

Review Article

Thiolated Chitosans: Novel Polymers for Mucoadhesive Drug Delivery – A Review

SA Sreenivas^{1*} and KV Pai²

¹Department of Pharmaceutics, KLES's College of Pharmacy, Vidyanagar, Hubli, Karnataka. ²Department of Industrial Chemistry, Shankaraghatta, Shimoga, Karnataka, India.

Abstract

Chitosan is a natural polycationic copolymer consisting of glucosamine and N-acetylglucosamine units. The polymer has valuable properties as a biomaterial because it is considered to be biocompatible, biodegradable and non-toxic. The purpose of this review article is to provide detailed information about thiolated chitosans which are gaining popularity because of their high mucoadhesiveness and extended drug release properties. The derivatization of the primary amino groups of chitosan with coupling reagents bearing thiol functions leads to the formation of thiolated chitosans. Various properties of chitosan are improved by the immobilization of thiol groups. Due to the formation of disulfide bonds with mucus glycoproteins, mucoadhesiveness is augmented. The permeation of paracellular markers through mucosa can be enhanced by utilizing thiolated instead of unmodified chitosan. Moreover, thiolated chitosans display in situ gelling features due to the pH-dependent formation of inter- as well as intra-molecular disulfide bonds. This latter process provides, strong cohesion and stability of carrier matrices, being based on thiolated chitosans. The in situ gel formation within the pH range of 5 to 6.8 makes the application of thiolated chitosans on vaginal, nasal and ocular mucosa also possible. Thiolated chitosans can guarantee prolonged controlled release of embedded therapeutic ingredients.

Key words: Thiolated chitosan, Thiomers, Mucoadhesion, Permeation enhancement, In situ gelation

*Corresponding author: Email: saiseenu7@rediffmail.com; Tel: 09242892034

INTRODUCTION

Mucoadhesive drug delivery systems

Mucoadhesion can be defined as the ability of synthetic or biological macromolecules to adhere to mucosal tissues such as the mucosa of the small intestine. Since the early 1980s, the concept of mucoadhesion has gained considerable interest in pharmaceutical technology. If this concept can reach its full potential, it might open the door for novel, highly efficient dosage forms especially for oral drug delivery. Mucoadhesive drug delivery systems promise several advantages that arise from localization at a given target site, prolonged residence time at the site of drug absorption, and an intensified contact with the mucosa increasing the drug concentration gradient¹. Hence, uptake and consequently bioavailability of the drug may be increased and frequency of dosing reduced with the result that patient compliance is improved. Various natural and synthetic polymers have been discovered as mucoadhesive excipients. Their mucoadhesive properties can be explained by their interaction with the glycoproteins of the mucus, based mainly on non-covalent bonds such as ionic interactions, hydrogen bonds and van der Waals forces².

Chitosan

The biopolymer chitosan is obtained by alkaline deacetylation of chitin, which is one of the most abundant polysaccharides in nature. Shell wastes of shrimp, lobster and crab are the main industrial sources of chitin³. Chitosan is a polysaccharide consisting of copolymers of glucosamine and N-acetylglucosamine. The primary amino group accounts for the possibility of relatively easy chemical modification of chitosan and salt formation with acids. At acidic pH, the amino groups are protonated, which promotes solubility, whereas chitosan is insoluble at alkaline and neutral pH⁴. Because of its favorable properties, such as enzymatic biodegradability, non-toxicity and biocompatibility³, chitosan has received considerable attention as a novel excipient in drug delivery systems, and has been included in the European Pharmacopoeia since 2002. So far, chitosan

has been utilized in various fields of pharmaceutical technology, including the formulation of controlled release dosage forms such as tablets, gels and microspheres, as mucoadhesive and/or permeation enhancing excipient for oral, nasal, ocular and buccal drug delivery⁵⁻⁸ and in non-viral gene delivery^{9,10}.

Thiolated chitosans

Recently, it has been shown that polymers with thiol groups provide much higher adhesive properties than polymers generally considered to be mucoadhesive. The enhancement of mucoadhesion can be explained by the formation of covalent bonds between the polymer and the mucus layer which are stronger than non-covalent bonds. These thiolated polymers (see Fig. 1), known as *thiomers*, interact with cysteine-rich subdomains of mucus glycoproteins via disulfide exchange reactions¹¹ or via simple oxidation process as shown in Fig. 2.

To further enhance the solubility of chitosan and to improve its mucoadhesive and/or permeation enhancing properties, various derivatives such as trimethylated chitosan,¹² mono-N-carboxymethyl chitosan¹³, N-sulfochitosan¹⁴ and chitosan-EDTA conjugates¹⁵ were developed. A further modification is based on the immobilization of thiol bearing moieties on the polymeric backbone of chitosan. To date, three different thiolated chitosan derivatives have been synthesized: chitosan-thioglycolic acid conjugates^{16,17}, chitosan-cysteine conjugates¹⁸ and chitosan-4-thio-butyl-amidine (chitosan-TBA) conjugates¹⁹.

These thiolated chitosans have numerous advantageous features in comparison to unmodified chitosan, such as significantly improved mucoadhesive and permeation enhancing properties¹⁸⁻²¹. The strong cohesive properties of thiolated chitosans make them highly suitable excipients for controlled drug release dosage forms^{19,22}. Moreover, solutions of thiolated chitosans display *in situ* gelling properties at physiological pH values¹⁷.

