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**Original Research Article** 

# Expression profiling of miR-96, miR-584 and miR-422a in colon cancer and their potential involvement in colon cancer pathogenesis

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### Abstract

**Purpose:** To determine the correlation between miRNAs; miR-96, miR-422a and miR584, and colon cancer, and also to test whether any of these miRNAs can act as non-invasive biomarkers in colon cancer

**Methods:** The tumor samples and the corresponding normal mucosa used in this study were collected from 60 patients diagnosed with colon cancer. HT-29, HCT-116 and SW-620 cell lines were used for miRNA expression profiling. Total RNA was extracted using kit-based methods with minor modifications, and specific miRNAs were detected and quantified by quantitative real-time polymerase chain reaction [RT-PCR].

**Results:** The results indicate down-regulation of miR-96, miR-584 and miR-422a in colon cancer tissue and decreased expression levels in the three colon cancer cell lines (all p < 0.01). Lower miRNA expression levels are correlated with increased tumor size.

**Conclusion:** This study shows that miR-96, miR-584 and miR-422a are down-regulated in colon cancer and are associated with tumor size. Thus, the ratio of miR-96/miR-638 in plasma is a potential non-invasive biomarker in colon cancer.

Keywords: Colon cancer, miR-96, miR-422a, miR-584, Expression profiling, Biomarker

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### INTRODUCTION

MicroRNAs (miRNAs) are relatively newly discovered class of molecules that play crucial roles in cellular processes in humans and other mammals. miRNAs are transcribed by RNA polymerase II and do not code for any proteins, and are of varying lengths ranging between 20 and 25 nucleotides [1]. Previously unknown, miRNAs have been shown to play significant roles in cancer related pathways such as invasion, migration, proliferation, apoptosis and other aspects of tumorigenesis [2,3]. The precise mechanisms of miRNA function in mammals are still debatable. However, several years of literary evidence suggest that miRNAs regulate gene expression through precise sequence specific base pairing at the 3' UTR of the target mRNA or at the translational level [4].

Colon cancer is the cancer of the colon with a million new cases reported each year globally [5]. In the United States alone, colon cancer is the third most prevalent form of cancer with 95,000

cases reported each year, and second most leading cause of cancer related deaths. Similar to other types of cancer, survival rates of colon cancer patients depend on the early detection of colon cancer. Therefore, it is important to detect colon cancer at an early stage. However, only 40 % of colon cancers are detected at an early stage. Several studies have shown that early detection of colon cancer is highly curable. The

present methods of treatment include independent or a combination of surgical procedures, chemotherapy, radiotherapy and other targeted therapies depending on the stage of cancer. Since the afore-stated procedures are based on the late onset of colon cancer, it is important to develop early stage detection methods that have the potential to complement the aforementioned standard procedures to cure colon cancer or improve the life span of an individual.

Given the crucial association between miRNAs and cancer, several studies have classified different types of human cancers based on miRNA expression profiles [6,7]. Aberrant expression of miRNAs has been associated with almost every type of cancer. Expression analysis by RT-PCR revealed miR-221, miR-376a and miR-301 to be differentially expressed in pancreatic cancer providing new insights into pancreatic tumorigenesis [8]. A recent study has shown the let-7 miRNA cluster, miR-214, miR-199a\*, miR-200a and miR-100 to be frequently deregulated in ovarian cancer [9]. Similarly, miRNA expression profiling of prostate and breast cancer cells have revealed a total of 51 miRNAs that were either upregulated or downregulated in prostate cancer and a total of 133 miRNAs in breast tumors [10,11].

Expression profiling of miRNAs have been carried out in the last decade in colon cancer cell lines or tissues and these studies have established that aberrant expression of miRNAs is associated with colon or colorectal cancer. One of the studies analyzed the expression of 156 miRNAs in colorectal cancer and found a group of 13 miRNAs with aberrant expression profiles of which miR-31, miR-96, miR-133b, miR-135b, miR-145, and miR-183 were most significantly deregulated [12]. In another study, a total of 95 plasma miRNAs have been analyzed in colorectal cancer patients and 5 miRNAs have been found to be upregulated compared to their corresponding controls [13].

