

Original Research Article

Anti-cancer and antioxidant properties of phenolics isolated from *Toona sinensis* A Juss acetone leaf extract

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Abstract

Purpose: To investigate the antioxidant and anticancer activities of phenolics from the leaf extract of *Toona sinensis* (TS).

Methods: Acetone leaf extract of TS was screened for total phenolic and flavanoid contents, and the flavonoids were subjected to high performance liquid chromatographic (HPLC) analysis. Antioxidant properties were assessed via oxygen radical absorbance capacity (ORAC), peroxy radical scavenging capacity (PSC) and cellular antioxidant activity (CAA), while anti-proliferative activity ins HepG2 cell line was assessed using methylene blue assay.

Results: The extract contained 36.02 ± 0.24 mg of gallic acid equiv/g dry weight (DW) and 20.24 ± 1.73 mg of catechin equiv/g DW of total phenolic and total flavanoid, respectively. The levels of rutin and quercitrin were 0.51 and 19.55 mg/g, respectively. Epicatechin, gallic acid, quercitrin, isoquercetin were not detected. The extract showed significant antioxidant potential and high anti-proliferation capacity with low cytotoxicity against HepG2 cell in vitro. The underlying mechanism of anti-proliferative effect was induction of apoptosis.

Conclusion: TS leaf extract possesses significant in vitro antioxidant properties and anti-proliferative effect against HepG2 cells, which make it a potential anticancer drug source.

Keywords: *Toona sinensis*, Phenolics, Antioxidants, HepG2 cells, Anti-proliferation

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INTRODUCTION

Phytochemicals are plant-based non-nutrient secondary metabolites, which provide multiple positive health effect. These metabolites are increasingly recognized as efficacious components in the prevention and management of chronic disease, such as cancer, diabetes, and cardiovascular diseases [1].

Toona sinensis A. Juss is a deciduous tree which belongs to the family Meliaceae. All parts of TS are medicinally useful. Indeed the plant possesses many biological functions such as

antitumor activity [2], antioxidant capacity [3], and anti-inflammatory properties [4]. It is also used for treating severe acute respiratory syndrome [5], and for improving hyperglycemia and dyslipidemia [6]. The young leaves of TS are popular in traditional Chinese medicines and have been used for long as a safe vegetable source [7]. Non-polar extracts of TS leaves contain constituents that can prevent type-2 diabetes and hepatosteatorosis [8]. Previous studies showed that TS leaves possess anti-oxidative activity and anti-inflammatory activities [9]. TS is used for treatment of liver cancer in Gansu province of China [10]. Although the anti-

diabetic, antioxidant and anti-inflammatory activities of TS leaves have been reported, there is lack of information on their phenolic profiles, cellular antioxidant activity and *in vitro* anti-proliferative activity.

EXPERIMENTAL

Chemicals and materials

2,2'-Azobis-amidino-propane (ABAP), catechin hydrate, gallic acid, Folin-Ciocalteu reagent and dichlorofluorescein diacetate (DCFH-DA) were obtained from Sigma Chemical Co. (St. Louis, MO, US). All solvents used were of analytical grade. Williams medium E (WME), insulin, and other cell culture media were purchased from Gibco U.S. Biotechnology Co. Fetal bovine serum (FBS) was bought from Tianhang Biotech Co. Inc. (Zhejiang, China). HepG2 cell line was obtained from the Sun Yat-sen University Culture Collection.

Samples treatment and preparation

One kilogram of the leaves was collected from Anhui-China in spring 2014. The leaves were washed with distilled water to remove dirt, followed by freezing at -80 °C for 24 h and vacuum freeze-drying. The dried leaves were ground into powders (60 mesh) and stored at -80 °C prior to analysis.

Extraction procedures

The powder extraction procedure was a modification of the classical method described by Chu *et al* [11]. The powder (10 g) was homogenized with chilled acetone (80 %) for 5 min. The homogenate was filtered under vacuum, and the residue was re-extracted and filtered three times. The extracts were pooled in flasks and evaporated at 45 °C until no acetone was left. The residue was dissolved in a little volume of distilled water and stored at -40 °C until used.

Estimation of total phenolic content

Total phenolic content was determined as gallic acid equivalent using gallic acid standard curve [12], and the final value was expressed as milligram of gallic acid equivalent (GAE) per gram DW.

Measurement of total flavonoid content

The total flavonoid content was determined using the sodium borohydride/chloranil-based method as described earlier [13]. Values were calculated

using catechin hydrate standard curve and expressed as milligram of catechin equivalent (CE)/g DW. Data are showed as mean \pm SD.

Analysis of flavonoids by HPLC

Flavonoid compounds in the sample extract were quantified using HPLC. HPLC system was assembled with Waters Associates chromatography (Waters Corp, Milford, US), and spectrophotometer set at 360 nm was used. A Sun Fire C18 phase column (5 μ m particle size, 250 mm \times 4.6 mm) was employed for chromatographic separation at 30 °C. The binary mobile phase consisted of methanol (A) and water (B). After injection of 10 μ L sample, the system was eluted with 5 % A for 5 min; then increased to 10, 15, 20 and 45 % each at 5-min intervals. Then the gradient solvent A was increased to 100 % in 5 min, kept at 100 % for 10 min, and recycled back to 5 % after 5 min, and kept for 5 min prior to running the next sample. All process flow rate was 1.0 mL/min. The peaks were identified through their retention times by comparing with UV-visible spectra of standards.

