Tropical Journal of Pharmaceutical Research November 2017; 16 (11): 2629-2635 ISSN: 1596-5996 (print); 1596-9827 (electronic) © Pharmacotherapy Group, Faculty of Pharmacy, University of Benin, Benin City, 300001 Nigeria. All rights reserved.

> Available online at http://www.tjpr.org http://dx.doi.org/10.4314/tjpr.v16i11.9

Original Research Article

Effect of natural antioxidants on the aggregation and disaggregation of beta-amyloid

Jae-Eun Lee, Min-Suk Kim and So-Young Park*

Laboratory of Phramacognosy, College of Pharmacy, Dankook University, Cheonan 330-714, Korea

*For correspondence: Email: soypark23@dankook.ac.kr; Tel: +82 41 550 1434; Fax: +82 559 7899

Sent for review: 9 July 2017

Revised accepted: 16 October 2017

Abstract

Purpose: To examine the relationship between higher antioxidant activity and aggregation or disaggregation of beta-amyloid ($A\beta$) for 21 plants.

Methods: Twenty-nine natural plant extracts and their antioxidant activities were analyzed using DPPH assay. The aggregation and disaggregation of $A\beta$ were analyzed using Thioflavin-T assay.

Results: Eleven plant extracts exhibited high antioxidant activities with the half-maximal inhibitory concentration (IC_{50}) values < 20.0 µg/mL. Furthermore, the plant extracts efficiently inhibited A β aggregation with a mean IC_{50} value of 17.0 µg/mL. However, four plant extracts exhibiting low antioxidant activities (IC_{50} > 80.0 µg/mL) inhibited A β aggregation less efficiently with a mean IC_{50} value of 75.7 µg/mL. Furthermore, plant extracts with high antioxidant activities were not invariably efficient for disaggregating pre-formed A β aggregates.

Conclusion: High antioxidant activities were positively correlated with the inhibition of $A\beta$ aggregation, although not with the disaggregation of pre-formed $A\beta$ aggregates. Nevertheless, potent antioxidants may be helpful in treating Alzheimer's disease.

Keywords: Alzheimer's disease, β-Amyloid, Aggregation, Disaggregation, Antioxidants

Tropical Journal of Pharmaceutical Research is indexed by Science Citation Index (SciSearch), Scopus, International Pharmaceutical Abstract, Chemical Abstracts, Embase, Index Copernicus, EBSCO, African Index Medicus, JournalSeek, Journal Citation Reports/Science Edition, Directory of Open Access Journals (DOAJ), African Journal Online, Bioline International, Open-J-Gate and Pharmacy Abstracts

INTRODUCTION

 β -amyloid (A β) is a polypeptide of 36-43 amino acids, and is a major causative factor of Alzheimer's disease (AD). A β aggregation into senile plaques is a pathological characteristic detected in the brains of patients with AD. A β is produced by the sequential cleavage of amyloid precursor proteins by β - and γ -secretases. A β molecules undergo self-aggregation to form oligomers and fibrils, which cause neurotoxicity via oxidative stress, disturb calcium homeostasis, and induce mitochondrial dysfunction, and neuroinflammation. Thus, the discovery of compounds or extracts that inhibit A β aggregation, and induce the disaggregation of A β aggregates could be an effective approach to the therapy and prevention of AD.

Antioxidants are molecules that scavenge free radicals and reactive oxygen species (ROS), can induce oxidative stress. which and consequently, cellular damages. Antioxidant intake is helpful in preventing human diseases [1]. For example, antioxidants can reduce the risk of cardiovascular diseases by inhibiting vascular inflammation, lipid peroxidation, and platelet aggregation [2]. Additionally, antioxidants are known to have anti-cancer, anti-diabetic, and anti-aging effects [3-5]. Antioxidants are also considered helpful in treating AD, because the ROS generated in AD pathology causes neuronal damage [6]. Furthermore, certain antioxidants such as polyphenols inhibit $A\beta$ aggregation, and destabilize preformed $A\beta$ aggregates [7,8]. However, the relationships between antioxidant activity and inhibition of $A\beta$ aggregation, and between antioxidant activity and disaggregation of preformed $A\beta$ aggregates are not analyzed. Therefore, in this study, 29 natural plant extracts with diverse antioxidant activity were analyzed for effects on the aggregation and disaggregation of $A\beta$, and the correlation between them was analyzed as well.