Similarly, in this study, we carried out expression profiling of miR-96, miR-422a and miR-584 in

colon cancer and show that all the three miRNAs are downregulated. Lower expression levels of these miRNAs have been correlated with increased tumor size. Also, we demonstrate that circulating level of miR-96 in plasma has the potential as a non-invasive biological marker for diagnosis of colon cancer.

### EXPERIMENTAL

### Patient characteristics and tissue samples

The tumor samples and the corresponding normal mucosa used in this study were collected from 60 patients diagnosed with CRC. After collection, all the samples were immediately flash frozen and stored at – 80 °C. The patients were being treated at The Second Affiliated Hospital of Wenzhou Medical University. The Medical Ethics Committee of The Second Affiliated Hospital of Wenzhou Medical University granted approval for this study. The samples were collected only after a written consent was obtained from the patients or their guardians. The histological grade of cancer was classified using the TNM staging system as per World Health Organization standards [20].

### Cell lines and culture

HT-29, HCT-116 and SW-620 cell lines were obtained from The Second Affiliated Hospital of Wenzhou Medical University. HT-29 and HCT-116 cell lines were cultured in McCoy's 5a media (Invitrogen, USA) supplemented with 10 % Fetal Bovine Serum (FBS). SW-620 cell line was cultured in L15 media (Sigma Aldrich, USA) supplemented with 2 mM glutamine and 10 % FBS. All the cell lines were cultured at 37 °C in a humidified atmosphere of 5 % CO<sub>2</sub>.

### RNA extraction and purification

RNA was extracted from the normal mucosa and the cancerous tissue samples according to the following procedure. Briefly, the flash frozen tissue samples were homogenized in presence of 0.5 ml of TRIZOL/50 mg of tissue. In the next phase separation step, 0.1 ml of chloroform was added to the aforementioned suspension solution. Following the phase separation step, RNA was precipitated by addition of 0.25 ml of isopropanol. Following centrifugation at 12000 g, the RNA pellet was washed with 80 % ethanol, centrifuged at 8000 g/10 min, air-dried and redissolved in 50 µL of DPEC treated water. The contaminating DNA was removed by DNAse treatment kit according to the manufacturer's instructions (Ambion, USA). For the purification of RNA, RNeasy column purification kit was used

(Qiagen, USA) according to the manufacturer's instructions. The total RNA was eluted in 50  $\mu$ L of DPEC treated water and quantified on a NanoDrop 1000 (Thermo Scientific, USA).

### Quantitative real-time polymerase chain reaction [RT-PCR]

The expression profiling of colon tissue miRNA was performed by Tagman assays. The miRNAs, miR-96, miR-584 and miR-422a, endogenous 5s rRNA primers and probes were purchased from Invitrogen, USA. The sequences of specific primers used in this study are indicated in Table 1. The final volume of the PCR mixture was 50 µL and it contained 0.3 µM probe, 0.3 µM of forward and reverse primers, 1X PCR buffer, 0.4 mM dNTPs and Taq DNA polymerase. The thermocycling conditions included initial denaturation at 95 °C for 5 min, followed by 35 cycles of 95 °C for 20 s, 58 °C for 30 s, and 68 •C for 40 s. Corresponding negative control without a template was used in all the runs and all the PCR runs were carried out in triplicates. miRNA expression levels were quantified using the ABI Prism 7000 Sequence detection system (Applied Biosystems, USA). The relative expression levels of the miRNAs were calculated using the 2- $\Delta\Delta$ CT method. 5s rRNA was used as an internal control to normalize the data.