It is the aim of this review to provide an overview about different thiolated chitosan derivatives that have been synthesized so far,

as well as their characterization and optimization utilizing various *in vitro* test systems. The performance of thiolated chitosan in *in vivo* studies, providing proof of their applicability in peroral peptide delivery systems, will be discussed as well.

Synthesis of thiolated chitosans

The primary amino group at the 2-position (Fig. 3) of the glucosamine subunit of chitosan is the main target for the immobilization of thiol groups. As shown in Fig. 3 sulfhydryl bearing agents can be covalently attached to this primary amino group via the formation of amide or amidine bonds. In case of the formation of amide bonds the carboxylic acid group of the ligands cysteine and thioglycolic acid reacts with the primary amino group of chitosan mediated by a water soluble carbodiimide². The formation of disulfide bonds by air oxidation during synthesis is avoided by performing the process at a pH below 5. At this pH-range the concentration of thiolate anions, representing the reactive form for oxidation of thiol groups, is low, and the formation of disulfide bonds can be almost excluded. Alternatively, the coupling reaction can be performed under inert conditions.

In the case of the formation of amidine bonds, 2-iminothiolane is used as a coupling reagent¹⁹. It offers the advantage of a simple one step coupling reaction. In addition, the thiol group of the reagent is protected against oxidation because of the chemical structure of the reagent. Orientating studies with all these thiolated chitosans showed that a degree of modification of 25–250 mmol thiol groups per gram chitosan leads to the highest improvement in the mucoadhesive and permeation enhancing properties. The amount of immobilized thiol groups in reduced and oxidized form can be determined via Ellman's reagent¹⁷ with and without previous quantitative reduction of disulfide bonds with borohydride²³.

PROPERTIES OF THIOLATED CHITOSANS

Mucoadhesive properties

The improved mucoadhesive properties of thiolated chitosans are explained by the formation of covalent bonds between thiol

groups of the polymer and cysteine rich subdomains of glycoproteins in the mucus layer²⁴. These covalent bonds are supposedly stronger than noncovalent bonds such as ionic interactions of chitosan with anionic substructures of the mucus layer. This theory was supported by the results of tensile studies with tablets of thiolated chitosans which demonstrated a positive correlation between the degree of modification with thiol bearing moieties and the adhesive properties of the polymer^{2,20}. These findings were confirmed by another *in vitro* mucoadhesion test system where the time of adhesion of tablets on intestinal mucosa was determined. The contact time of the thiolated chitosan derivatives increased with increasing amounts of immobilized thiol groups^{2,19}.

With chitosan-thioglycolic acid conjugates a 5–10-fold increase in mucoadhesion in comparison to unmodified chitosan was achieved. The mucoadhesive properties of chitosan-TBA (chitosan-4-thio-butyl-amidine) conjugates were even further improved. One explanation for this phenomenon can be given by the theory that chitosan-TBA conjugates additionally increased mucoadhesive properties due to improved ionic interactions between the additional cationic amidine substructure of the conjugate (see Fig 3) and anionic substructures within the mucus layer. Tensile studies with chitosan-TBA conjugates of low, medium and high molecular mass (150, 400 and 600 kDa) furthermore indicated that medium molecular mass thiolated chitosans displayed relatively, the highest mucoadhesiveness. Utilizing a medium molecular mass chitosan-TBA conjugate displaying 264 mM thiol groups per gram polymer led to a more than 100-fold improvement in mucoadhesion compared to unmodified chitosan. This represents the greatest progress made so far in the development of mucoadhesive polymers²⁰.

Permeation enhancing effect

In 1994 Illum et al showed the permeation enhancing capabilities of chitosan for the first time⁶. Chitosan is able to enhance the paracellular route of absorption, which is important for the transport of hydrophilic

compounds such as therapeutic peptides and antisense oligonucleotides across the membrane. Various studies carried out on Caco-2 cell monolayers demonstrated a significant decrease in the transepithelial electrical resistance after the addition of chitosan²⁵⁻²⁷. The mechanism underlying this permeation enhancing effect seems to be based on the positive charges of the polymer which interact with the cell membrane resulting in a structural reorganization of tight junction-associated proteins²⁸. In the presence of the mucus layer, however, this permeation enhancing effect is comparatively lower, as chitosan cannot reach the epithelium because of size limited diffusion and/or competitive charge interactions with mucins²⁹. Nevertheless, these results obtained on Caco-2 cell monolayers could be confirmed by *in vivo* studies, showing an enhanced intestinal absorption of the peptide drug, buserelin, in rats due to the co-administration of chitosan hydrochloride³⁰.

The permeation enhancing effect of chitosan can be greatly improved by the immobilization of thiol groups. The effect of thiolated chitosans has been shown in various permeation studies in Ussing type chambers using freshly excised intestinal mucosa¹⁸. The uptake of fluorescence-labeled bacitracin, for instance, was improved 1.6-fold utilizing 0.5% of chitosan-cysteine conjugate instead of unmodified chitosan¹⁸. In another study, the permeation enhancing effect of chitosan-TBA, in comparison to the permeation enhancing effect of unmodified chitosan, was shown. The uptake of the cationic marker compound, rhodamine-123 was 3-fold higher in the presence of thiolated chitosan than in unmodified chitosans²¹.

The likely mechanism responsible for this improved permeation enhancement has been attributed to the inhibition of the protein, tyrosine phosphatase. This enzyme seems to be involved in the opening and closing process of the tight junctions. Tyrosine phosphatase is responsible for the dephosphorylation of tyrosine subunits of occludin, representing an essential transmembrane protein of the tight junctions.