EXPERIMENTAL

Materials and reagents

Thioflavin T (ThT), DPPH, and dimethyl sulfoxide (DMSO) were purchased from Sigma-Aldrich (MO, USA). Ethanol and methanol were purchased from Samchun (Pyeong-Taek, Korea). $A\beta_{1-42}$ was purchased from GL Biochem (Shanghai, China).

Plant materials

Twenty-nine plant materials (Table 1) were purchased from a commercial market (Samhing medicinal herb market; Seoul, Korea) in 2014. One of the authors (S-Y Park) performed botanical identification and voucher specimens were deposited in Pharmacognosy laboratory of College of Pharmacy, Dankook University.

Preparation of plant extracts

Dried and pulverized plant materials were extracted 3 times with 90 % methanol at room temperature. The MeOH filtrate was evaporated under vacuum to yield the MeOH extract. Each extract was dissolved in DMSO and stored at -20 °C until use for the assay.

Antioxidant assay

The antioxidant activities of the plant extracts were determined using a stable free radical, DPPH. The ability of the plant extracts to scavenge DPPH, and convert it to 1,1-diphenyl-2-picrylhydrazine was determined colorimetrically. In brief, the plant extracts at different concentrations of (4, 20, and 100 µg/mL) were mixed with 190 µL of DPPH (0.316 mM in ethanol) and the mixtures were incubated at 37 °C for 30 min. The optical density values of the mixtures were measured at 517 nm with a microplate reader (Molecular Devices. Sunnyvale, CA, USA). All measurements were conducted out at least thrice.

ThT assay of inhibition of Aβ aggregation

To quantify A β aggregates, the ThT assay was performed. A β_{1-42} was dissolved in DMSO at 1 mL, and stored at -20 °C until use. To monitor their effects on A β aggregation, the plant extracts (4, 20, and 100 µg/mL) were incubated with 20 µM of A β (49 µL) at 37 °C for 24 h. Then, 3 µM of ThT (50 µL) was added, and the fluorescence was measured after 30 min with an EMax precision microplate reader (Molecular Devices) with excitation at 442 nm and emission at 485 nm. A β treated with DMSO was used as the control, and each assay was performed in triplicate.

ThT assay of disaggregation of preformed A β aggregates

To monitor the effects of the plant extracts on the disaggregation of the A β aggregates, 20 μ M of A β (49 μ L) was incubated at 37 °C. After 24 h, the plant extracts (4, 20, and 100 μ g/mL) were added, and incubated for an additional 24 h. Subsequently, 3 μ M of ThT (50 μ L) was added, and the fluorescence was measured as described in the previous section.

Statistical analysis

Data were expressed as means \pm SD. Two or more group comparisons were performed by one-way analysis of variance (ANOVA), followed by Tukey's post-hoc test (SPSS version 17.0; IBM SPSS, Armonk, NY, USA). Differences were considered statistically significant at *P* < 0.05.

RESULTS

Antioxidant effects of the plant extracts

The results of DPPH assay showed that Agrimonia pilosa, Cornus officinalis, Rubus coreanus, Sophora japonica, and Paeonia suffruticosa exhibited strong antioxidant effects with IC_{50} values below than 10 µg/mL. Contrastingly, Houttuynia cordata, Torilis japonica, Rehmannia glutinosa, and Arisaema erubescens exhibited relatively poor antioxidant activities with IC_{50} values higher than 100 µg/mL (Table 1).

Inhibition of $A\beta$ aggregation by the plant extracts

The ThT assay demonstrated that Cornus officinalis exhibited the highest inhibitory effect on A β aggregation with an IC₅₀ value of 8.2 µg/mL (Table 1).