 Table 1: Sequences of specific primers used in this study

miR-96	Forward: 5' CGGAGCACCTTACCCACTT 3' Reverse: 5' CAACCCACGGCACCATTCA 3'
miR- 584	Forward: 5' CAGGGTTGCTGTTACTGGAG 3'
	Reverse: 5' AATCTAACCCCACATTTCCCC 3'
miR- 422a	Forward: 5' GCTCCCATGCTATTAATATTCCTGGA 3' Reverse: 5' CGATCGCGCTTCAGCTCA 3'

### **Statistical analysis**

The differential expression of miRNA between colon cancer samples and the corresponding normal mucosa was assessed by a two-sided Student's t-test. The relationship between miR-96, miR-584 and miR-422a expression levels and clinicopathological characteristics were analyzed by non-parametric tests.

A *p*-value of < 0.05 was considered statistically significant. All statistical analyses were performed using GraphPad Prism 5 software and Microsoft Excel Version 14.6.5.

### RESULTS

### Expression levels of miR-96, miR-584 and miR-422a in colon cancer tissues

The expression levels of miR-96, miR-584 and miR-422a were analyzed in 60 colon cancer patient tissue samples and their adjacent normal mucosal counterparts using quantitative RT-PCR. Clinical characteristics of the 60 patients involved in this study are summarized in Table 2. Quantification analysis indicated that the levels of miR-584 and miR-422a miR-96, were significantly downregulated in most of the colon cancer tissue samples (Fig 1a, b and c). Next, we went ahead to analyze the differential expression levels of all the three miRNAs with respect to the adjacent normal mucosal tissues. Differential expression analysis using  $\Delta Ct$ method revealed that the expression levels for miR-96 decreased by a factor of 40-fold in 50 out of 60 cancer tissue samples relative to the normal mucosal tissue counterparts (Fig 2a, p <0.01). Similarly, the expression levels of miR-584 and miR-422a were decreased by a factor of 70fold in 55 out of 60 cancer tissue samples, and by a factor of 60-fold in 45 out of 60 cancer tissue samples (Fig 2b and c, p < 0.01).

### Expression of miR-96, miR-584 and miR-422a in colon cancer cell lines

We evaluated the expression levels of miR-96, miR-584 and miR-422a in the colon cancer cell lines HCT-116, SW-620 and HT-29. The results indicated significantly lower expression levels of miR-96, miR-584 and miR-422a in all the three colon cancer cell lines compared to the randomized normal mucosal tissue from patients (Fig 2d).

## Association between miRNA expression and clinicopathological characteristics of colon cancer patients

We evaluated the association between miR-96, miR-584 and miR-422a expression levels and clinicopathological characteristics in colon cancer patients. No significant correlation could be obtained between low expression levels of miR-96, miR-584 and miR-422a and characteristics such as patients' gender, age, the tumor and the clinical stages, lymph node metastasis and invasion. However, our correlation analysis suggested a significant correlation between lower expression levels of miR-96, miR-584 and miR-422a and the size of the tumor, irrespective of other clinicopathological characteristics. Our

results suggest that, lower expression levels of the aforementioned miRNAs are associated with increased tumor size (Table 2, all p < 0.05).

### Ratio of miR-96/miR-638 is a potential biomarker for colon cancer

We next wanted to investigate whether the expression levels of miR-96, miR-584 or miR-422a in blood could act as a non-invasive viable biological marker to discriminate colon cancer patients from healthy individuals. We measured the expression levels of miR-96, miR-584 and miR-422a independently in plasma samples from 25 colon cancer patients with respect to the healthy individual samples by quantitative RT-PCR. Recent studies have shown that, since the levels of plasma miR-638 are quite stable, miR-

638 can be used as a standard to improve the precision of the analysis [14]. Therefore, we analyzed the ratios of miR-96, miR-584 and miR422a levels to miR-638 levels in the plasma samples from healthy and colon cancer patients. The ratio of plasma levels of miR-584/miR-638 and miR-422a/miR638 from colon cancer patients could not be unambiguously distinguished from the aforementioned ratios from healthy individuals (Figure 3 a and b). However, the analysis revealed an increased ratio of miR- 96/miR-638 in the plasma samples from the colon cancer patients with respect to the healthy individuals (Figure 3c). Thus, the levels of miR-96/miR-638 can act as a potential biomarker in the prognosis of colon cancer.