Antioxidant assays

Total antioxidant activities were determined by hydrophilic peroxy radical scavenging capacity assay and oxygen radical scavenging capacity assay. Hydrophilic peroxy radical scavenging capacity of the extract was evaluated using the method as outlined previously [14]. In a run, pure compounds or sample extract appropriately diluted in phosphate buffer were transferred into reaction cells on a 96-well plate followed by the addition of DCFH. The reaction was initiated by adding ABAP and read at once. Each set of dilutions and control was added in triplicate in 96-well plate. Fluorescence was monitored at 485 nm excitation and 535 nm emission in a fluorescent spectrophotometer (SoftMax systems, Molecular Devices, US). Result was presented as micromoles of vitamin C equivalent (VCE) per gram of sample.

Oxygen radical scavenging capacity was measured as described by Prior *et al* [15,16], using black, clear-bottom, 96-well microplates. Extract diluent and trolox standard were added to appropriate wells and incubated at 37 °C for 10 min. Fluorescein working solution was added to each well and incubated at 37 °C for 20 min. After incubation, freshly prepared ABAP was added to each well (except the control well) and the wells were immediately read in Fluoroskan Ascent FL plate-reader at excitation of 485 nm and emission of 535 nm for 35 cycles every 4.5

min. ORAC value was expressed as mean micromoles of trolox equivalent (TE) per g DW.

Cell culture

HepG2 cells were cultured in WME, containing 2 mM L-glutamine, 10 mM Hepes, 5 µg/mL insulin, 0.05 µg/mL hydrocortisone, 50 µg/mL streptomycin, 50 units/mL penicillin, 5 % FBS and 100 µg/mL gentamicin. Cells were incubated at 37 °C with 5 % CO₂, and seeded/sub-cultured when in exponential growth phase.

Cytotoxicity of extract against HepG2

Cytotoxicity was measured as described formerly [17]. The concentration of extract that decreased cell numbers by 10 % when compared to the control was considered to be cytotoxic.

Cellular antioxidant activity (CAA)

The cellular antioxidant activity of the extract was determined following the method of Wolfe *et al* [18]. HepG2 cells were seeded into black, clear-bottom, 96-well microplate. Cell medium was removed after 24 h of seeding, and the cells were washed with PBS, and treated in triplicate with solutions containing different concentrations of quercetin or extracts plus DCFH-DA dissolved in antioxidant treatment medium for 1 h. If a PBS wash was to be utilized, wells were washed with PBS followed by addition of ABAP solution. The microplate was read in a Fluoroskan Ascent FL plate-reader at excitation of 485 nm and emission of 535 nm for 13 cycles every 5 min. The area under the fluorescence-versus-time curve was calculated to determine CAA value. EC₅₀ values were converted to CAA values, which were expressed as micromoles of quercetin equivalent (QE) per 100 g of sample, using the mean EC₅₀ value for quercetin from three separate experiments.

Inhibition of cell proliferation

The anti-proliferation effects of the extract were determined using cell methylene blue colorimetric assay [17]. The anti-proliferative effects were assessed by the EC₅₀ values, expressed as milligrams of sample extract per milliliter.

Assessment of apoptosis by flow cytometry

The apoptosis of HepG2 cell was analyzed by flow cytometer employing an annexin V-FITC and PI apoptosis detection kit. In brief, the cells were treated with 0, 4, 6 and 8 mg/mL of extract for 24 h followed by harvesting, washing and

incubation with annexin V-FITC and PI at room temperature for 20 min. Aluminum foil was used during all staining processes to avoid light and; the cells were analyzed by flow cytometer.

Statistical analysis

Statistical analysis was performed using SPSS software 13.0 (SPSS Inc., Chicago, IL, USA), while dose-effect analysis was done using Calcsyn software version 2.0 (Biosoft, Cambridge, UK). All data are reported as mean ± SD (n = 3).

RESULTS

Total phenolic and flavonoid contents

Measured values of total phenolic and flavonoid contents are given in Table 1. The total phenolic content of the leaf extract of TS was 36.02 mg of GCE/g DW, which was less than reported level of polyphenolic content (79.90 mg of GCE/g) in TS leaf [19]. However, this value was 45.28 times and 98.25 times higher compared to spinach and cabbage, respectively [11]. This indicated that TS leaf is a good source of phenolics.

Total flavonoid content of TS leaves extract was 20.24 ± 1.73 mg of CE/g DW. This amount was relatively higher than the value reported earlier (12.92 ± 0.95 mg of CE/g sample) in the aqueous extract of TS leaf [20]. In the studied sample, contribution of total flavonoids to total phenolics was 32.92 ± 2.65 %.