| Plant | Plant part used | Antioxidant effect (IC₅₀, μg/mL) | Inhibition of Aβ aggregation (IC ₅₀ , μg/mL) | Disaggregation of Aβ aggregates (ED₅₀, µg/mL) |
|--|-----------------|--|---|--|
| <i>Agrimonia pilosa</i> Ledebour (Rosaceae) | Whole plant | 5.1 | 9.1 | 102.3 |
| <i>Amomum villosum</i> Loureiro (Zingiberaceae) | Fruit/Seed | 17 | 13.8 | 39.9 |
| Arisaema erubescens Schott (Araceae) | Root | 90.9 | 98.3 | 114.2 |
| <i>Chelidonium majus</i> Linné var. asiaticum Ohwi (Papaveraceae) | Aerial parts | 78.2 | 72.4 | 104.2 |
| <i>Chrysanthemum zawadskii</i> Herbich var. latilobum (Maxim.) Kitamura (Compositae) | Whole plant | 27.8 | 46.4 | 179.7 |
| <i>Cimicifuga heracleifolia</i> Komarov (Ranunculaceae) | Rhizome | 52.7 | 76.1 | 115.7 |
| <i>Cornus officinalis</i> Siebold et Zuccarini (Cornaceae) | Fruit | 5.3 | 8.2 | 42.1 |
| Cyperus rotundus L (Cyperaceae) | Rhizome | 26 | 15.2 | 42.9 |
| <i>Drynaria fortunei</i> J. Smith (Polypodiaceae) | Rhizome | 19.7 | 16.1 | 134.9 |
| <i>Epimedium koreanum</i> Nakai (Berberidaceae) | Aerial parts | 46.8 | 39.4 | 110.6 |
| <i>Eucommia ulmoid</i> es Oliver (Eucommiaceae) | Leaves | 72.5 | 56.1 | 234.1 |
| <i>Houttuynia cordata</i> Thunberg (Saururaceae) | Aerial parts | 134.4 | 99.3 | 276.4 |
| <i>Hovenia dulci</i> s Thunb (Rhamnaceae) | Fruit/Seed | 31.4 | 46.6 | 124.6 |
| <i>Leonurus japonicus</i> Houttuyn (Labiatae) | Aerial parts | 64.1 | 47.9 | 25.7 |
| <i>Magnolia obovata</i> Thunberg (Magnoliaceae) | Bark | 30 | 49.3 | 130.3 |
| Morus alba Linne (Moraceae) | Leaves | 74.2 | 44.5 | 73.8 |
| <i>Myristica fragrans</i> Houttuyu (Myristicaceae) | Seed | 59 | 51.3 | 88.4 |
| <i>Paeonia lactiflora</i> Pall (Ranuculaceae) | Root | 10 | 24.4 | 48.2 |
| <i>Paeonia suffruticosa</i> Andrews (Paeoniaceae) | Root bark | 7.8 | 20.8 | 92.9 |
| <i>plantago asiatica</i> L (Plantaginacea) | Seed | 12.8 | 20.6 | 127.7 |
| <i>Prunella vulgari</i> s L (Labiatae) | Stem | 14.6 | 24.8 | 45 |
| <i>Rehmannia glutinosa</i> Liboschitz ex Steudel (Scrophulariaceae) | Root | 109.5 | 80.2 | 396.9 |
| Rubus coreanus Miquel (Rosaceae) | Fruit | 6 | 12.8 | 146.1 |
| <i>Schizonepeta tenuifolia</i> Briquet (Labiatae) | Flower | 29.8 | 32.1 | 57.7 |
| Scrophularia buergeriana Miquel (Scrophulariaceae) | Root | 53.8 | 72.8 | 67.4 |
| Sophora japonica L (Leguminosae) | Flower | 7.1 | 25.2 | 134.3 |
| <i>Thuja orientalis</i> L (Curpressaceae) | Leaves | 16.7 | 10.8 | 14.7 |
| <i>Torilis japonica</i> Decandolle (Umbelliferae) | Fruit | 100.9 | 87.5 | 377.6 |
| Zingiber officinale Roscoe | Rhizome | 65.7 | 47.2 | 137.4 |

(Zingiberaceae)

Table 1: List of some plant species, and their antioxidant, anti-amyloidogenic and Aß disaggregating activities

Aβ incubation with 100 µg/mL C. officinalis reduced the extent of A β aggregation to 23 %, compared with the DMSO-treated control groups (Figure 1A). Similarly, Thuja orientalis and Agrimonia pilosa had strong inhibitory effects on A β aggregation with IC₅₀ values of 9.1 and 10.8 µg/mL (Table 1), respectively. T. orientalis and A. pilosa at 100 µg/mL reduced Aβ aggregation to 16.5 % and 26.2 %, respectively, compared with the DMSO-treated control groups (Figure 1A). However, Torilis japonica, Arisaema erubescens, and Houttuynia cordata were less efficient in inhibiting AB aggregation, and their IC_{50} values were 87.5, 98.3 and 99.3, respectively (Table 1). Incubation with 100 µg/mL T. japonica, A. erubescens, and H. cordata reduced AB aggregation to 47.2 %, 49.9 %, and 43.6 %., whereas these plant extracts at 4 µg/mL exhibited negligible inhibitory effects on AB aggregation (Figure 1B).