 Table 2: Association between miR-96, miR-584 and miR-422a expression levels and clinicopathological characteristics in colon cancer patients

Patient	N	miP-96	mi <b>P</b> _584	miP_422a	
characteristics	IN	11117-30	11111-304	1111 <b>1-4</b> 22a	
Gender					
Male	35	0.264	0.179	0.243	
Female	25	0.149	0.215	0.243	
P-value		0.318	0.209	0.196	
Age (years)					
> 70	37	0.125	0.116	0.232	
< 70	23	0.187	0.288	0.148	
P-value		0.576	0.132	0.212	
Histological grade					
Well defined	43	0.115	0.252	0.174	
Poorly defined	17	0.198	0.165	0.279	
P-value		0.15	0.798	0.5	
pTNM stage					
+	42	0, 179	0.219	0.137	
	18	0.129	0.176	0.185	
P-value		0.379	0. 252	0.58	
pN stage					
NO	33	0.156	0.129	0.165	
N1	19	0.172	0.184	0.149	
N2	8	0.155	0.169	0.183	
P-value	-	0.698	0.539	0.236	
Lymphatic vessel					
invasion					
Positive	49	0.126	0.167	0.136	
Negative	11	0.18	0.148	0.194	
P-value		0.29	0.376	0.295	
Tumor size (cm)					
< 5	26	0.272	0.317	0.184	
> 5	34	0.269	0.242	0.211	
P-value		0.025	0.034	0.03	



**Figure 1:** Expression levels of (a) miR-96, (b) miR-584 and (c) miR-422a in 60 colon cancer patient tissues. Quantification analysis indicated that the levels of miR-96, miR-584 and miR-422a were significantly downregulated in most of the colon cancer tissue samples

### DISCUSSION

The last decade has witnessed advancement in colon cancer research and diagnosis. however it is still one of the deadliest form of cancer with a number of fatalities per year. Since the survival rates of colon cancer patients depend on the early detection of cancer it has become necessary to identify biological markers for a better diagnosis. In the last decade, different classes of molecules have been identified as potential biomarkers for colon cancer and one such intriguing class of molecules is the miRNAs [15]. Large-scale miRnome studies have been carried out in different human solid tumors of breast, colon, pancreas, lung and stomach and identified a large cohort of miRNAs that have or under-expressed been over-expressed [16,17]. Several other studies identified such "miRNA signatures" in cancer and conclusively establish that miRNAs play a critical role in cancer pathogenesis, playing role of either oncogenes or tumor suppressor genes.

Several published literature has provided sufficient evidence that miR-96, miR-584 and miR-422a are often de-regulated in cancer causing pathways. While these miRNAs have been associated with colon cancer and reported previously [18], to the best of our knowledge, a comprehensive validation and characterization of these miRNAs in colon cancer patient tissue samples and cell lines, and the ability of these miRNAs to act as a viable biomarker in the prognosis of colon cancer is still required.

Firstly, the levels of miR-96, miR-584 and miR-422a were assessed in colon cancer tissue and adjacent normal mucosal tissue counterparts using real-time PCR owing to its rapid quantification as well as sensitivity. Significantly lower expression levels of these miRNAs in the cancer tissue compared with the healthy tissue indicated that miR-96, miR-584 and miR-422a are down-regulated in colon cancer. Next, the levels of these miRNAs were assessed in some routinely used colon cancer cell lines.