Table 1: Phytochemical content of TS leaf extract

Parameter	Value
Total phenolics (mg of gallic acid equiv./g DW)	36.02±0.24
Total flavonoids (mg of catechin equiv./g DW)	20.24±1.73
Percent contribution of total flavonoids to total phenolics (%)	32.92±2.65
PSC values (µmol of vitamin C equiv./g DW)	243.26±26.59
ORAC values (µmol of trolox equiv./g DW)	328.84±32.75

Values are mean ± SD (n = 3)

Flavonoid compounds

Flavonoids such as quercitrin, rutin, epicatechin, gallic acid, quercetin and isoquercetin were identified and quantified by comparing retention times with those of authentic standards using HPLC method. Only two compounds quercitrin and rutin were detected and quantified at concentrations of 19.55 mg/g DW (1.95 %) and

0.51 mg/g DW (0.51 %), respectively (Table 2, Figure 1). Epicatechin, gallic acid, quercetin, and isoquercetin were below the detection limit.

Table 2: Content of antioxidant compounds of TS leaf extract (mg/g DW)

Compound	Content (mg/g of DW)
Quercitrin	19.55±0.55
Rutin	0.51±0.02
Epicatechin	ND
Gallic acid	ND
Quercetin	ND
Isoquercetin	ND

Total *in vitro* antioxidant activity

Reactive oxygen species (ROS) have been reported as the causative factor in many chronic diseases. Oxidative stress caused by ROS can be prevented by natural antioxidant compounds [21]. The results of *in vitro* antioxidant activities of the extract are displayed in Table 1 and Figure 2. Although antioxidant activities of plant extracts can be determined by various methods, PSC and ORAC methods are the latest and more authentic techniques. PSC assay incorporates DCFH-DA as a fluorescent probe, and is suitable for analyzing both hydrophilic and lipophilic antioxidants. This method is reliable, sensitive, rapid, precise, and reproducible [22]. The PSC value was $243.26 \pm 26.59 \mu\text{mol VCE/g DW}$

(Table 1 and Figure 2). Food and pharmaceutical industries usually use oxygen radical absorbance capacity (ORAC) assay to test total antioxidant activity. In a typical ORAC assay, the loss of fluorescence of B-phycoerythrin is an indication of the degree of damage from its reaction with the peroxy radical [16]. The antioxidant activity is valued by assessing the area under the fluorescence decay curve of the extract relative to that of the blank containing PBS [14]. The TS leaf extract had an ORAC value $328.84 \pm 32.75 \mu\text{mol TE/g DW}$ (Table 1).

Cellular antioxidant activity

The kinetic of DCFH oxidation by peroxy radicals generated from ABAP is exhibited in Figure 3. Increases in fluorescence due to formation of DCF were inhibited by quercetin or TS leaf extract in a dose-dependent manner. DCFH oxidation was suppressed whether the cells had been washed with PBS or not between the antioxidant and the ABAP treatments. The EC_{50} values of TS leaf extract by CAA were $1518.6 \pm 170.74 \text{ mg/mL}$ in wash protocol, and $567.48 \pm 1845 \text{ mg/mL}$ in no-wash protocol. The EC_{50} values were converted to CAA values, which gave $762.55 \pm 85.97 \mu\text{mol QE/100 g sample}$ in wash protocol, and $1719.8 \pm 56.04 \mu\text{mol QE/100 g sample}$ in no-wash protocol. In other words, 44.34 % of the constituents can enter the cells.