Effects of the plant extracts on disaggregation of A β aggregates

The disaggregating effects of the 29 plant extracts on pre-aggregated A β aggregates were determined by the ThT assay. *Thuja orientalis* exhibited the highest disaggregating activity among the 29 plant extracts with an IC₅₀ value of 14.7 µg/mL (Table 1). Similarly, *Leonurus japonicus* efficiently induced the disaggregation of preformed A β aggregates with an IC₅₀ value of 25.7 µg/mL. Contrarily, the effects of *Torilis japonica* and *Rehmannia glutinosa* on the disaggregation of preformed A β aggregates were extremely minimal with IC₅₀ values of 377.6 and 396.9 µg/mL, respectively.

Antioxidant activity and inhibition of $A\beta$ aggregation

To determine the relationship between the inhibition of A β aggregation and antioxidant activity, twenty-nine plants were divided into 4 groups based on the IC₅₀ values for antioxidant activity (group 1, IC₅₀ value: 0–20; group 2, IC₅₀

value: 21-50; group 3, IC₅₀ value: 51-80, and group 4, IC_{50} value over 81). The average IC_{50} values for the antioxidant activities of the groups 1, 2, 3, and 4 were 11.1, 32.0, 65.0, and 86.1 μ g/mL, respectively (Figure 2A), whereas the average IC_{50} values for anti- A β aggregation in the groups 1, 2, 3, and 4 were 17.0, 38.2, 58.5 and 75.7 μ g/mL, respectively. The average IC₅₀ value for anti- Aß aggregating activities of the plant extracts in group 1 was significantly lower than those of the groups 2, 3, and 4. These results indicated that the plant extracts with higher antioxidant activities exhibited higher anti-Aβ aggregating activities. Therefore, plants with higher antioxidant activities could be beneficial for patients with AD because of their possibly higher anti-Aß aggregating activities.

Antioxidant activity and disaggregation of $A\beta$ aggregates

То determine the relationship between antioxidant activity and the disaggregation of $A\beta$ aggregates, the mean IC₅₀ values of groups 1-4 for the disaggregation of AB aggregates were compared (Figure 2C). The average IC₅₀ values of the groups 1, 2, 3, and 4 for A β disaggregation were 84.3, 107.6, 105.8, and 291.2 µg/mL, respectively. Group 1 exhibited a relatively low IC_{50} value for A β disaggregation; however, it was not significantly different from those of groups 2 and 3. However, the disaggregating activities of groups 1, 2, and 3 were significantly different from that of group 4.

DISCUSSION

Natural products containing polyphenolic compounds, such as berries, spices, and green tea, are known to have antioxidant and neuroprotective activities. Furthermore, the neuroprotective effects of natural polyphenolic compounds, such as myricetin and luteolin are exerted, at least partially, by attenuating Aβ-induced toxicity [9].





Trop J Pharm Res, November 2017; 16(11): 2632

Lee et al



Figure 2: Mean values of the antioxidant, anti-A β aggregating and A β -disaggregating activities of 29 plant extracts divided into 4 groups based on antioxidant activity. (A) Average values of antioxidant activities, (B) average values of anti-A β aggregating activities and (C) average values of A β -disaggregating activities. ^a P < 0.05, different from Group 1, ^b P < 0.05, different from Group 2, ^c P < 0.05, different from Group 3, ^d P < 0.05, different from Group 4

However, this does not indicate that potent antioxidants are efficient in the inhibition of $A\beta$ aggregation and disaggregation of $A\beta$ aggregates. Therefore, in this study, the relationships between antioxidant activity and inhibition of $A\beta$ aggregation, and between antioxidant activity and disaggregation of preformed $A\beta$ aggregates were examined.