When these tyrosine subunits of occludin are dephosphorylated, the tight junctions are closed. In contrast, when these tyrosine subunits are phosphorylated, the tight junctions are opened. The inhibition of tyrosine phosphatase by compounds such as phenylarsine oxide, pervanadate or reduced glutathione leads consequently to phosphorylation and opening of the tight junctions³¹⁻³³. In contrast to the stable but toxic tyrosine phosphatase inhibitors phenylarsine oxide and pervanadate, the inhibitory effect of glutathione is lower as it is rapidly oxidized on the cell surface, losing its inhibitory activity³⁴. Due to the combination of reduced glutathione with thiolated chitosans, however, the oxidation of the inhibitor on the membrane can be restricted, as thiomers are capable of reducing oxidized glutathione³¹.

Thiolated chitosans as matrices for controlled drug release

Chitosan represents, primarily due to its mucoadhesive properties, a valuable tool for non-invasive drug delivery⁵. The longer residence time of formulations based on mucoadhesive polymers at the absorption site is believed to contribute to an increased absorption rate of the incorporated drug. However, such an enhanced bioavailability can be achieved only if a controlled release of the active agent out of the formulation is provided.

Thiolated chitosans also display, besides their strong mucoadhesive and permeation enhancing properties, excellent cohesive properties. The reduced thiol functions on the chitosan backbone enable thiolated chitosans not only to form disulfide bonds with mucus glycoproteins, but also to form inter- as well as intra-molecular disulfide bonds. Such a crosslinking of the polymeric chains²⁰ results in a high stability of drug carrier systems based on thiolated chitosans (Fig.4).

The cohesion and stability of a drug delivery system over the intended duration of drug liberation is often a substantial requirement for a controlled release. The usefulness of thiolated chitosans as carrier matrices for controlled drug release was demonstrated with model drugs, such as clotrimazole¹⁹⁻²²

and salmon calcitonin³⁶⁻³⁷. Clotrimazole is well-established as an antimycotic drug in the treatment of vaginal infections. In order to improve its therapeutic efficacy, a sustained release of the drug over a period of several days might be highly beneficial. The release of clotrimazole out of matrix tablets based on either chitosan-thioglycolic acid conjugate or chitosan-TBA conjugate was quantified. Both thiolated chitosan tablets remained stable during the whole period of the experiment (6 hours) and no disintegration could be observed. However, only the chitosan-TBA conjugate was able to guarantee a significant delay in drug release, compared to unmodified chitosan, leading to a sustained release over a much longer time period^{19,22}.

Furthermore, the release profile of salmon calcitonin out of matrix tablets based on the chitosan-TBA conjugate was determined. A pseudo zero order release profile of salmon calcitonin over the first 8 hours was observed in simulated intestinal fluid. During the experiment the tablets swelled continuously, maintaining good cohesiveness and releasing the active agent via a controlled diffusion process. These release studies, in which a peptide drug was liberated from a thiolated chitosan matrix system, permit information concerning the chemical events within the formulation to be gained. Strong unintended interactions between the polymeric matrix system and the peptide drug could be excluded, based on the according to this controlled and sustained release profile³⁷. Both studies confirm that, controlled drug release out of thiolated chitosan drug carrier systems can be achieved.

***In situ* gelling properties**

Rapid clearance from the site of drug action is one important factor that limits the efficacy of drugs administered to the ocular, nasal and vaginal mucosa. It is widely accepted that limiting the clearance by increasing the viscosity of a drug formulation will result in increased bioavailability of these drugs. A very promising strategy to obtain drug formulations of sufficient viscosity is based on *in situ* gel formation. The formation of a gel at the site of drug delivery combines the advantages of a

solution, which can be easily administered, with the favorable viscoelastic properties of a gel, providing a prolonged residence time of the formulation. The sol-gel transition occurs in the physiological environment as a result of physicochemical changes, such as changes in the pH³⁸, temperature^{38,39} or electrolyte concentration^{40,41}. Thiolated chitosans display *in situ* gelling properties due to the oxidation of thiol groups at physiological pH-values, which results in the formation of inter- and intramolecular disulfide bonds. This cross-linking process can be observed within the pH range of 5–6.8.

The *in situ* gelling behavior of thiolated chitosans was characterized *in vitro* by rheological measurements. The sol-gel transition of thiolated chitosans at pH 5.5 was completed after 2 hours when highly cross-linked gels were formed. In parallel, a significant decrease in the thiol group content of the polymers was observed, indicating the formation of disulfide bonds^{17,19}. The rheological properties of unmodified chitosan remained constant over the whole observation period. Rheological investigation of thiolated chitosans furthermore demonstrated a clear correlation between the total amount of polymer-linked thiol groups and the increase in elasticity of the formed gel. The more thiol groups were immobilized on chitosan, the higher was the increase in elastic modulus in solutions of thiolated chitosan^{17,19}.

Thiolated chitosan derivatives, therefore, seem to be promising new excipients for liquid or semisolid formulations, which should stabilize themselves once applied on the site of drug delivery. The *in situ* gel formation within the pH range of 5 to 6.8 makes the application of thiolated chitosans on vaginal, nasal and ocular mucosa plausible.

