Tropical Journal of Pharmaceutical Research June 2016; 15 (6): 1151-1157 ISSN: 1596-5996 (print); 1596-9827 (electronic) © Pharmacotherapy Group, Faculty of Pharmacy, University of Benin, Benin City, 30001 Nigeria. All rights reserved.

> Available online at http://www.tjpr.org http://dx.doi.org/10.4314/tjpr.v15i6.6

# **Original Research Article**

# Licochalcone A exerts antitumor activity in bladder cancer cell lines and mice models

Yue-Yang Zhang<sup>1</sup>, Chang-Ting Huang<sup>2</sup>, Shao-Ming Liu<sup>3\*</sup>, Bin Wang<sup>4</sup>, Jun Guo<sup>5</sup>, Jian-Qi Bai<sup>6</sup>, Xiao-Jing Fan<sup>7</sup> and Yu-Sen Jia<sup>8</sup>

<sup>1</sup>Department of Andrology, <sup>2</sup>Department of Pediatrics, Wangjing Hospital of China Academy of Chinese Medical Sciences, Beijing 100102, <sup>3</sup>Department of Urology, Beijing University of Chinese Medicine Third Affiliated Hospital, Beijing 100029, <sup>4</sup>Department of Andrology, Dongzhimen Hospital, Beijing University of Chinese Medicine, Beijing 100700, <sup>5</sup>Department of Andrology, Xiyuan Hospital of China Academy of Chinese Medical Sciences, Beijing, 100091, <sup>6</sup>Department of Pathology, Wangjing Hospital of China Academy of Chinese Medical Sciences, Beijing 100102, <sup>7</sup>Department of Acupuncture, Second Affiliated Hospital of Tianjin University of TCM, Tianjin, 300150, <sup>6</sup>Department of Urology, Dongfang Hospital of Beijing University of Chinese Medicine, Beijing, 100078, China

\*For correspondence: Email: liushaoming59@gmail.com; Tel/Fax: 0086-010-52075325

Received: 15 July 2015

Revised accepted: 9 May 2016

## Abstract

**Purpose:** To investigate the effect of licochalcone A (LA) on the inhibition of cell proliferation and ERK1/2 phosphorylation in bladder carcinoma cell lines.

**Methods:** Cell viability was investigated using 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazoliumbromide (MTT) assay. Dye-binding method was used to examine the concentration of proteins. Lymphocytes were extracted from mice and after surface staining were subjected to BD fixation and permeabilization for intracellular staining. Flow cytometry was used to measure cellular fluorescence.

**Results:** MTT results revealed a significant decrease in the proliferation of UM-UC-3, J82 and HT-1197 cell lines on treatment with LA. LA also induced reduction in phosphorylation of ERK1/2 in all three carcinoma cell lines. In the mouse model, licochalcone A treatment via intraperitoneal (ip) administration induced a significant decrease in the level of regulatory T cells (Tregs). Comparison of the mouse interferon- $\alpha$  (IFN- $\alpha$ )-treated and LA-treated groups revealed that LA treatment caused enhancement of cytotoxic T lymphocyte (CTL) activity similar to that of IFN- $\alpha$ . Administration of UM-UC-3 cells in C3H/HeN mice resulted in marked reduction in the counts for splenocytes and CD4+ CD25+ Foxp3+ T (regulatory T cells) cell proportion in LA-treated mice compared to untreated control group.

**Conclusion:** Licochalcone A may be of therapeutic importance for the prevention of bladder carcinoma. However, studies are required to ascertain the compound's usefulness in this regard.

