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Original Research Article

Etanercept Inhibits Pro-inflammatory Cytokines Expression in Titanium Particle-Stimulated Peritoneal Macrophages

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Abstract

Purpose: To investigate the inhibitory role of Etanercept in pro-inflammatory cytokines such as TNF- α , *IL*-1 β and *IL*-6 production in titanium (Ti) particle stimulated macrophages.

Methods: Peritoneal macrophages were stimulated with 1×10^9 Ti particles and treated simultaneously with or without 10, 100, or 1000 ng/mL Etanercept. The levels of TNF- α , IL-1 β and IL-6 in the culture supernatants were measured using ELISA.

Results: Titanium particles could stimulate TNF- α , IL-1 β and IL-6 secretion in peritoneal macrophages. Etanercept inhibited Ti particle-induced TNF- α release by 29.7 % at 10 ng/ml (19.19 ± 4.72 pg/mL, p < 0.01), 49.3 % at 100 ng/mL (13.83 ± 3.72 pg/ml, p < 0.01) and 60.4 % at 1000 ng/mL (10.82 ± 3.87 pg/mL, p < 0.001), IL-1 β release by 5.23 % at 10 ng/mL (34.79 ± 7.83 pg/mL, p > 0.05), 21.06 % at 100 ng/mL (28.98 ± 4.81 pg/mL, p < 0.01) and 29.83 % at 1000 ng/mL (25.76 ± 5.23 pg/ml, p < 0.001), and IL-6 release by 38.69 % at 10 ng/mL (25.68 ± 99.56 pg/mL, p < 0.01), by 42.13 % at 100 ng/mL (242.4 ± 33.26 pg/mL, p < 0.01) and 53.4 % at 1000 ng/ml (195.2 ± 48.82 pg/mL, p < 0.001).

Conclusion: Etanercept has potent ability to prevent wear debris-induced osteolysis and may be valuable as a therapeutic agent for the treatment of prosthetic loosening in humans.

Keywords: Etanercept; titanium particle; proinflammatory cytokines; peritoneal macrophages

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INTRODUCTION

Total joint arthroplasties is a widely used final treatment option in many cases to reduce pain, restore joint function and allow patients to return to normal living [1]. Wear debris, which is generated from all the components of the prosthetic joint surface, is widely recognized as the major cause of aseptic loosening [2,3]. Wear debris are phagocytosed by macrophages, resulting in the secretion of proinflammatory cytokines, such as tumor necrosis factor- α (TNF-

α), interleukin-1β (IL-1β) and IL-6 [4-6]. Among them, TNF-α is the pivotal cytokine involved in wear debris-induced osteolysis. Blockage of TNF-α activity inhibits both osteoclast differentiation and osteolysis induced by wear particles in the murine calvaria model [7,8]. Thus, TNF-α has been proved to be potential treatment target for wear debris induced osteolysis.

Etanercept, which is a soluble fusion protein composed of the extracellular domains of type 2 TNF receptor (p75) fused to the Fc region of a IgG1, is a TNF- α antagonist that has been approved by U.S. F.D.A. for the treatment of rheumatoid, juvenile rheumatoid and psoriatic arthritis [9,10]. Recent clinical data have indicated that Etanercept also has beneficial effects on preventing bone erosions in rheumatoid arthritis and psoriasis [11,12]. Given the emerging evidence that TNF- α plays a similar and central pathogenetic role in prosthetic loosening and rheumatoid arthritis, we proposed that Etanercept has the ability to inhibit wear debris induced osteolysis. The objective of current study was to determine if Etanercept was capable of inhibiting pro-inflammatory cytokines such as TNF- α , IL-1 β and IL-6 production in titanium (Ti) particle stimulated macrophages.

EXPERIMENTAL

Animals and reagents

BALB/c mice (6 - 8 weeks old) were purchased from laboratory animal center of Ningxia medical college (Ningxia, China) and maintained in a temperature controlled room with 12 h light /dark cycles with regular chow and water. The experimental procedures were approved by the Laboratory Animal Care Committee at Ningxia medical college. All animals received humane care according to the criteria outlined in the "Guide for the Care and Use of Laboratory Animals" prepared by the National Academy of Sciences and published by the National Institutes of Health (NIH publication 86 - 23 revised 1985). Etanercept (trade name Enbrel) was purchased from Amgen & Wyeth (Philadelphia, PA, USA). Titanium particles (Ti6Al4V alloy) were a gift from Zimmer (Warsaw, IN, USA). Mouse tumor necrosis factor- α (TNF- α), interleukin -1 β (IL-1 β) and interleukin-6 (IL-6) ELISA kit were from R&D Systems (Minneapolis, MN, USA). RPMI-1640, L-glutamine, penicillin, streptomycin, and fetal bovine serum (FBS) were from Gibco (Grand Island, NY, USA). Limulus assay kit was from Horseshoe Crab Reagent Manufactory (Xiamen, China).