The aggregation of $A\beta$ monomers to form oligomers and fibrils (aggregates) induces neurotoxicity. Oxidative stress, induced during Aβ aggregation, disrupts synapses, and impairs the functions of synapses and membranes [10,11]. Thus, antioxidants could be helpful in treating AD by reducing oxidative stress-related neuronal damage. Additionally, the use of compounds or extracts inhibiting Aß aggregation is considered a good approach to the therapy and prevention of AD. For example, flavonoids such as myricetin, quercetin, and kaempferol inhibited AB aggregation (exhibited antiamyloidogenic activities) [12-14]. Additionally, rosmarinic acid and curcumin are other compounds that inhibit Aß aggregation efficiently [15,16]. Interestingly, these compounds are known to have high antioxidant activities. Consistent with previous reports, our results suggested that the plant extracts with relatively high antioxidant activity inhibited Aß aggregation efficiently. This could be because potent antioxidants, such as polyphenols, in the plant

extracts interact with aromatic regions in A β , disrupt the self-assembly of A β into β -sheet conformational stacking, and eventually, inhibit A β aggregation [17].

Antioxidants are also considered efficient in disaggregating AB. Curcumin and B-carotene exhibited high Aβ-disaggregating activities owing to their symmetric rod-like structures and hydrophobic moieties that wedged into A^β cores, and disrupt the β -sheet structures [18,19]. However, the increases in hydrophilicity reduced Aβ-disaggregating activity [18]. Furthermore, morin is an efficient Aβ-disaggregating flavonoid. The aromatic moiety and hydrogen-bonding capacity of morin are suggested to cause the disaggregation of AB aggregates [20]. These reports suggest that molecules with hydrophobicity, aromatic moiety, and hydrogen bonding-capacity might, at least, partially contribute to the disaggregation of Aβ aggregates. Consistent with these results, our study similarly suggested that antioxidants are not invariably positively correlated with high Aβdisaggregating activity. Altogether, these results suggested that the plant extracts with high antioxidant activities are efficient in inhibiting Aß aggregation; however, they were not invariably efficient in disaggregating Aβ aggregates. Nonetheless, antioxidants could be beneficial for patients with AD owing to their possible anti-Aß aggregating and Aβ-disaggregating activities.

Trop J Pharm Res, November 2017; 16(11): 2633

CONCLUSION

Twenty-nine plant extracts have been analyzed for antioxidant, anti-amyloidogenic, and $A\beta$ disaggregating activities. The plant extracts with high antioxidant activity inhibit $A\beta$ aggregation more efficiently than those with low antioxidant activity; however, antioxidant activity correlates weakly with the disaggregation of pre-formed $A\beta$ aggregates. Nevertheless, good antioxidants may be helpful in treating AD.

DECLARATIONS

Acknowledgement

This work was supported by Research Fund of Dankook University in the form of a grant to one of the authors (SY Park) in 2015.

Conflict of Interest

No conflict of interest associated with this work.

Contribution of Authors

The authors declare that this work was done by the authors named in this article and all liabilities pertaining to claims relating to the content of this article will be borne by them.

Open Access

This is an Open Access article that uses a funding model which does not charge readers or their institutions for access and distributed under the terms of the Creative Commons Attribution License (http://creativecommons.org/licenses/by/ 4.0) and the Budapest Open Access Initiative (http://www.budapestopenaccessinitiative.org/rea d), which permit unrestricted use, distribution, and reproduction in any medium, provided the original work is properly credited.

REFERENCES

- Wang J, Hu S, Nie S, Yu Q, Xie M. Reviews on Mechanisms of In Vitro Antioxidant Activity of Polysaccharides. Oxid Med Cell Longev 2016; 2016: 5692852
- 2. Adegbola P, Aderibigbe I, Hammed W, Omotayo T. Antioxidant and anti-inflammatory medicinal plants have potential role in the treatment of cardiovascular disease: a review. Am J Cardiovasc Dis 2017; 20177: 19-32
- Wu X, Cheng J, Wang X. 2017. Dietary Antioxidants: Potential Anticancer Agents. Nutr Cancer 2017; 69: 521-533