*Keywords:* Licochalcone A, Bladder carcinoma, Splenocytes, Phosphorylation, Cell proliferation, Interferon

Tropical Journal of Pharmaceutical Research is indexed by Science Citation Index (SciSearch), Scopus, International Pharmaceutical Abstract, Chemical Abstracts, Embase, Index Copernicus, EBSCO, African Index Medicus, JournalSeek, Journal Citation Reports/Science Edition, Directory of Open Access Journals (DOAJ), African Journal Online, Bioline International, Open-J-Gate and Pharmacy Abstracts

#### INTRODUCTION

Cancer is the leading cause of death in economically developed countries and second leading cause of death in developing countries [1]. The tendency of cancer cells to proliferate uncontrollably, escape apoptosis and spread to other tissues makes it the most intractable disease [2]. Bladder carcinoma constitutes the most frequently detected cancer in industrialized countries [3]. It is associated with complex genetic events and may involve various molecular pathways, including the Ras-MAPK pathway [4]. Earlier reports show that RAS proteins transfer information from the receptors in cell surface to protein through different pathways [5]. It has been estimated that in 30 % of cancers RAS-RAF/MEK (mitogen extracellular kinase)/ERK (extracellular signal-related kinase) pathway is enhanced [6]. The first line of treatment for advanced renal cancers is sunitinib [7]. It is believed that suppression of tyrosine kinase can be a promising strategy for the inhibition of cancer cell proliferation. Tyrosine kinases mediate the binding of growth factors with their corresponding receptors [8].

Chalcones because of their potent biological activities have been the target of interest for the chemists as well as clinicians globally. They exhibit anti-inflammatory [9], anti-invasive [10], anti-tumour [11] and antibacterial [12] properties. Chalcone treatment causes induction of apoptosis and alters mitochondrial membrane potential in various types of cancers [13]. Licochalcone A (LA), an oxygenated chalcone was isolated from the roots of the Chinese liquorice (Glycyrrhiza uralensis). It is reported to exhibit various bioactive properties including antiparasitic, estrogenic, antimalarial and antitumor activities [14]. LA regulates cell growth and induces cell apoptosis in prostate cancer cell lines [15]. In the present study the effect of licochalcone A (LA) on bladder carcinoma in human bladder cancer cells and mice model was investigated. The results revealed that licochalcone A treatment inhibits bladder cancer proliferation and reduces **ERK1/2** cell phosphorylation. Furthermore, licochalcone A treatment caused a marked reduction in the regulatory T cell count in the mouse model. Also the cytotoxic T lymphocyte (CTL) activity was induced similar to that of IFN-α.

### **EXPERIMENTAL**

#### **Drug and chemicals**

Licochalcone A (LA) was isolated from the roots of the Chinese liquorice (*Glycyrrhiza uralensis*). The drug was dissolved in DMSO and stored at -20 °C. Mouse interferon- $\alpha$  (IFN- $\alpha$ ) was purchased from BD Pharmingen (San Diego, CA). The antibodies against ERK1/2 (Thr202/Tyr204) and  $\beta$ -actin were purchased from Santa Cruz Biotechnology (Santa Cruz, CA).

#### Animals and ethical statement

Female C3H/HeN mice (12 weeks old) and female nu/nu mice (10 weeks old) were obtained from the Laboratory Animal Center, Third Military

Medical University (Chongqing, China). The study was approved by the Ethics Committee of the Wangjing Hospital of China Academy of Chinese Medical Sciences (approval ref no. 201323765757/2014). All experimental animal procedures were conducted in compliance with State and Federal laws, standards of the US Department of Health and Human Services, and guidelines established by Tulane University Animal Care and Use Committee, accredited by Assessment Association for the and Accreditation of Laboratory Animal Care [16].

#### **Tumor cell line**

The bladder cancer cell lines, UM-UC-3, J82 and HT-1197 were obtained from The Cell Bank of Type Culture Collection of Chinese Academy of Sciences, Shanghai Institute of Cell Biology (Shanghai, China). The cells were cultured in DMEM containing antibiotics and were maintained in an incubator at 37 °C with humidified atmosphere of 5 % CO<sub>2</sub>.

#### MTT assay

Into a 96-well flat bottom micro plate (BD Falcon, Franklin, NJ) cells at a density of  $2.5 \times 10^5$  cells per well were distributed. To each of the well different concentration of licochalcone A was added and incubated for 72 h. MTT solution (10 µL) was added to each well and the plates were incubated for 4 h at 37 °C in 5 % CO<sub>2</sub> incubator. Formazan crystals were dissolved by adding 0.04 N HCl in 2-propanol (100 µL). The microplate reader (MPR-A4i; Tosoh Corporation, Tokyo, Japan) was used to measure absorption at 570 nm. All the measurements were carried out in triplicate.