***IN VIVO* STUDIES: PROOF OF CONCEPT**

The potential of thiolated chitosans for the oral administration of hydrophilic macromolecules could meanwhile be shown by various *in vivo* studies^{35,36}. As model drug, for instance, salmon calcitonin was utilized, which is a peptide drug of cationic net charge and a molecular mass of 3.2 kDa. Salmon calcitonin

is used for the treatment of chronic bone diseases^{37,42}. It is currently marketed in nasal spray and injectable forms, both having the drawback of low patient acceptance. A higher patient compliance should be achieved by the application of an oral delivery system for this drug. However, the oral bioavailability thus far obtained is too low to permit therapeutic employment⁴³. Therefore, this peptide was regarded as a challenging model drug for testing the potential of thiolated chitosans.

Different drug carrier matrices, comprising chitosan-TBA conjugate as substantial polymeric excipient and containing equal amounts of salmon calcitonin and optionally the permeation mediator, reduced glutathione, were developed. In order to avoid an enzymatic degradation of the peptide drug in the gastrointestinal tract, chitosan-enzyme inhibitor conjugates were added. All compounds were homogenized and directly compressed to tablets. To enteric-coated tablets targeted to the small intestine, a chitosan-BBI (Bowman-Birk inhibitor) conjugate⁴⁴ and a chitosan-elastatinal conjugate⁴⁵ were added. Furthermore, an alternative strategy was evaluated, focusing on targeted drug release and absorption in the stomach. Tablets targeted to the stomach contained chitosan-pepstatin A conjugate³⁶ which should avoid pepsinic digestion of salmon calcitonin. In order to prevent mucoadhesion in the oral cavity and oesophagus, these tablets were coated with a triglyceride. The different tablets were orally given to rats and the plasma calcium level was monitored as a function of time. Pharmacological efficacy was calculated on the basis of the area under the reduction in plasma calcium levels of the oral matrix tablets versus intravenous injection. The main biofeedback parameters after application of the drug carrier matrices for the oral delivery of salmon calcitonin are shown in Table 1. *In vivo* studies showed no statistically significant ($P < 0.05$) reduction of the plasma calcium level caused by salmon calcitonin, which was orally given in solution. Furthermore, no significant effect was observed after oral administration of tablets comprising the peptide drug and

unmodified chitosan, although the native polymer is reported to be mucoadhesive and to exhibit permeation enhancing effect for hydrophilic macromolecules⁴⁶ (see Table 1).

Table 1 shows that the presence of the chitosan-TBA conjugate is essential for calcitonin absorption, since only tablets based on, thiolated chitosan caused a decrease in plasma calcium level of more than 5% for several hours. The increased absorption of the peptide, when embedded in a thiolated chitosan matrix, occurred due to the properties of the polymer derivative: the high stability and cohesiveness can provide a sustained release of the peptide⁴⁷, while the mucoadhesive features should lead to a prolonged residence time of the dosage form at the site of absorption. Moreover, the combination of thiolated chitosan with the permeation mediator, reduced glutathione, seems to have an impact on the bioresponse of orally given calcitonin. The significantly higher pharmacological efficacy of thiolated chitosan tablets containing glutathione in comparison to corresponding tablets without glutathione (see Table 1) indicates that glutathione contributes to the drug absorption process. These results are in good agreement with *in vitro* results demonstrating that thiomers show strong permeation enhancing effect, which can be further improved by the addition of glutathione³¹. Therefore, the high *in vivo* efficacy of thiolated chitosans can be additionally raised by the use of glutathione.

Among all thiolated chitosan formulations, stomach targeted tablets based on chitosan-TBA conjugate with the addition of both glutathione and chitosan-pepstatin A conjugate, showed the strongest effect. They led to a decrease of the plasma calcium level of more than 10% for at least 12 hours, thus demonstrating the validity of systemic peptide delivery via the stomach. Moreover, a faster and more reproducible onset of action was obtained by this novel approach^{35,36}. According to these results, the applicability of thiolated chitosans for the oral administration of other peptide drugs seems also likely and is the subject of ongoing studies.