Mouse peritoneal macrophage culture

Resident peritoneal macrophages were collected from mice as previously described [13]. Briefly, peritoneal macrophages were recovered by peritoneal lavage with 5 ml of cold RPMI-1640 medium and isolated by centrifugation (1000 rpm/min, 10 min). Cells (2×10^5) were plated on 24-well tissue culture plates and cultured in RPMI 1640 medium containing 10 % FBS, penicillin (100 U/ml) and streptomycin (100 U/mL) at 37 °C, 5 % CO₂ for 12 h. The no adherent cells were removed, and the adherent cells were washed twice with RPMI-1640 medium and used in experiments.

Preparation of Ti particles

Commercially pure Titanium (Ti) particles (Zimmer, Warsaw, USA) were sterilized by incubation in 25 % nitric acid at 70 °C for 1 h and then suspended in phosphate-buffered saline (PBS) at a concentration of 1×10^9 particles /ml [8]. Particle size was confirmed with a Coulter Channelizer (Beckman Coulter, Fullerton, CA, USA), which determined 95 % of the particles to be < 5 µm in diameter.

Limulus amebocyte lysate assay

For determination of endotoxin concentration in Ti particle suspension, limulus amebocyte lysate (LAL) assays were performed according to the manufacturer's protocol. Briefly, Ti particles suspension serum samples were diluted 10 % (v/v) in LAL reagent water, and heated to 70 °C for 5 min to remove any nonspecific inhibition to the assay. Samples were incubated with equal volumes of LAL for 10 min at 37 °C and developed with equal volumes of substrate solution for 6 min. The absorbance of the assay plate was read at 405 nm using a micro plate reader (BioTek, Winooski, VT). Samples and standards were run in duplicate, and the detection limit was 0.15 EU/mL.

ELISA

Peritoneal macrophage at 1×10^5 cells/well were stimulated with 1×10^9 Ti particles and treated simultaneously with or without 10, 100, or 1000 ng/mL Etanercept. After 18 h, culture supernatants were collected and centrifuged at 1500 rpm for 5 min to remove the Ti particles and cells, the supernatants were collected, and the levels of TNF- α , IL-1 β and IL-6 in the culture were measured using ELISA kits (R&D Systems, Minneapolis, MN, USA) according to the manufacturer's instructions.

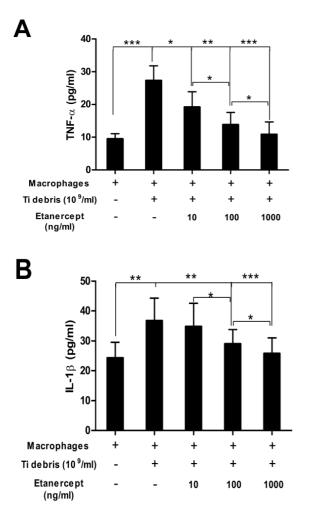
Statistics

All data were analyzed by Student's t test and were expressed as means \pm SEM, differences were considered statistically significant when *p* < 0.05. The significance levels are marked **p* < 0.05; ** *p* < 0.01; *** *p* < 0.001.

RESULTS

It has been shown that particulate debris could stimulate the production of pro-inflammatory

cytokines such as TNF- α , IL-1 β and IL-6 in macrophages [14,15]. We first determined the effect of Ti particles that induces TNF-a, IL-1β release in resident peritoneal and IL-6 macrophages. Limulus assay has been performed and data showed that the Ti particle suspension was free of endotoxin (< 0.25 EU/mL). 1 × 10^9 Ti particles were then used to treat macrophages and TNF-α, IL-1β and IL-6 production were measured by ELISA. Data showed that TNF- α (9.48 ± 1.58 pg/mL vs. 27.3 ± 4.47 pg /mL, p < 0.001), IL-1 β (24.25 ± 5.25 pg/mL vs. 36.71 ± 7.57 pg/mL, p < 0.01) and IL-6 (122.3 ± 37.78 pg/mL vs. 418.9 ± 31.98 pg/mL, p < 0.001) production was substantially induced by Ti particles (Fig. 1 A-C). To evaluate the inhibitory effect of Etanercept on Ti particleinduced TNF- α , IL-1 β and IL-6 release, peritoneal macrophages were treated with or without Etanercept (10, 100, or 1000 ng/mL) for 18 h. Data showed that Etanercept significantly inhibited Ti particle-induced TNF- α release by 29.7 % at 10 ng/mL (19.19 ± 4.72 pg/ml, p < 0.01), 49.3 % at 100 ng/mL.



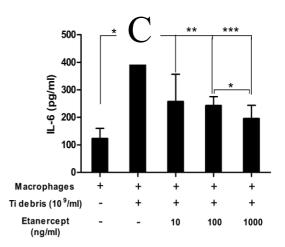


Figure 1: Inhibitory effect of Etanercept on titanium particle-induced TNF- α , IL-1 β and IL-6 release by macrophages. Peritoneal macrophage (1 × 10⁵ cells/well) were stimulated with 1 × 10⁹ Ti particles and treated simultaneously with or without 10, 100, or 1000 ng/mL Etanercept. After 18 h, culture supernatants were collected and centrifuged at 1500 rpm for 5 min to remove the Ti particles and cells, the supernatants were collected, and the levels of TNF- α (A), IL-1 β (B) and IL-6 (C) in the culture were measured using ELISA kits. The data shown are representatives of three independent experiments. Data are shown as mean ± SEM (*p < 0.05, **p < 0.01, ***p < 0.001)