#### Western blotting

The cells treated with licochalcone A were lysed and the concentration of proteins was determined by dye-binding method (Bio-Rad). The proteins were resolved on 15 % SDS-PAGE and transferred to nitrocellulose membranes. Digitonin-based subcellular fractionation cytosolic technique was used for and mitochondrial fractions. Onto DS-PAGE equal volumes of cytosolic and mitochondrial fractions were resolved and transferred to nitrocellulose membranes. The membranes after incubation with primary antibody were washed, and then incubated with horseradish peroxidase antimouse or horseradish peroxidase antirabbit antibodies. chemiluminescence Enhanced system used for visualization of was immunoreactive bands.

#### **Intracellular FACS**

The lymphocytes extracted from the mice after surface staining were subjected to BD fixation and permeabilization (BD Biosciences) for intracellular staining using the manual protocol. After incubation with 100 μL of BD Cytofix/Cytoperm solution the cells were washed with BD perm-wash buffer and stained with FITCconjugated anti-Foxp3 mAb (FJK-16s). FACS Calibur flow cytometer (BD Biosciences, San Jose, CA) was used to measure the cellular fluorescence.

#### **Treatment strategy**

UM-UC-3 cells at a density of  $2.5 \times 10^7$  were administered subcutaneously into the lateral flanks of the mice. When tumor achieved palpable stage, the mice in the treatment and control group were given licochalcone A (40 mg/kg) or phosphate buffered saline (PBS), respectively through the intravenous route for 4 weeks. After every 2 days the size of the tumor was measured with calipers and converted to volume.

#### **Determination of CTL activity**

The mice administered UM-UC-3 cells were sacrificed on day 14 following treatment with licochalcone A or PBS. The splenocytes were

extracted and cultured with mitomycin C (MMC)treated UM-UC-3 cells for 5 days after adding 25 ng/mL IL-2 (Takeda Chemical Industries, Tokyo, Japan). The Percoll (Amersham Biosciences, Piscataway, NJ) density gradient centrifugation was used to separate the viable cells whereas the effector cells were incubated with 51Crlabeled UM-UC-3 cells for 6 h. After incubation the cytotoxicity was evaluated.

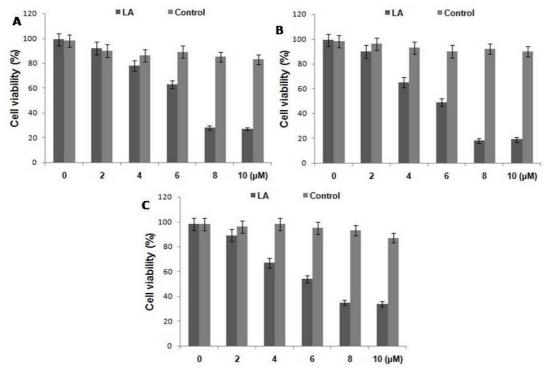
#### **Statistical analysis**

All the experiments were performed in triplicates and Student's t test was used for the determination of statistical significance. Differences were considered statistically significant at p < 0.05. Graph Pad Prism 5 (Graph Pad Software, Inc., San Diego, CA, USA) was used for the analysis of the data.

## RESULTS

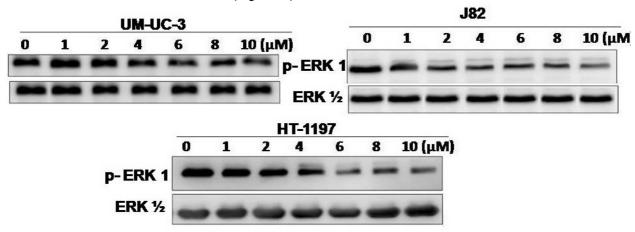
# Effect of licochalcone A on bladder cancer cell growth and ERK1/2 expression

The effect of licochalcone A on the growth of UM-UC-3, J82 and HT-1197 bladder cancer cells was analyzed. It was observed that licochalcone A suppressed the growth of all the three tested bladder cancer cell lines in a dose-dependent manner (Figure 1).