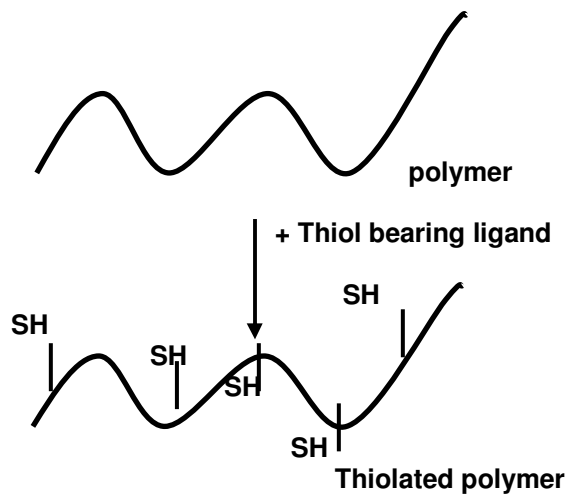


Fig 1: Simple diagrammatic representation for preparing thiolated polymers

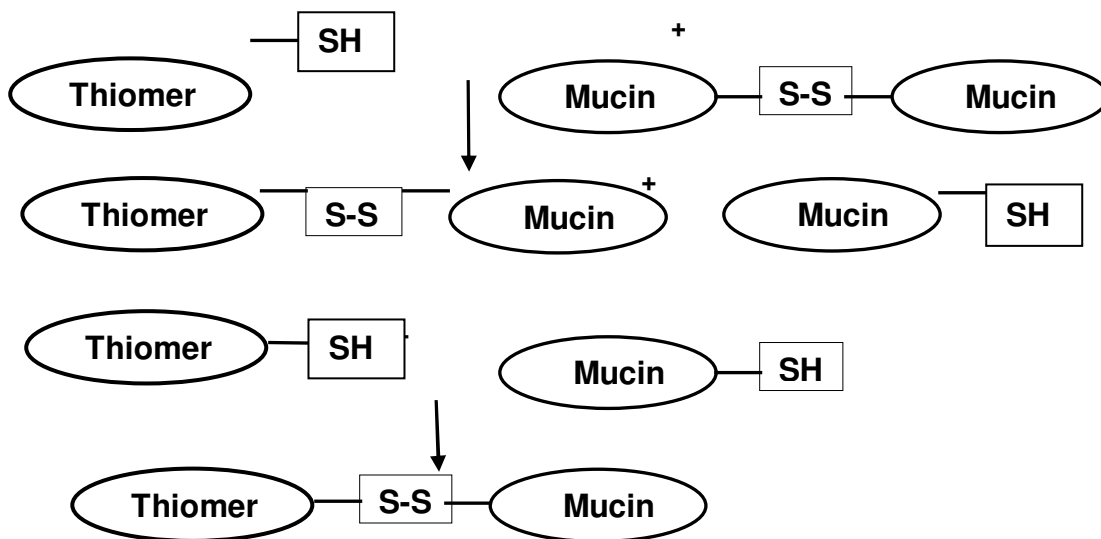


Fig 2: Mechanism of disulfide bond formation between thiomers and mucus glycoproteins (mucins)

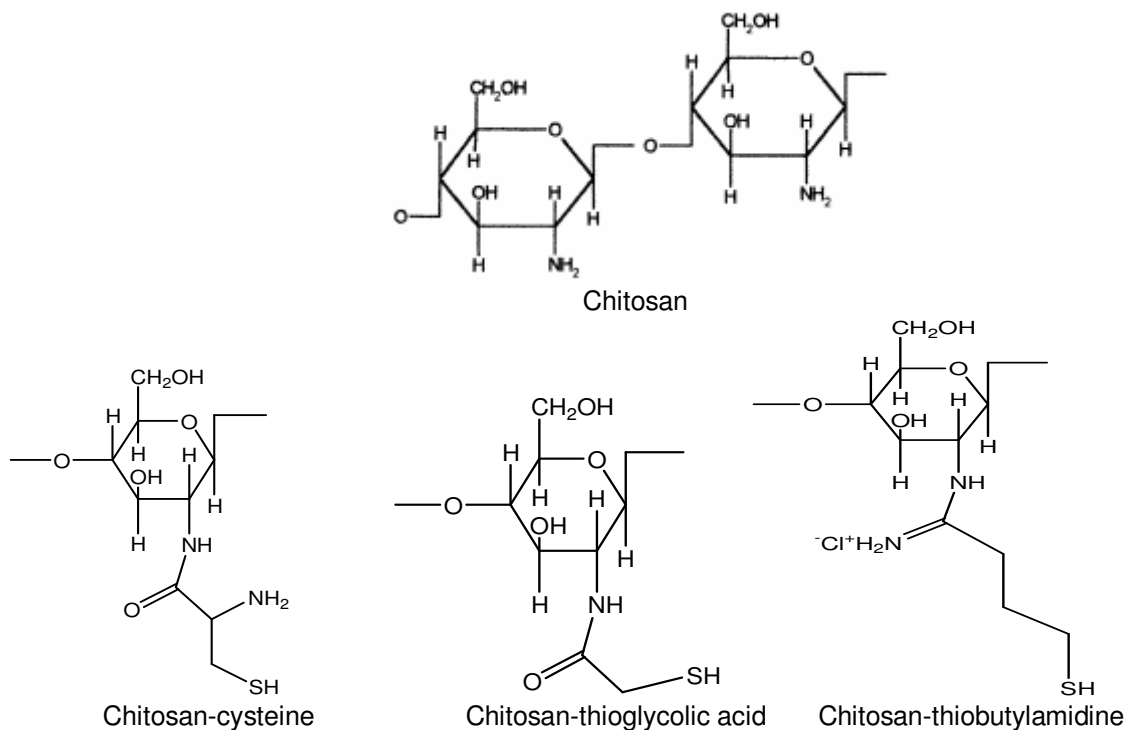


Fig 3: Structures of chitosan and thiolated chitosans

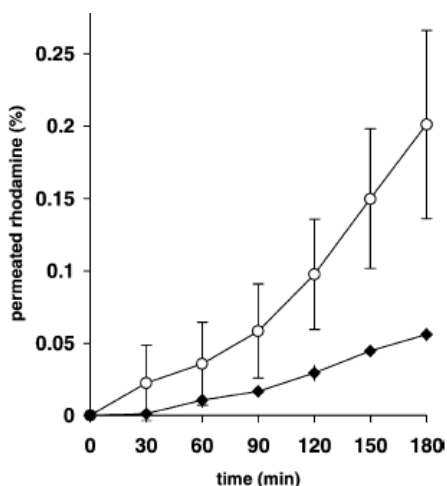


Fig 4: Permeation enhancing effect of 0.5% (m/v) Chitosan-TBA conjugate with 5% (m/v) glutathione (o) and 0.5% (m/v) unmodified chitosan (■) on small intestinal mucosa²¹

FUTURE TRENDS

Non-invasive peptide delivery

The incorporation of peptide drugs exhibiting a cationic net charge in anionic mucoadhesive

polymers on the one hand leads to a strong reduction in the mucoadhesive properties and, on the other hand, may hinder drug release as a result of strong ionic interactions between