- Eid HM, Haddad PS. The Antidiabetic Potential of Quercetin: Underlying Mechanisms. Curr Med Chem 2017; 24: 355-364
- 5. Obrenovich ME, Nair NG, Beyaz A, Aliev G, Reddy VP. The role of polyphenolic antioxidants in health, disease, and aging. Rejuvenation Res 2010; 13: 631-643
- Manoharan S, Guillemin GJ, Abiramasundari RS, Essa MM, Akbar M, Akbar MD. The Role of Reactive Oxygen Species in the Pathogenesis of Alzheimer's Disease, Parkinson's Disease, and Huntington's Disease: A Mini Review. Oxid Med Cell Longev 2016; 2016: 8590578
- Xie H, Wang JR, Yau LF, Liu Y, Liu L, Han QB, Zhao Z, Jiang ZH. Catechins and procyanidins of Ginkgo biloba show potent activities towards the inhibition of betaamyloid peptide aggregation and destabilization of preformed fibrils. Molecules 2014; 19: 5119-5134
- Pasinetti GM. Novel role of red wine-derived polyphenols in the prevention of Alzheimer's disease dementia and brain pathology: experimental approaches and clinical implications. Planta Med 2012; 78: 1614-1619
- Ma H, DaSilva NA, Liu W, Nahar PP, Wei Z, Liu Y, Pham PT, Crews R, Vattem DA, Slitt AL, et al. Effects of a Standardized Phenolic-Enriched Maple Syrup Extract on beta-Amyloid Aggregation, Neuroinflammation in Microglial and Neuronal Cells, and beta-Amyloid Induced Neurotoxicity in Caenorhabditis elegans. Neurochem Res 2016; 41: 2836-2847
- 10. Mattson MP. Pathways towards and away from Alzheimer's disease. Nature 2004; 430:631-639
- Butterfield DA, Drake J, Pocernich C, Castegna A. 2001. Evidence of oxidative damage in Alzheimer's disease brain: central role for amyloid beta-peptide. Trends Mol Med 2001; 7: 548-554
- Hamaguchi T, Ono K, Murase A, Yamada M. Phenolic compounds prevent Alzheimer's pathology through different effects on the amyloid-beta aggregation pathway. Am J Pathol 2009; 175: 2557-2565
- Jimenez-Aliaga K, Bermejo-Bescos P, Benedi J, Martin-Aragon S. Quercetin and rutin exhibit antiamyloidogenic and fibril-disaggregating effects in vitro and potent antioxidant activity in APPswe cells. Life Sci 2011; 89: 939-945
- 14. Sharoar MG, Thapa A, Shahnawaz M, Ramasamy VS, Woo ER, Shin SY, Park IS. Keampferol-3-O-rhamnoside abrogates amyloid beta toxicity by modulating monomers and remodeling oligomers and fibrils to nontoxic aggregates. J Biomed Sci 2012; 19: 104
- Ono K, Hasegawa K, Naiki H, Yamada M. Curcumin has potent anti-amyloidogenic effects for Alzheimer's betaamyloid fibrils in vitro. J Neurosci Res 2004; 75: 742-750
- Yamada M, Ono K, Hamaguchi T, Noguchi-Shinohara M. Natural Phenolic Compounds as Therapeutic and Preventive Agents for Cerebral Amyloidosis. Adv Exp Med Biol 2015; 863: 79-94
- Ahmad E, Ahmad A, Singh S, Arshad M, Khan AH, Khan RH. A mechanistic approach for islet amyloid polypeptide aggregation to develop anti-amyloidogenic agents for type-2 diabetes. Biochimie 2011; 93: 793-805

Trop J Pharm Res, November 2017; 16(11): 2634

- Ono K, Yoshiike Y, Takashima A, Hasegawa K, Naiki H, Yamada M. Vitamin A exhibits potent antiamyloidogenic and fibril-destabilizing effects in vitro. Exp Neurol 2004; 189: 380-392
- 19. Das S, Stark L, Musgrave IF, Pukala T, Smid SD. Bioactive polyphenol interactions with beta amyloid: a comparison of binding modelling, effects on fibril and

aggregate formation and neuroprotective capacity. Food Funct 2016; 7: 1138-1146

 Lemkul JA, Bevan DR. Destabilizing Alzheimer's Abeta(42) protofibrils with morin: mechanistic insights from molecular dynamics simulations. Biochem 2010; 49: 3935-3946.