244-247.
- Chanda S, Nagani K. In vitro and in vivo methods for anticancer activity evaluation and some Indian medicinal plants possessing anticancer properties: an overview. J Pharmacog Phytochem 2013; 2(2): 140-152.
- Anand U, Jacobo-Herrera N, Alternimi A, Lakhssassi N. A comprehensive review on medicinal plants as antimicrobial therapeutics: potential avenues of biocompatible drug discovery. Metabolites 2019; 9(11): 258.
- Cohen GM. Caspases: the executioners of apoptosis. Biochem J 1997; 326(1): 1-16.
- Seca AM, Pinto DC. Plant secondary metabolites as anticancer agents: successes in clinical trials and therapeutic application. Int J Mol Sci 2018; 19(1): 263.
- Shaikh JR, Patil M. Qualitative tests for preliminary phytochemical screening: An overview. Int J Chem Stu 2020; 8(2): 603-608.
- El Aziz M, Ashour A, Melad AG. A review on saponins from medicinal plants: chemistry, isolation, and determination. J Nanomed Res 2019; 8(1): 282-288.
- Lerma-Herrera MA, Beiza-Granados L, Ochoa-Zarzosa A, López-Meza JE, Hernández-Hernández JD, Aviña-Verduzco J, et al. In vitro cytotoxic potential of extract from Aristolochia foetida Kunth against MCF-7 and bMECs cell lines. Saudi J Biol Sci 2021; 28(12): 7082-7089.
- 11. Shahneh FZ, Valiyari S, Azadmehr A, Hajiaghaee R, Yaripour S, Bandehagh A. Inhibition of growth and induction of apoptosis in fibrosarcoma cell lines by Echinophora platyloba DC: in vitro analysis. Adv Pharmacol Pharm Sci 2013; 2013.
- Abdul-Hafeez EY, Orabi MA, Ibrahim OH, Ilinskaya O, Karamova NS. In vitro cytotoxic activity of certain succulent plants against human colon, breast and liver cancer cell lines. S Afr J Bot 2020; 131: 295-301.

Trop J Pharm Res, March 2024; 23(3): 498

- Pinky SS, Monira S, Hossain MA, Hossain A. Antioxidant, Anti-inflammatory, Cytotoxic and Analgesic Activities of Sensevieria trifasciata. Banglad Pharm J 2020; 23(2): 195-200.
- Nayim P, Sudhir K, Mbaveng AT, Kuete V, Sanjukta M. In vitro anticancer activity of Imperata cylindrical root extract toward human cervical cancer and identification of potential bioactive compounds. BioMed Res Int 2021, 1-12.
- 15. Krzywik J, Mozga W, Aminpour M, Janczak J, Maj E, Wietrzyk J. Synthesis, antiproliferative activity and

molecular docking studies of novel doubly modified colchicine amides and sulfonamides as anticancer agents. Molecul 2020; 25(8): 1789.

- Iqbal J, Abbasi BA, Mahmood T, Kanwal S, Ali B, Shah SA. Plant-derived anticancer agents: A green anticancer approach. Asian Pacific J Trop Biomed 2017; 7(12): 1129-1150.
- 17. Safarzadeh E, Shotorbani SS, Baradaran B. Herbal medicine as inducers of apoptosis in cancer treatment. Adv Pharm Bull 2014, 4(Suppl 1): 421.