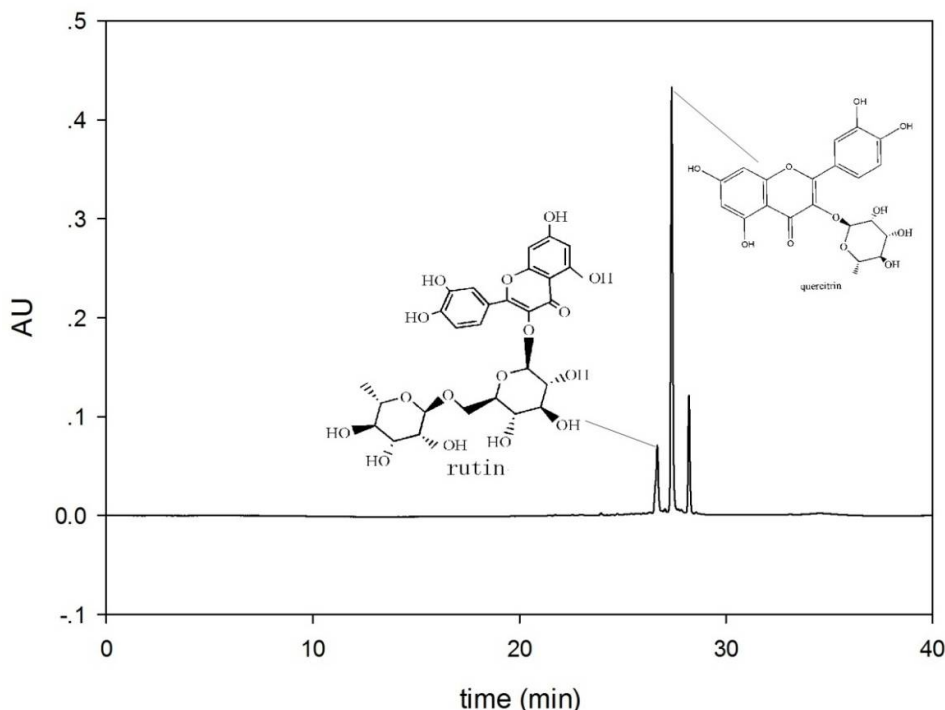


Figure 1: HPLC analysis of flavonoids from TS leaf extract

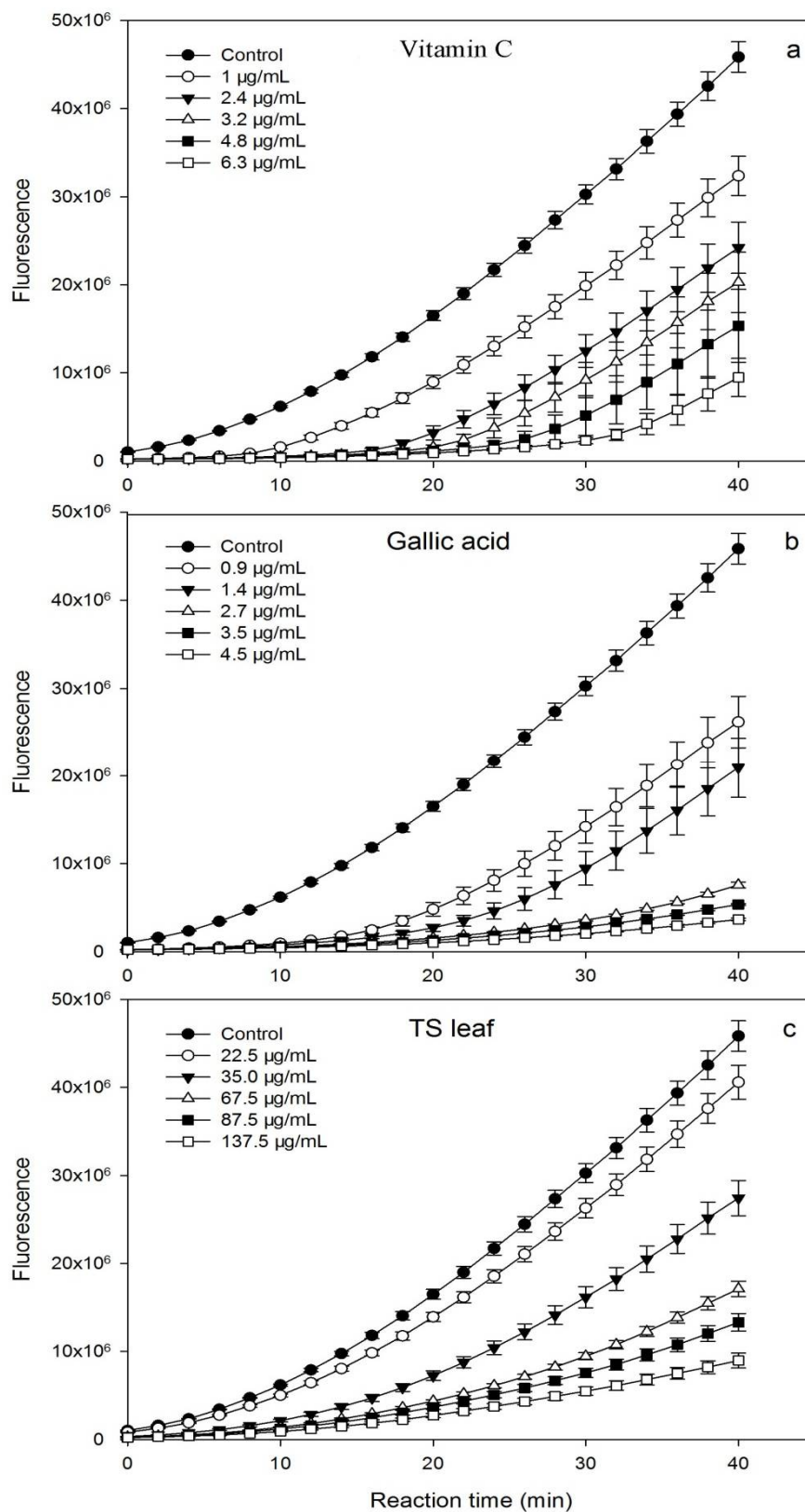


Figure 2: Time kinetics and dose-response of hydrophilic antioxidant activities of Vitamin C (a), gallic acid (b) and TS leaf extract (c) (mean ± SD, n = 3)