Table 1: Main biofeedback parameters after oral administration of tablets containing all equal amounts of Salmon calcitonin to rats (n=5)

Tablet composition	Maximal reduction of Ca-level (% of initial value)	Time point of maximal reduction of Ca-level (hour)	Pharmacological
Small Intestine targeted tablets (2 mg)	89.9		1.3
Chitosan-TBA conjugate,		12	
Citosan-BBI conjugate,	91.0		0.9
Chitosan-elastatinal conjugate, glutathione.	a	12	0
Chitosan-TBA conjugate,		-	
Citosan-BBI conjugate,			
Chitosan-elastatinal conjugate.			
Unmodified chitosan.			
Stomach targeted tablets (5 mg)	88.8	6	1.3
Chitosan-TBA conjugate,			
glutathione	a	-	0
Chitosan-pepstatin A conjugate.			
Unmodified chitosan.			

The share of the basic excipient (written in italic and bold) in each tablet type was of at least 65%. (Adapted from Andreas BS et al.³⁵ and Guggi et al.³⁶) ^aDecrease in plasma calcium level was not significant in comparison to physiological daily variations in rats.

the therapeutic ingredient and the polymeric network. Consequently, cationic therapeutic peptides or peptidomimetics such as calcitonin or desmopressin need to be embedded in cationic or non-ionic mucoadhesive polymers. As non-ionic polymers cannot provide sufficient high mucoadhesion and thiolated chitosans display comparatively the highest mucoadhesive properties among cationic polymers, this type of thiomers seems to be a favorable tool for the oral administration of cationic hydrophilic macromolecules. Apart from oral delivery systems thiolated chitosans seem to be useful also for other non-invasive routes of peptide drug administration. In particular, the nasal, vaginal, buccal and ocular mucosae are interesting targets.

Production of micro- and nano-particles

Microparticles based on chitosan disintegrate very rapidly unless they are combined with multivalent anionic compounds such as

sodium sulfate⁴⁸ or alginate leading to stabilization by an ionic cross-linking process. Due to the addition of such multivalent anionic compounds, however, the mucoadhesive properties of chitosan are strongly reduced. In contrast, microparticles that are based on thiolated chitosan do not disintegrate. Because of the formation of disulfide bonds within the polymeric network, microparticles are strongly stabilized⁴⁹. Consequently, a controlled drug release out of thiolated chitosan microparticles can be provided. In contrast to the addition of multivalent anionic compounds, the immobilization of thiol groups on chitosan leads to strongly improved mucoadhesive properties.

Tissue engineering

A further interesting application of thiolated chitosans is their use in tissue engineering. The expanding field of tissue engineering applications has accelerated the need for materials which are tissue compatible,

biodegradable and with mechanical properties similar to the target tissues. Biodegradable and biocompatible polymers have been attractive candidates for scaffolding materials because they degrade as the new tissues are formed, eventually without inflammatory reactions or toxic degradation. Recently, Kast et al demonstrated the biodegradability of thiolated chitosan, paving the way for its use as novel scaffold material⁵⁰. Further, studies in this direction were performed with L-929 mouse fibroblasts seeded onto chitosan-thioglycolic acid sheets. The results of this study showed that thiolated chitosan can provide a porous scaffold structure guaranteeing cell anchorage, proliferation and tissue formation in three dimensions⁵⁰. Due to the *in situ* gelling properties, it seems possible to provide a certain shape of the scaffold material by pouring a liquid thiolated chitosan⁵¹ cell suspension in a mold. Furthermore, liquid polymer cell suspensions may be applied by injection forming semi-solid scaffolds at the site of tissue damage. Since low concentrated aqueous solutions of thiolated chitosan remain liquid when stored under inert conditions and are rapidly gel under access of oxygen, they seem to be promising candidates for such applications.

Coating of stents

Another promising application of thiolated chitosans is their use as coating material for stents. Polymer-coated drug-eluting stents are a potential technique to achieve high local tissue concentrations of an effective drug at the precise site and at the time of vessel injury. First orientating studies demonstrated that by simply dipping the stent in a thiolated chitosan solution and drying it on air, a stable coating could be achieved. During the drying process, cross-linking of chitosan by the formation of disulfide bonds due to air oxidation, takes place. The polymeric network is thereby stabilized on the stent. The chitosan coating should allow sustained release of incorporated drugs such as anti-inflammatory agents or agents avoiding cell proliferation. Recently, it was shown that stents can be successfully coated with thiolated poly(acrylic acid) and that sustained release of a model

peptide drug out of this thiomeric coating can be provided. Similar results can be expected for thiolated chitosans but have to be verified by ongoing studies.

CONCLUSION

The chemical modification of chitosan via derivatization with various reagents bearing sulfhydryl functions causes a dramatic change in the polymer's properties. Mucoadhesiveness and cohesiveness are strongly improved. A comparatively stronger permeation enhancing effect is provided which can be further raised by the combination of thiolated chitosans with the permeation mediator, glutathione. Furthermore, thiolated chitosans display *in situ* gelling features and facilitate controlled drug release. Due to these advantageous features thiolated chitosans have been successfully used for peroral administration of peptide drugs. They seem to represent a promising new generation of polymeric excipients, in particular for the non-invasive administration of hydrophilic macromolecular drugs.