 $(13.83 \pm 3.72 \text{ pg/mL}, p < 0.01)$ and 60.4 % at 1000 ng/mL (10.82 \pm 3.87 pg/mL, p < 0.001) compared to the Ti particles stimulated control $(27.3 \pm 4.47 \text{ pg/mL})$ (Fig.1 A), and significantly inhibited Ti particle-induced IL-1ß release by 5.23 % at 10 ng/mL (34.79 ± 7.83 pg/mL, p > 0.05), 21.06 % at 100 ng/mL (28.98 ± 4.81 pg/mL, p < 0.01) and 29.83 % at 1000 ng/ml $(25.76 \pm 5.23 \text{ pg/mL}, p < 0.001)$ compared to the Ti particles stimulated control (36.71 ± 7.57 pg/mL) (Fig. 1B). Etanercept could also significantly inhibit Ti particle-induced IL-6 release by 38.69 % at 10 ng/mL (256.8 ± 99.56 pg/mL, p < 0.01), 42.13 % at 100 ng/mL (242.4 ± 33.26 pg/mL, p < 0.01) and 53.4 % at 1000 ng/mL (195.2 ± 48.82 pg/mL, p < 0.001) compared to the Ti particles stimulated control (418.9 ± 31.98 pg/mL) (Fig. 1C). These results demonstrate that Etanercept has a dosedependent inhibitory effect on Ti particle-induced pro-inflammatory cytokines such as TNF-a, IL-1β and IL-6 production in macrophages.

DISCUSSION

The major finding of this study is that Etanercept, a TNF- α antagonist, could dose-dependently inhibit the production of TNF- α , IL-1 β and IL-6 by

macrophages stimulated with Ti particles. TNF-a may be a dominant upstream event that has dramatic consequences on periprosthetic osteolysis, because blockade of TNF-a also reduces the secretion of IL-1ß and IL-6, TNF-a acts as an autocrine/paracrine stimulator of additional TNF- $\!\alpha$ release. These results are consistent with our hypothesis that Etanercept inhibit the pro-inflammatory cytokine can stimulation induced by cascade Ti of macrophages. Similar results were recently published by other investigators examining the effects of Etanercept on wear debris induced osteolysis using macrophage cell lines [16]. Combined with previously reports, It suggests that TNF- α is critically involved in osteolysis and aseptic loosening and that soluble TNF- α inhibitors, such as Etanercept, may be useful as therapeutic agents for the treatment of prosthetic loosening.

TNF- α expression has been studied extensively in response to various stimuli. In this study, we used Ti particles to stimulate a pro-inflammatory response in macrophage cultures *in vitro*. Ti particles are readily available and the majority of the particles are small enough to be phagocytosed by macrophages (< 10 µm in diameter) [17]. Meanwhile, numerous literatures have shown that Ti particles induce the same reaction as other types of implant wear debris [18].

Macrophages play a crucial role in inflammatory bone disorder such as wear debris-induced aseptic loosening and are major sources of inflammatory mediators. Here we chose resident peritoneal macrophages, which have been widely used as a macrophage source in mice as the exposed cells. Previous studies of macrophage physiology customarily used macrophage cell lines such as RAW264.7 and J774 cells [19]. These cells, derived from pristane-elicited murine peritoneal macrophages transformed with Abelson leukemia virus, have been particularly valuable because of their ease of culture, rapid growth rate, and phenotypic resemblance to primary macrophages. However, these cell lines cannot fully mimic the phenotype of primary macrophages in vivo and the uptake efficiency of Ti particles by the peritoneal macrophages maybe higher than that of RAW264.7 cells.

Although we have shown that Etanercept is able to prevent pro-inflammatory cytokines production by macrophages stimulated with wear debris, the precise mechanism by which Etanercept is involved in wear debris induced osteolysis remains unknown. As the first pro-inflammatory cytokine produced by macrophages in response to particles, TNF- α has been shown to control the release of other pro-inflammatory mediators such IL-1ß and IL-6. One primary mechanism may be that, as soluble receptor of TNF, Etanercept can competitively bind to the TNF membrane receptor (mTNF-R) on the cell surface and block all signaling through the membrane bound TNF receptors, thus inhibiting osteoclastogenesis and osteoclast activity. Another important mechanism by which TNF-a contributes to the pathogenesis of aseptic loosening is its inhibitory effects on osteoblasts. Etanercept may inhibit osteoclastogenesis by down regulating the expression of osteoprotegerin ligand and M-CSF, IL-6 and PEG2 in osteoblastic cells [20].

CONCLUSION

In summary, the present study suggests that Etanercept inhibits Ti particle-induced TNF- α , IL-1 β and IL-6 release by macrophages, and therefore, may be a potential therapeutic agent for the prevention and/or treatment of osteolysis and loosening after TJA. Further investigations are required to examine the inhibitory effects of Etanercept on wear debris induced osteolysis *in vivo* and the underlying molecular mechanism.

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