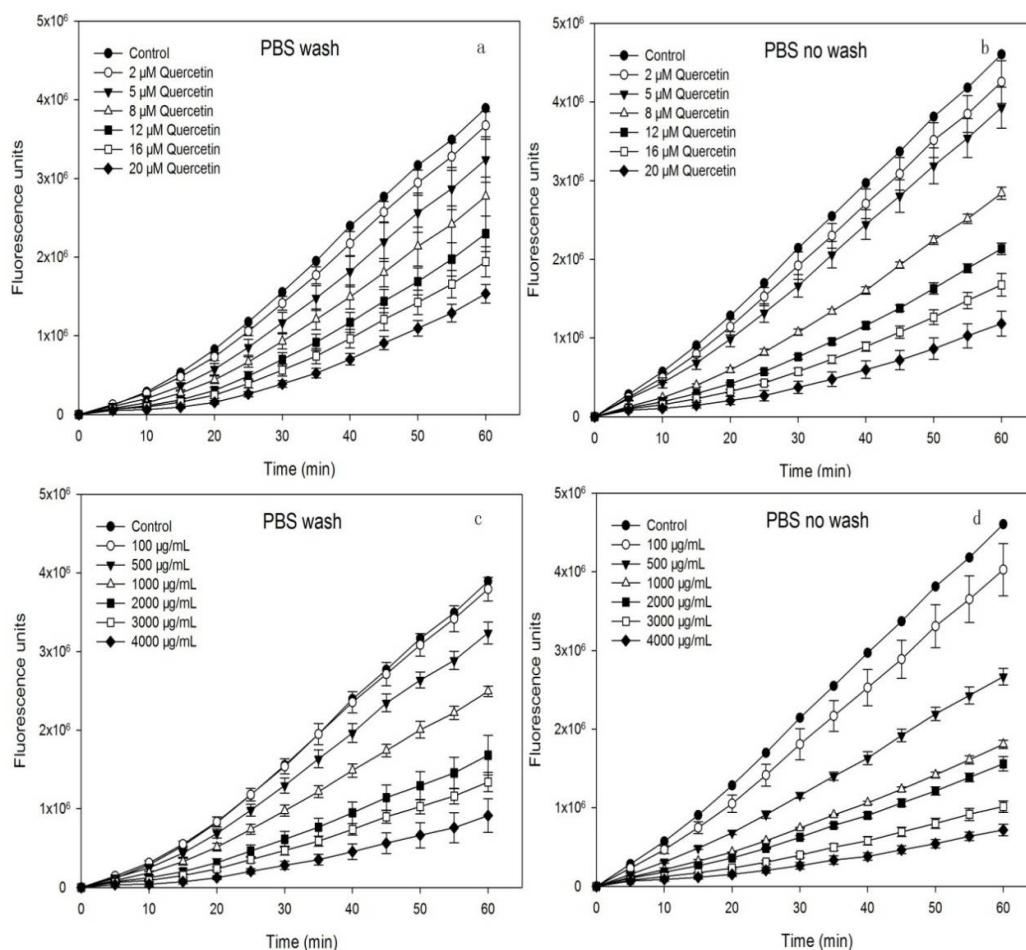


Figure 3: Peroxyl radical-induced oxidation of DCFH to DCF in hepG2 cells and the antioxidant activity by quercetin (a, b) and TS leaf extract (c, d) over time (Mean \pm SD, n = 3)

Cellular antioxidant quality

The cellular antioxidant qualities of the extract were evaluated from their CAA values, phenolic content and flavonoid content, and expressed as micromole of QE/100 μ mol of phenolics or flavonoids. In the PBS wash protocol, CAA value of TS leaf extract relative to total phenolics was $3.60 \pm 0.43 \mu\text{mol QE}/100 \mu\text{mol}$; while CAA value relative to total flavonoids was $11.00 \pm 1.71 \mu\text{mol QE}/100 \mu\text{mol}$. In the no-wash protocol, CAA value of the extract relative to total phenolics was $8.12 \pm 0.26 \mu\text{mol QE}/100 \mu\text{mol}$; while CAA value relative to total flavonoids was $24.75 \pm 1.48 \mu\text{mol QE}/100 \mu\text{mol}$.

Anti-proliferative and cytotoxic effect on HepG2 cancer cells

Figure 4 shows anti-proliferative activity and cytotoxic effects of the extract. The median effective dose (EC_{50}) for inhibition of HepG2 cell proliferation was $7.45 \pm 0.37 \text{ mg/mL}$.

Induction of apoptosis by TS leaf extract

Pro-apoptotic effect is regarded as the most important route for anti-proliferative action of phytochemicals toward cancer cells [23]. The apoptosis effect of TS leaf extract was assayed by flow cytometric analysis and the results of annexin V-FITC/PI staining assay are displayed in Figure 5. It was observed that the extract exhibited apparent effect on the percentage of early apoptotic cells. HepG2 cells treated with the leaf extract had an increase in the ratio of early stage apoptosis cells compared to the blank group. Remarkable enhancement was observed in cells treated with 6 mg/mL and 8 mg/mL of the extract. However, the percentage of late apoptosis in HepG2 cells was minor. The inhibiting effect of the extract on HepG2 cells proliferation was demonstrated mainly in early stage, implying a potential value of TS leaf as anticancer remedy.