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**Figure 2:** Differential expression analysis of all (a) miR-96, (b) miR-584 and (c) miR-422a in colon cancer tissues (CCT) with respect to the adjacent normal mucosal tissues (NAT). Differential expression analysis was carried out by  $\Delta C_t$  method. Expression levels of miR-96, miR-584 and miR-422a were decreased by a factor of 40-fold, 70-fold and 60-fold respectively (d) Expression levels of miR-96, miR-584 and miR-422a in HCT-116, SW-620 and HT-29 colon cancer cell lines. miRNA expression analysis in colon cancer cell lines is relative to three randomly selected normal colorectal tissues from 60 colon cancer patients (*bars, s. d* All, *p* < 0.05)



**Figure 3:** Comparison of miR-584, miR-422a and miR-96 levels in the plasmas from normal individuals and colon cancer patients. The ratio between (a) miR-584, (b) miR-422a and (c) miR-96 to miR-638 were quantitated by RT-PCR. *Bars*, s.d. As it is evident from the figure, the miR-96/miR-638 ratio from the colon cancer plasma samples could be unambiguously distinguished from the normal plasma samples

Similar to the patient tissue samples, HCT-116, SW-620 and HT-29 colon cancer cell lines showed subdued expression levels of these miRNAs. Further, the correlation between low miRNA expression levels and clinicopathological characteristics of colon cancer patients was evaluated. This study indicated an interesting correlation between low expression levels of miR-96, miR-584 and miR-422a with increase in size of the tumor in colon cancer patients.

A similar study has been carried out in the past in 29 colorectal cancer tissues and the study shows the expression of miR-21, -31, -143 and -145 to

be altered in CRC and is related to clinicopathological characteristics such as tumor size and lymph node metastasis [19]. Thus, it is evident that miR-96, miR-584 and miR-422a show altered expression levels in colon cancer and it is likely that these miRNAs have the potential as a biomarker in the prognosis of colon cancer.

miRNAs have been shown to have a promising potential as prognostic biomarkers for various types of cancer. Therefore, we investigated the predictive value of miR-96, miR-584 and miR-422a in blood plasma as a viable prognostic biomarker for colon cancer. Quantification of the ratios of miR-584/miR-638 levels and miR-422a/miR638 levels did not yield success, as a clear distinction could not be obtained with respect to healthy individuals. However, we found the ratio of miR-96 to miR-638 to be increased in the plasma samples of colon cancer patients compared to healthy individuals. The distinction was remarkable and thus the levels of miR-96/miR-638 have the potential as a viable non-invasive biomarker for colon cancer. Similar quantification studies were carried out in the plasma samples of hepatocellular carcinoma patients to identify potential biomarkers [14].

### CONCLUSION

This study shows that miR-96, miR-584 and miR-422a are downregulated in colon cancer and are associated with tumor size. The levels of miR-96/miR-638 has the potential of a non-invasive biomarker in colon cancer. Translating this finding into clinical practice would require a thorough and systematic development of diagnostics followed by randomized investigations and validations for demonstrating the efficacy of such non-invasive biomarkers in colon cancer diagnosis.

### DECLARATIONS

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#### **Conflict of Interest**

No conflict of interest associated with this work.