**Figure 1:** Inhibition of growth by licochalcone A in bladder cancer cell lines, UM-UC-3, J82 and HT-1197 cells. The cells were treated with various concentrations of licochalcone A for 24 h to measure the cell survival rates in comparison to control cells

Licochalcone A treatment also caused decrease in the phosphorylation of ERK1/2 in all the three tested bladder carcinoma cell lines (Figure 2). ERK 1/2 has been reported to be involved in drug resistance [11-17].



**Figure 2:** Down regulation of ERK1/2 phosphorylation by licochalcone A. Licochalcone A was added to media containing, UM-UC-3, J82 and HT-1197 cells at the concentrations and durations indicated. Whole-cell extracts were subjected to SDS-PAGE. Western blotting was performed using the antibodies indicated

# Antitumor effect of licochalcone A in nude mouse model

UM-UC-3, J82 and HT-1197 at a density of  $2.5 \times 10^6$  was administered to each of the mice. After the tumor attained palpable stage, the animal in the treatment and control groups were given licochalcone A (40 mg/kg) and saline, respectively. Analysis of the animals revealed that tumor volumes in control group were markedly higher compared to treatment group (Figure 3).

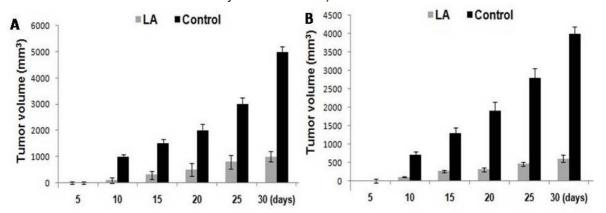
# Antitumor effect of licochalcone A in the C3H/HeN mouse model

The mice were treated with 40 mg/kg doses of the licochalcone A on the day 4 after

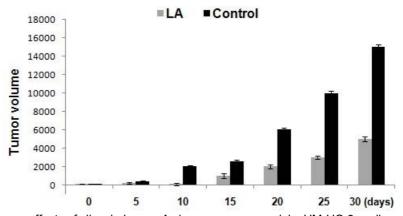
administration of  $2.5 \times 10^5$  UM-UC-3 cells. Tumor growth was significantly (p < 0.005) reduced in the licochalcone A treated mice compared with those in the control group (Figure 4).

# Effect of licochalcone A on T cells and generation of tumor-specific CTL activity in splenocytes of tumor-bearing C3H/HeN mice

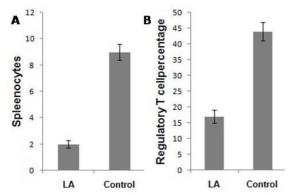
The effect of licochalcone A was also examined on effector cells for anti-tumor activity using flow cytometry on day 21 after administration of UM-UC-3cells in C3H/HeN mice. The results showed a marked reduction in the counts for splenocytes and CD4+ CD25+ Foxp3+ T (regulatory T cells) cell proportion in the licochalcone A treated mice compared to untreated control group (Figure 5A, B).



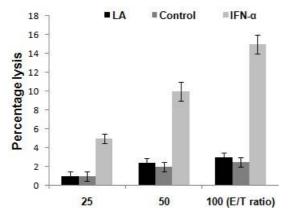
**Figure 3.** Antitumor effects of licochalcone A in the nude mouse model. UM-UC-3, J82 or HT-1197 cells ( $2.5 \times 10^6$ ) were inoculated subcutaneously into the lateral flanks of mice. The animals in the treatment and control groups were treated with licochalcone A (40 mg/kg) and PBS, respectively for 4 weeks. The size of tumor was measured using calipers after every 2 days