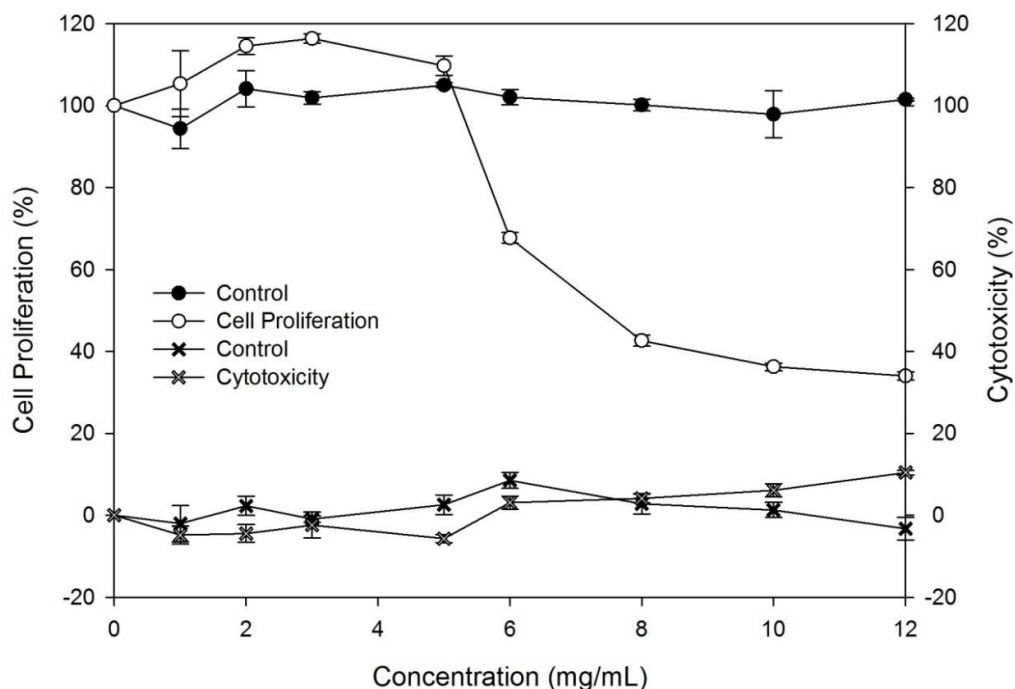


Figure 4: Inhibition of HepG2 cancer cells proliferation and cytotoxicity by TS leaf extract (mean \pm SD, n = 3)

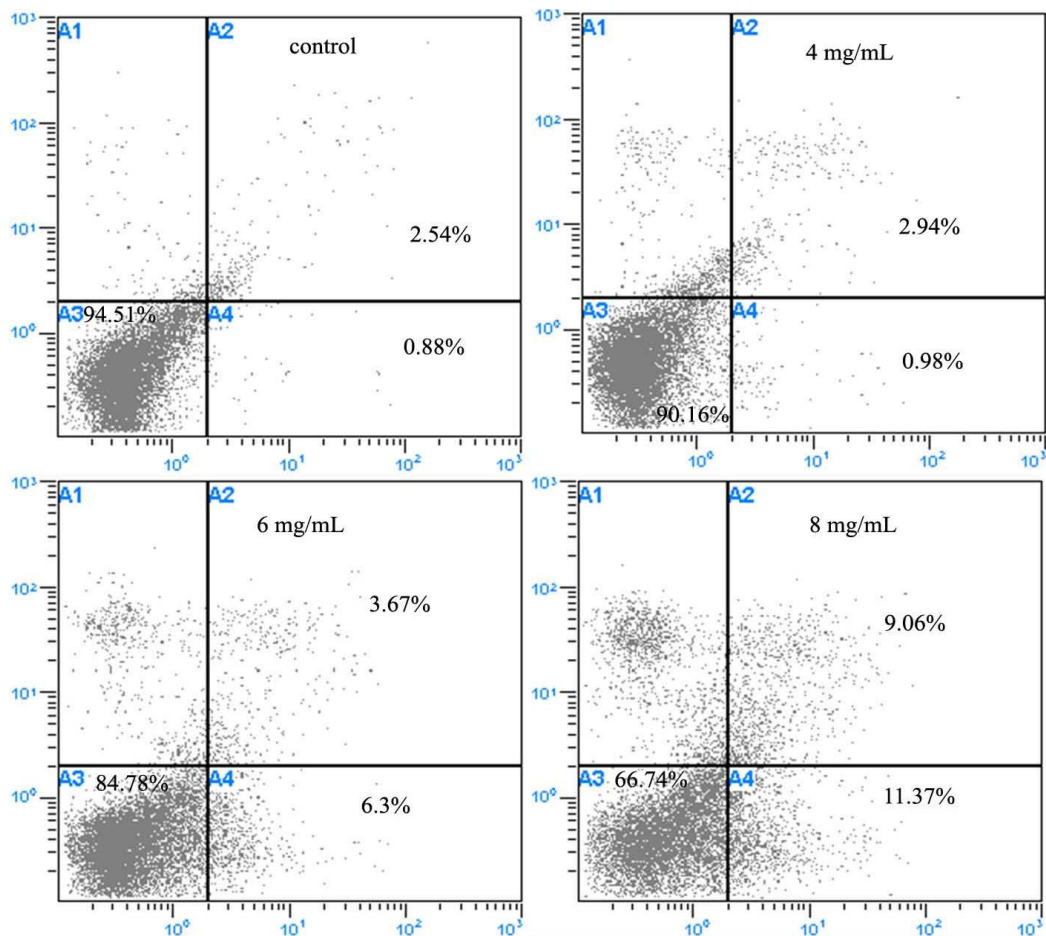


Figure 5: Effect of TS leaf extract on apoptosis in HepG2 cells, determined by flow cytometry with Annexin-V/PI