REFERENCES

1. Lehr CM. From sticky stuff to sweet receptors-achievements, limits and novel approaches to bioadhesion. *Eur. J. Drug Metab. Pharm.* 1996; 21: 139-48.
2. Kast CE, Andreas BS. Thiolated polymers – thiomers: development and *in vitro* evaluation of chitosan-thioglycolic acid conjugates. *Biomaterials* 2001; 22: 2345–2352.
3. Felt O, Buri P, Gurny R. Chitosan: a unique polysaccharide for drug delivery. *Drug Dev. Ind. Pharm.* 1998; 24: 979–993.
4. Fini A, Orienti I. The role of chitosan in drug delivery. *Am. J. Drug Deliv.* 2003; 1: 43–59.
5. Takeuchi H, Yamamoto H, Kawashima Y. Mucoadhesive nanoparticulate systems for peptide drug delivery. *Adv. Drug Deliv. Rev.* 2001; 47: 39–54.
6. Illum L, Farraj NF, Davis SS. Chitosan as a novel nasal delivery system for peptide drugs. *Pharm. Res.* 1994; 11: 1186–1189.
7. Felt O, Furrer P, Mayer JM, Plazonnet B, Buri P, Gurny R. Topical use of chitosan in ophthalmology: tolerance assessment and evaluation of precorneal retention. *Int. J. Pharm.* 1999; 180: 185–193.
8. Senel S, Kremer M, Kas S, Wertz PW, Hincal AA, Squier CA. Enhancing effect of chitosan on peptide drug delivery across buccal mucosa. *Biomaterials* 2000; 21: 2067–2071.

9. Borchard G. Chitosans for gene delivery. *Adv. Drug Deliv. Rev.* 2001; 52: 145–150.
10. Liu WC, Yao KD. Chitosan and its derivatives – a promising non-viral vector for gene transfection. *J. Control. Rel.* 2002; 83: 1–11.
11. Snyder GH, Reddy MK, Cennerazzo MJ, Field D. Use of local electrostatic environments of cysteines to enhance formation of a desired species in a reversible disulfide exchange reaction. *Biochim. Biophys. Acta* 1983; 749: 219–226.
12. Thanou M, Florea BI, Langemeyer MW, Verhoef JC, Junginger HE. N-trimethylated chitosan chloride (TMC) improves the intestinal permeation of the peptide drug busserelin in vitro (Caco-2 cells) and in vivo (rats). *Pharm. Res.* 2000; 17: 27–31.
13. Thanou M, Nihot MT, Jansen M, Verhoef JC, Junginger JC. Mono-N-carboxymethyl chitosan (MCC), a polyampholytic chitosan derivative, enhances the intestinal absorption of low molecular weight heparin across intestinal epithelia in vitro and in vivo. *J. Pharm. Sci.* 2001; 90: 38–46.
14. Baumann H, Faust V. Concepts for improved regioselective placement of O-sulfo, N-sulfo, N-acetyl, and N-carboxymethyl groups in chitosan derivatives. *Carbohydr. Res.* 2001; 331: 43–57.
15. Andreas BS, Krajčec ME. Mucoadhesive polymers as platforms for peroral peptide delivery and absorption: synthesis and evaluation of different chitosan-EDTA conjugates. *J. Control. Rel.* 1998; 50: 215–223.
16. Andreas BS, Hopf TE. Synthesis and in vitro evaluation of chitosan-thioglycolic acid conjugates. *Sci. Pharm.* 2001; 69: 109–118.
17. Hornof MD, Kast CE, Andreas BS. In vitro evaluation of the viscoelastic behavior of chitosan – thioglycolic acid conjugates. *Eur. J. Pharm. Biopharm.* 2003; 55: 185–190.
18. Andreas BS, Brandt UM, Clausen AE. Synthesis and in vitro evaluation of chitosan-cysteine conjugates. *Sci. Pharm.* 1999; 67: 196–208.
19. Andreas BS, Hornof M, Zoidl T. Thiolated polymers – thiomers: modification of chitosan with 2-aminothiolane. *Int. J. Pharm.* 2003; 260: 229–237.
20. Roldo M, Hornof M, Caliceti P, Andreas BS. Mucoadhesive thiolated chitosans as platforms for oral controlled drug delivery: synthesis and in vitro evaluation. *Eur. J. Pharm. Biopharm.* 2004; 57(1): 115–21.
21. Langoth N, Guggi D, Pinter Y, Andreas BS. Thiolated Chitosan: In Vitro Evaluation of its Permeation Enhancing Properties. *J. Control. Rel.* 2004; 94(1): 177–86.
22. Kast CE, Valenta C, Leopold M, Andreas BS. Design and in vitro evaluation of a novel bioadhesive vaginal drug delivery system for clotrimazole. *J. Control. Rel.* 2002; 81: 347–354.
23. Leitner VM, Marschutz MK, Andreas BS. Mucoadhesive and cohesive properties of poly(acrylic acid)-cysteine conjugates with regard to their molecular mass. *Eur. J. Pharm. Sci.* 2003; 18: 89–96.
24. Leitner VM, Walker GF, Andreas BS. Thiolated polymers: evidence for the formation of disulphide bonds with mucus glycoproteins. *Eur. J. Pharm. Biopharm.* 2003; 56: 207–214.
25. Artursson P, Lindmark T, Davis SS, Illum L. Effect of chitosan on the permeability of monolayers of intestinal epithelial cells (Caco-2). *Pharm. Res.* 1994; 11: 1358–1361.
26. Borchard G, Luessen HL, Boer AGD, Verhoef JC, Lehr CM, Junginger HE. The potential of mucoadhesive polymers in enhancing intestinal peptide drug absorption. III: Effects of chitosanglutamate and carbomer on epithelial tight junctions in vitro. *J. Control. Rel.* 1996; 39: 131–138.
27. Dodane V, Amin Khan M, Merwin JR. Effect of chitosan on epithelial permeability and structure. *Int. J. Pharm.* 1999; 182: 21–32.
28. Schipper NGM, Olsson S, Hoogstraate JA, Boer AGD, Varum KM, Artursson P. Chitosans as absorption enhancers for poorly absorbable drugs. 2: Mechanism of absorption enhancement. *Pharm. Res.* 1997; 14: 923–929.
29. Schipper NGM, Varum KM, Stenberg P, Ocklind G, Lennernas H, Artursson P. Chitosans as absorption enhancers for poorly absorbable drugs. 3: Influence of mucus on absorption enhancement. *Eur. J. Pharm. Sci.* 1999; 8: 335–343.
30. Luessen HL, Leeuw BJD, Langemeyer MW, Boer AGD, Verhoef JC, Junginger HE. Mucoadhesive polymers in peroral peptide drug delivery. VI. Carbomer and chitosan improve the intestinal absorption of the peptide drug busserelin in vivo. *Pharm. Res.* 1996; 13: 1668–1672.
31. Clausen AE, Kast CE, Andreas BS. The role of glutathione in the permeation enhancing effect of thiolated polymers. *Pharm. Res.* 2002; 19: 602–608.
32. Barrett WC, Gnore JPD, Konig S, Fales HM, Keng YF, Zhang ZY. Regulation of PTP1B via glutathionylation of the active site cysteine 215. *Biochemistry* 1999; 38: 6699–6705.
33. Staddon JM, Herrenknecht K, Smales C, Rubin LL. Evidence that tyrosine phosphorylation may increase tight junction permeability. *J. Cell Sci.* 1995; 108: 609–619.
34. Grafstrom R, Stead AH, Orrenius S. Metabolism of extracellular glutathione in rat small-intestinal mucosa. *Eur. J. Biochem.* 1980; 106: 571–577.
35. Andreas BS, Kast CE, Guggi D. Permeation enhancing polymers in oral delivery of hydrophilic macromolecules: thioimer/ GSH systems. *J. Control. Rel.* 2003; 93(2): 95–103.
36. Guggi D, Krauland AH, Andreas BS. Systemic peptide delivery via the stomach: in vivo evaluation of an oral dosage form for salmon calcitonin. *J. Control. Rel.* 2003; 92: 125–135.