**Figure 4:** Anti-tumor effect of licochalcone A in a mouse model. UM-UC-3 cells were administered subcutaneously into the lateral flanks of mice. When tumor attained palpable stage the animals were assigned to two groups, treatment and control groups. The treatment and control groups were given licochalcone A (40 mg/kg) and PBS, respectively for 4 weeks daily. The size of tumor was measured after every 2 days by calipers



**Figure 5:** Effects of licochalcone A on the proportion of lymphocytes and the number of splenocytes. The number of splenocytes was examined by a counting chamber (A) and a FACS Calibur flow cytometer at 32 days after tumor inoculation. (B) Representative FACS patterns of CD4+ CD25+ Foxp3+ after either licochalcone A or PBS treatment



**Figure 6:** Effect of licochalcone A and IFN- $\alpha$  on generation of tumor-specific CTL activity in spleen cells from tumor-bearing mice. After 2 weeks of the treatment mice were sacrificed to extract spleen cells and used for the assay of CTL activity. E:T ratio, effector cells : target cells ratio; CTL, cytotoxic T lymphocyte, LA, licochalcone A

The effect of licochalcone A on the level of CTL activity was also investigated and compared with IFN- $\alpha$ -treated mouse. It was observed that the level of CTL activity was significantly higher in IFN- $\alpha$ -treated mouse compared to licochalcone A treated mouse (Figure 6).

#### DISCUSSION

Bladder carcinoma constitutes the most frequently detected cancer in industrialized countries and is associated with complex genetic events involving various molecular pathways, including the Ras-MAPK pathway [3,4]. In the present study use of licochalcone A in bladder carcinoma treatment was investigated. The results revealed that licochalcone A significantly proliferation suppressed the of bladder carcinoma cells and inhibited the tumor growth in mouse models. It is well known that ERK1/2 belonging to MAPK pathway plays a crucial role for the growth and survival of carcinoma cells [17]. It also induces resistance to various drugs used against cancers [18,19]. Therefore, it is believed that inhibition of ERK1/2 signaling pathway can be of great therapeutic importance for cancer treatment. The results from the present study showed that licochalcone A treatment inhibited phosphorylation of ERK1/2.

In patients suffering from chronic myelogenous leukemia the CTL activity is promoted on treatment with IFN- $\alpha$  [20,21]. The results of the present study revealed that licochalcone A treatment proved more effective for the improvement of CTL activity in the spleen compared to IFN- $\alpha$ . The CD4+CD25+Foxp3+ Tregs cell count in the licochalcone A treated group was significantly decreased compared to untreated group. Tregs have been shown to

produce immunosuppressive cytokines as well as negative regulation of immunity [22]. Therefore, reduction in Tregs will promote immunity and enhance activity of the T-cells. In the licochalcone A treated mice the activity of CTL was significantly increased than those of untreated control mice. In addition the splenocyte count was also decreased in licochalcone A treated mice.

### CONCLUSION

The results obtained show that licochalcone A has an anti-tumor effect in murine models of bladder cancer when used as a single agent. Furthermore, the findings suggest that licochalcone may enhance the therapeutic efficacy of existing immune-based therapies for metastatic cancer patients.

### DECLARATIONS

#### **Conflict of Interest**

No conflict of interest associated with this work.

#### **Contribution of Authors**

The authors declare that this work was done by the authors named in this article and all liabilities pertaining to claims relating to the content of this article will be borne by them.

#### REFERENCES

- Jemal A, Bray F, Center MM, Ferlay J, Ward E, Forman D. Global cancer statistics. CA Cancer J Clin 2011; 61: 69-90.
- Hiss DC, Gabriels GA. Implications of endoplasmic reticulum stress, the unfolded protein response and apoptosis for molecular cancer therapy. Part I: targeting p53, Mdm2, GADD153/CHOP, GRP78/BiP and heat shock proteins. Expert Opin. Drug Discov 2009; 4: 799-821.
- 3. Jemal A, Siegel R, Ward E. Cancer statistics, 2008. CA Cancer J Clin 2008; 58: 71-96.
- Zachos I, Konstantinopoulos PA, Tzortzis V. Systemic therapy of metastatic bladder cancer in the molecular era: current status and future promise. Expert Opin Investig Drugs 2010; 19: 875-887.
- Sridhar SS, Hedley D, Siu LL. Raf kinase as a target for anticancer therapeutics. Mol Cancer Ther 2005; 4: 677-685.
- Gollob JA, Wilhelm S, Carter C. Role of Raf kinase in cancer: therapeutic potential of targeting the Raf/MEK/ERK signal transduction pathway. Semin Oncol 2006; 33: 392-406.