DISCUSSION

This study was focused on antioxidant and anti-proliferation activities of TS leaves extract towards HepG2 liver cancer cell line. Quercitrin and rutin were identified as the main flavonoids. Quercitrin (glycosylated form of quercetin), is the most common flavonoid found in nature and has been reported in vegetables and fruits such as apples, grapes, onions, tea [24]. Quercitrin is a bioflavonoid with antioxidant properties and is more easily absorbed than other forms of quercetin [25]. It is protective against UVB irradiation-induced oxidative damage to skin [24]. Rutin is a kind of flavanol glycoside, richly present in black tea, apple skin peels and buckwheat. It has been reported that rutin possesses antioxidant properties, scavenges hydroxyl and superoxide radicals and inhibits lipid peroxidation [26]. Thus the presence of quercitrin and rutin indicate that TS leaf has significant nutrition and health benefits. In plant extracts antioxidant activity cannot be ascertained by a single method due to the complex reactivities of phytochemicals.

Three methods i.e. PSC, ORAC and CAA were used to evaluate antioxidant potential of TS leaves. PSC and ORAC assays incorporate fluorescent probe to monitor reaction. These methods are rapid, precise, reliable, reproducible, and sensitive; and can produce acceptable results comparable to common chemical assays [12]. The assays also take into account the bioavailability, uptake and metabolism of the extract [18].

HepG2 cell culture model is relatively fast, represents the complexity of biological systems and is cost-effective. TS extract had lower EC₅₀ value and higher antioxidant activity in the no-wash protocol than in wash-PBS protocol. This is due to the fact that PBS affects extracellular antioxidant activity, thereby reducing the intracellular antioxidant activity. These data demonstrate that the phytochemicals in the extract were able to pass through the cell membrane. Liao *et al* [27] observed that the inhibitory 50 % concentration values in the DPPH radical-scavenging activity of TS leaf extracts was 0.209 mg/mL. Although the mechanism of different antioxidant activities vary, our results are nonetheless comparable to that of Liao *et al* [27]. This indicates that TS leaf extract inhibits oxidative stress.

In an interesting study, TS leaf extract significantly inhibited ovarian cancer at the G2 phase in an in vivo xenograft model with no significant toxicity [28]. Furthermore, it has been

reported that water leaf extracts of TS s had no acute lethal effect at test dose in male and female mice [29]. Our data also showed that the leaf extract of TS had antitumor effect but no cytotoxic effect.

CONCLUSION

The findings of this study reveal that TS leaf extract is rich in flavonoids, bioactive compounds responsible for its cellular antioxidant capacity and antitumor effects. These results provide scientific basis for the use of TS as a folk medicine for treating patients with liver cancer. The safety and efficacy of TS leaf makes it a potential natural medicine for managing hepatocellular carcinoma.

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REFERENCES

1. Liu RH. Potential synergy of phytochemicals in cancer prevention: Mechanism of action. *J Nutr* 2004; 134(12): 3479S-3485S.
2. Yang S, Zhao Q, Xiang H, Liu M, Zhang Q, Xue W, Song B, Yang S. Antiproliferative activity and apoptosis-inducing mechanism of constituents from *Toona sinensis* on human cancer cells. *Cancer Cell Int* 2013; 13(1):1-8.
3. Dong X, Zhu Y, Bao G, Hu F, Qin G. New limonoids and a dihydrobenzofuran norlignan from the roots of *Toona sinensis*. *Molecules* 2013; 18(3): 2840-2850.
4. Wang C, Tsai Y, Hsieh Y, Lin R, Lin C. The aqueous extract from *Toona sinensis* leaves inhibits microglia-mediated neuroinflammation. *Kaohsiung J Med Sci* 2014; 30(2):73-81.
5. Chen C, Michaelis M, Hsu H, Tsai C, Yang KD, Wu Y, Jr. Cinatl J, Doerr HW. *Toona sinensis* roem tender leaf extract inhibits SARS coronavirus replication. *J Ethnopharmacol* 2008; 120(1): 108-111.
6. Yu BC, Tsai YT, Ta NL, Yu WJ, Chang SJ. Proteomic analysis of glucose and lipid metabolism in liver of diabetic mice fed with *toona sinensis* leaf ethanol extract. *Ann Nutr Metab* 2013; 63: 1292-1292.
7. Li J, Eidman K, Gan X, Haefliger OP, Carroll PJ, Pika J. Identification of (S,S)-gamma-Glutamyl-(cis-S-1-propenyl) thioglycine, a Naturally Occurring Norcysteine Derivative, from the Chinese Vegetable *Toona sinensis*. *J Agr Food Chem* 2013; 61(31):7470-7476.
8. Hsieh TJ, Tsai YH, Liao MC, Du YC, Lien PJ, Sun CC, Chang F, Wu YC. Anti-diabetic properties of non-polar *Toona sinensis* Roem extract prepared by supercritical-CO₂ fluid. *Food Chem Toxicol* 2012; 50(3): 779-789.