37. Torres-Lugo M, Peppas NA. Transmucosal delivery systems for calcitonin: a review. *Biomaterials* 2001; 21: 1191–1196.
38. Edsman K, Carlfors J, Petersson J. Rheological evaluation of poloxamer as an situ gel for ophthalmic use. *Eur. J. Pharm. Sci.* 1998; 6: 105–112.
39. Bromberg LE. Enhanced nasal retention of hydrophobically modified polyelectrolytes. *J. Pharm. Pharmacol.* 2001; 53: 109–114.
40. Deasy PB, Quigley KJ. Rheological evaluation of deacetylated gellan gum (Gelrite) for pharmaceutical use. *Int. J. Pharm.* 1991; 73: 117–123.
41. Paulsson M, Hagerstrom H, Edsman K. Rheological studies of the gelation of deacetylated gellan gum (Gelrite) in physiological conditions. *Eur. J. Pharm. Sci.* 1999; 9: 99–105.
42. Lee YH, Sinko PJ. Oral delivery of salmon calcitonin. *Adv. Drug Deliv. Rev.* 2000; 42: 225–238.
43. Shah RB, Ahsan F, Khan MA. Oral delivery of proteins: progress and prognostication. *Crit. Rev. Ther. Drug Carrier Syst.* 2002; 19: 135–169.
44. Guggi D, Andreas BS. In vitro evaluation of polymeric excipients protecting calcitonin against degradation by intestinal serine proteases. *Int. J. Pharm.* 2003; 252: 187–196.
45. Andreas BS, Scerbe-Saiko A. Synthesis and in vitro evaluation of chitosan-EDTA-protease-inhibitor conjugates which might be useful in oral delivery of peptides and proteins. *Pharm. Res.* 1998; 15: 263–269.
46. Andreas BS. Chitosan and its derivatives: potential excipients for peroral peptide delivery systems. *Int. J. Pharm.* 2000; 194: 1–13.
47. Guggi D, Kast CE, Andreas BS. In vivo evaluation of an oral calcitonin delivery system for rats based on a thiolated chitosan matrix. *Pharm. Res.* 2003; 20(12): 1989–1994.
48. Lubben IMV, Verhoef JC, Aelst ACV, Borchard G, Junginger HE. Chitosan microparticles for oral vaccination: preparation, characterization and preliminary in vivo uptake studies in murine Peyer's patches. *Biomaterials* 2001; 22: 687–694.
49. Coppi G, Iannuccelli V, Leo E, Bernabei MT, Cameroni R. Chitosan-alginate microparticles as a protein carrier. *Drug Dev. Ind. Pharm.* 2001; 27: 393–400.
50. Ma PX, Choi JW. Biodegradable polymer scaffolds with well defined interconnected spherical pore network. *Tissue Eng.* 2001; 7: 23–33.
51. Kast CE, Frick W, Losert U, Andreas BS. Chitosanthioglycolic acid conjugate: a new scaffold material for tissue engineering?. *Int. J. Pharm.* 2003; 256: 183–189.