- Mendel DB, Laird AD, Xin X. In vivo antitumor activity of SU11248, a novel tyrosine kinase inhibitor targeting vascular endo¬thelial growth factor and platelet-derived growth factor receptors: determination of a pharmacokinetic/pharmacodynamic relationship. Clin Cancer Res 2003; 9: 327-337.
- Cohen HT, McGovern FJ. Renal-cell carcinoma. N Engl J Med 2005; 353: 2477-2490.
- Nowakowska Z. A review of anti-infective and antiinflammatory chalcones. Eur J Med Chem 2007; 42: 125-137.
- Go ML, Wu X, Liu XL. Chalcones: An update on cytotoxic and chemoprotective properties. Curr Med Chem 2005; 12: 483-499.
- Kumar D, Kumar NM, Akamatsu K, Kusaka E, Harada H, Ito T. Synthesis and biological evaluation of indolyl chalcones as antitumor agents. Bioorg Med Chem Lett 2011; 20: 3916-3919.
- Sivakumar PM, Ganesan S, Veluchamy P, Doble M. Novel chalcones and 1, 3, 5-triphenyl-2-pyrazoline derivatives as antibacterial agents. Chem Biol Drug Des 2010; 76: 407-411.
- Sakai T, Eskander RN, Guo Y,Kim KJ, Mefford J, Hopkins J, Bhatia NN, Zi X, Hoang BH. Flavokawain B, a kava chalcone, induces apoptosis in synovial sarcoma cell lines. J Orthop Res 2011; 30: 1045-1050.
- Chimenti F, Fioravanti R, Bolasco A, Chimenti P, Secci D, Rossi F. Chalcones: A valid scaffold for monoamine oxidases inhibitors. J Med Chem 2009; 52: 2818-2824.
- Lust S, Vanhoecke B, Janssens A, Philippe J, Bracke M, Offner F. Xanthohumol kills B-chronic lymphocytic leukemia cells by an apoptotic mechanism. Mol Nutr Food Res 2005; 49: 844-850.
- Lyndsay V. R, Syreeta L. T, Stephen M. B, Shuchen W, HafezA K, Shannon E. M, Melyssa R. B, Qiang Z, Guangdi W, Matthew E. B, Bridgette M. C-B. Glyceollins as novel targeted therapeutic for the treatment of triple-negative breast cancer. Oncology lett. 2012; 3: 163-171.
- Zachos I, Konstantinopoulos PA, Tzortzis V. Systemic therapy of metastatic bladder cancer in the molecular era: current status and future promise. Expert Opin Investig Drugs 2010; 19: 875-887.
- Jeong EK, Lee SY, Jeon HM. Role of extracellular signalregulated kinase (ERK) 1/2 in multicellular resistance to docetaxel in MCF-7 cells. Int J Oncol 2010; 37: 655-661.
- Peng DJ, Wang J, Zhou JY. Role of the Akt/mTOR survival pathway in cisplatin resistance in ovarian cancer cells. Biochem Biophys Res Commun 2010; 394: 600-605.
- Frumento G, Piazza T, Di Carlo E. Targeting tumorrelated immunosuppression for cancer immunotherapy. Endocr Metab Immune Disord Drug Targets 2006; 6: 233-237.
- Molldrem JJ, Lee PP, Wang C. Evidence that specific T lymphocytes may participate in the elimination of chronic myelo-genous leukemia. Nat Med 2000; 6: 1018-1023.

22. Von Boehmer H. Mechanisms of suppression by

suppressor T cells. Nat Immunol 2005; 6: 338-344.