#### **Contribution of Authors**

The authors declare that this work was done by

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### REFERENCES

- 1. Bartel DP. MicroRNAs: genomics, biogenesis, mechanism, and function. Cell 2004; 116: 281-297.
- 2. Garzon R, Calin GA, Croce CM. MicroRNAs in cancer. Annu Rev Med 2009; 60: 167-179.
- 3. Esquela-Kerscher A, Slack FJ. Oncomirs—microRNAs with a role in cancer. Nat Rev Cancer2006; 6: 259-269.
- Pillai RS. MicroRNA function: multiple mechanisms for a tiny RNA? Wiley Interdiscip Rev RNA 2005; 11: 1753-1761.
- 5. Siegel R, Ma J, Zou Z, Jemal A. Cancer statistics, 2014. CA Cancer J Clin 2014; 64: 9-29.
- Lu J, Getz G, Miska EA, Alvarez-Saavedra E, Lamb J, Peck D, Sweet-Cordero A, Ebert BL, Mak RH, Ferrando AA. MicroRNA expression profiles classify human cancers. Nature 2005; 435: 834-838.
- Volinia S, Calin GA, Liu C-G, Ambs S, Cimmino A, Petrocca F, Visone R, Iorio M, Roldo C, Ferracin M. A microRNA expression signature of human solid tumors defines cancer gene targets. Proc Natl Acad Sci U S A 2006; 103: 2257-2261.
- Lee EJ, Gusev Y, Jiang J, Nuovo GJ, Lerner MR, Frankel WL, Morgan DL, Postier RG, Brackett DJ, Schmittgen TD. Expression profiling identifies microRNA signature in pancreatic cancer. Int J Cancer 2007; 120: 1046-1054.
- Yang H, Kong W, He L, Zhao J-J, O'Donnell JD, Wang J, Wenham RM, Coppola D, Kruk PA, Nicosia SV. MicroRNA expression profiling in human ovarian cancer: miR-214 induces cell survival and cisplatin resistance by targeting PTEN. Cancer Res 2008; 68: 425-433.
- Porkka KP, Pfeiffer MJ, Waltering KK, Vessella RL, Tammela TLJ, Visakorpi T. MicroRNA expression profiling in prostate cancer. Cancer Res 2007; 67: 6130-6135.
- Blenkiron C, Goldstein LD, Thorne NP, Spiteri I, Chin S-F, Dunning MJ, Barbosa-Morais NL, Teschendorff AE, Green AR, Ellis IO, et al. MicroRNA expression profiling of human breast cancer identifies new markers of tumor subtype. Genome Biol 2007; 8: R214.

- Bandrés E, Cubedo E, Agirre X, Malumbres R, Zarate R, Ramirez N, Abajo A, Navarro A, Moreno I, Monzó M, et al. Identification by Real-time PCR of 13 mature microRNAs differentially expressed in colorectal cancer and non-tumoral tissues. Mol Cancer 2006; 5: 29.
- Ng EK, Chong WW, Jin H, Lam EK, Shin VY, Yu J, Poon TC, Ng SS, Sung JJ. Differential expression of microRNAs in plasma of colorectal cancer patients: a potential marker for colorectal cancer screening. Gut 2009; 58: 1375-138.
- 14. Shigoka M, Tsuchida A, Matsudo T, Nagakawa Y, Saito H, Suzuki Y, Aoki T, Murakami Y, Toyoda H, Kumada T, et al. Deregulation of miR-92a expression is implicated in hepatocellular carcinoma development. Pathol Int 2010; 60: 351-357.
- Cho WCS. MicroRNAs: potential biomarkers for cancer diagnosis, prognosis and targets for therapy. Int J Biochem Cell Biol 2010; 42: 1273-1281.

- 16. Lee YS, Dutta A. MicroRNAs in cancer. Annu Rev Pathol 2009; 4: 199-227.
- 17. Calin GA, Croce CM. MicroRNA signatures in human cancers. Nat Rev Cancer 2006; 6: 857-866.
- Faltejskova P, Svoboda M, Srutova K, Mlcochova J, Besse A, Nekvindova J, Radova L, Fabian P, Slaba K, Kiss I, et al. Identification and functional screening of microRNAs highly deregulated in colorectal cancer. J Cell Mol Med 2012; 16: 2655-2666.
- Slaby O, Svoboda M, Fabian P, Smerdova T, Knoflickova D, Bednarikova M, Nenutil R, Vyzula R. Altered expression of miR-21, miR-31, miR-143 and miR-145 is related to clinicopathologic features of colorectal cancer. Oncology 2007; 72: 397-402.
- Edge SB, Compton CC. The American Joint Committee on Cancer: the 7th edition of the AJCC cancer staging manual and the future of TNM. Ann Surg Oncol 2010; 17(6):1471-1474.