9. Hsiang C, Hseu Y, Chang Y, Kumar KJS, Ho T, Yang H. *Toona sinensis* and its major bioactive compound gallic acid inhibit LPS-induced inflammation in nuclear factor-kappa B transgenic mice as evaluated by in vivo bioluminescence imaging. *Food Chem* 2013; 136(2): 426-434.
10. Zhen H, Zhang YF, Fang ZJ, Huang ZW, You CG, Shi P. *Toona sinensis* and moschus decoction induced cell cycle arrest in human cervical carcinoma HeLa cells. Evidence-based complementary and alternative medicine: eCAM 2014; 2014:121276.
11. Chu YF, Sun J, Wu XZ, Liu RH. Antioxidant and anti-proliferative activities of common vegetables. *J Agr Food Chem* 2002; 50(23):6910-6916.
12. Chen Y, Wang G, Wang H, Cheng C, Zang G, Guo X, Liu RH. Phytochemical profiles and antioxidant activities in six species of ramie leaves. *Plos One* 2014; 9(9).
13. He X, Liu D, Liu RH. Sodium Borohydride/Chloranil-Based assay for quantifying total flavonoids. *J Agr Food Chem* 2008; 56(20): 9337-9344.
14. Chen Y, Chen G, Fu X, Liu R. Phytochemical profiles and antioxidant activity of different varieties of adinandra tea (*Adinandra jack*). *J Agr Food Chem* 2014; 63(1): 169-176.
15. Prior RL, Hoang H, Gu LW, Wu XL, Bacchiocca M, Howard L, Hampsch-Woodill M, Huang DJ, Ou BX, Jacob R. Assays for hydrophilic and lipophilic antioxidant capacity (oxygen radical absorbance capacity (ORAC(FL)) of plasma and other biological and food samples. *J Agr Food Chem* 2003; 51(11): 3273-3279.
16. Huang DJ, Ou BX, Hampsch-Woodill M, Flanagan JA, Prior RL. High-throughput assay of oxygen radical absorbance capacity (ORAC) using a multichannel liquid handling system coupled with a microplate fluorescence reader in 96-well format. *J Agr Food Chem* 2002; 50(16): 4437-4444.
17. Felice DL, Sun J, Liu RH. A modified methylene blue assay for accurate cell counting. *J Funct Foods* 2009; 1(1):109-118.
18. Wolfe KL, Liu RH. Cellular antioxidant activity (CAA) assay for assessing antioxidants, foods, and dietary supplements. *J Agr Food Chem* 2007; 55(22): 8896-8907.
19. Hsieh CL, Lin YC, Ko WS, Peng CH, Huang CN, Peng RY. Inhibitory effect of some selected nutraceutical herbs on LDL glycation induced by glucose and glyoxal. *J Ethnopharmacol* 2005; 102(3):357-363.
20. Chen H, Lin Y, Hsieh C. Evaluation of antioxidant activity of aqueous extract of some selected nutraceutical herbs. *Food Chem* 2007; 104(4): 1418-1424.
21. Ames BN, Shigenaga MK, Hagen TM. Oxidants, antioxidants, and the degenerative diseases of aging. *P Natl Acad Sci Usa* 1993; 90(17): 7915-7922.
22. Adom KK, Liu RH. Rapid peroxy radical scavenging capacity (PSC) assay for assessing both hydrophilic and lipophilic antioxidants. *J Agr Food Chem* 2005; 53(17):6572-6580.
23. Fu YJ, Kadioglu O, Wiench B, Wei ZF et al. Cell cycle arrest and induction of apoptosis by cajanin stilbene acid from *Cajanus cajan* in breast cancer cells. *Phytomedicine* 2015; 22(4): 462 - 468.
24. Yin Y, Li W, Son Y, Sun L, Lu J, Kim D, Wang X, Yao H, Wang L, Pratheeshkumar P, Hitron AJ, Luo J, Gao N, Shi X, Zhang Z. Quercitrin protects skin from UVB-induced oxidative damage. *Toxicol Appl Pharm* 2013; 269(2): 89-99.
25. Morand C, Manach C, Crespy V, Remesy C. Quercetin 3-O-beta-glucoside is better absorbed than other quercetin forms and is not present in rat plasma. *Free Radical Res* 2000; 33(5): 667-676.
26. Aruna R, Geetha A, Suguna P. Expression of caspase activation recruitment and pyrin domain levels of apoptosis-associated speck-like protein complex in the pancreas of rats subjected to experimental pancreatitis: Influence of rutin administration. *Hum Exp Toxicol* 2014; 33(9): 940-948.
27. Liao J, Hsu C, Wang M, Hsu W, Chan Y. Beneficial effect of *Toona sinensis* Roemor on improving cognitive performance and brain degeneration in senescence-accelerated mice. *Brit J Nutr* 2006; 96(2): 400-407.
28. Chang H, Hsu H, Su J, Wang P, Chung Y, Chia Y, Tsai L, Wu Y, Yuan SF. The fractionated *Toona sinensis* leaf extract induces apoptosis of human ovarian cancer cells and inhibits tumor growth in a murine xenograft model. *Gynecol Oncol* 2006; 102(2): 309-314.
29. Liao J, Chung Y, Yeh J, Lin Y, Lin Y, Wu S, Chan Y. Safety evaluation of water extracts of *Toona sinensis* Roemor leaf. *Food Chem Toxicol* 2007; 45(8): 1393 - 1399.