

Original Research Article

Anticancer Activity of Tetrahydrocorysamine against Pancreatic Adenocarcinoma Cell Line PANC-1 *In vitro* and *In vivo*

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Abstract

Purpose: To investigate the cytotoxic activity of tetrahydrocorysamine (TCSM) from *Corydalis Rhizoma* W. T. Wang (Papaveraceae) against PANC-1 cells and its possible mechanisms.

Methods: TCSM was isolated from *Corydalis Rhizoma* by column chromatography and identified by mass spectrometry and nuclear magnetic resonance (NMR). The effects of TCSM on the proliferation and apoptosis of PANC-1 cells were determined by methyl thiazolyl tetrazolium (MTT) and flow cytometry assays. The effect of TCSM on the expressions of mitochondria-mediated apoptotic proteins were investigated by Western blot assay. Xenograft assay was used to evaluate the anticancer activity of TCSM *in vivo*.

Results: TCSM showed cytotoxic activity against PANC-1 cells with half-maximal concentration (IC_{50}) of 19.16 μ M in MTT assay. TCSM (5, 10 and 20 μ M) significantly ($p < 0.01$) increased the apoptosis rates of PANC-1 cells (24.45, 35.26 and 54.16 vs 6.12 %), compared with control group. The results of Western blot suggest that TCSM significantly ($p < 0.01$) down-regulated the expressions of anti-apoptotic proteins (Bcl-2 and Survivin), up-regulated the expressions of pro-apoptotic proteins (Bax, Smac, c-caspase-3 and c-caspase-9) and promoted the release of cytochrome C from mitochondria to the cytoplasm of PANC-1 cells, compared with control. TCSM (20 mg/kg) significantly ($p < 0.01$) inhibited the growth of PANC-1 cell-induced tumor (384 vs 1138 mm³) by regulating the expressions of mitochondria-mediated apoptotic proteins as stated above for xenograft assay, compared with control.

Conclusion: TCSM induces apoptosis in PANC-1 cells *in vitro* and *in vivo* via mitochondria-mediated apoptotic pathway.

Keywords: *Corydalis Rhizoma*, Tetrahydrocorysamine, PANC-1 cells, Flow cytometry, Western blot, Xenograft

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INTRODUCTION

Currently, pancreatic adenocarcinoma is still a major unsolved health problem, and almost all pancreatic adenocarcinoma patients suffer from metastases and deaths [1]. The pancreatic adenocarcinoma is usually diagnosed in the advanced state, when there are few or no

effective therapies, so the 5-year survival rate of pancreatic adenocarcinoma patients remains poor (about 3 %) [2]. Therefore, it is urgent and important to find new, safety and effective drugs for treating pancreatic adenocarcinoma. Nowadays, finding compounds from plants are an important pathway to obtain new drugs for treating pancreatic adenocarcinoma [3].

Corydalis Rhizoma [*Corydalis yanhusuo* W. T. Wang (Papaveraceae)] is a commonly used medicine in China, and its major constituents are alkaloids such as d-corydaline, dl-tetrahydropalmatine, protopine, berberine, l-tetrahydrocoptisine, dl-tetrahydrocoptisine, l-tetrahydrocolumbamine, d-corybulbine, d-glaucine, a-allocryptopine and tetrahydrocorysamine (TCSM) [4]. It is reported that *Corydalis Rhizoma* has anticancer activity [5], and the active constituents are the alkaloids [6]. The anticancer activities of alkaloids from *Corydalis Rhizoma* have been widely reported [7,8], and the mechanisms involve apoptosis, reversal of multidrug resistance of tumor cells, anti-angiogenic effect, the expression of microRNAs, etc.

In the present study, we investigated the cytotoxic activity of TCSM isolated from *Corydalis Rhizoma* against PANC-1 cells and possible mechanisms by methyl thiazolyl tetrazolium (MTT), flow cytometry, Western blot and xenograft assays.

EXPERIMENTAL

Plant materials

Corydalis Rhizoma was purchased from Anguo Chinese herbal medicine market (Hebei, China) in 2013, identified by Gang Peng, Suizhou Hospital, Hubei University of Medicine. A voucher specimen (voucher no. 201309113) was stored in Hubei University of Medicine for future reference.

Chemicals and reagents

Silica gel (300 - 400 mesh), all analytical grade reagents including ethanol, petroleum ether, chloroform, ethyl acetate and n-butyl alcohol, acetone and preparative TLC were purchased from Qingdao Haiyang Chemical Co., Ltd. (Qingdao, China). DMEM medium and fetal bovine serums (FBS) were obtained from Invitrogen (Carlsbad, CA, USA). MTT Cell Proliferation and Cytotoxicity Assay Kits and Enhanced BCA Protein Assay Kit were obtained from Applygen (Beijing, China) and Beyotime Institute of Biotechnology (Shanghai, China). Annexin V-FITC/PI apoptosis assay kits were purchased from Ruisai biomart (Shanghai, China). Primary antibodies for β -actin, Bcl-2, Survivin, Bax, Smac, cytochrome c, cleaved caspase-3 (c-caspase-3), c-caspase-9 and horseradish peroxidase (HRP)-conjugated goat anti-rabbit antibody were purchased from Cell Signaling Technology (Beverly, MA, USA).

Animals

Nude mice (BALB/C, 5 - 6 weeks old) were purchased from the SLRC Laboratory Animal Company (Shanghai, China). All experiments were strictly in accordance with international ethical guidelines and the National Institutes of Health Guide concerning the Care and Use of Laboratory Animals [9] and carried out with the approval of the Animal Experimentation Ethics Committee of Suizhou Hospital, Hubei University of Medicine (protocol no: HBUMSZH2014).

Preparation of TCSM

The air-dried *Corydalis Rhizoma* (120 kg) was finely ground and extracted by refluxing with 85 % ethanol. The ethanol solvent was evaporated under reduced pressure to obtain a crude extract, which was then suspended in water and partitioned with petroleum ether, chloroform, ethyl acetate and n-butyl alcohol. The petroleum ether fraction (1025 g) was subjected to column chromatography over silica gel (300 - 400 mesh), eluted with petroleum ether-acetone (99:1 - 40:60) to give 31 fractions. Fraction 8 (51.2 g) was separated by column chromatography over silica gel (petroleum ether-ethyl acetate) and further isolated by preparative TLC (petroleum ether-acetone) to afford TCSM (204 mg). The purity and structure of TCSM were verified and identified by HPLC, mass spectrometry and NMR. Additionally, the extraction and isolation were performed many times in order to obtain enough TCSM.

Cell culture

PANC-1 cells were purchased from the American Type Culture Collection (ATCC, Manassas, VA, USA) and routinely cultured in DMEM medium supplemented with 10 % FBS, 100 U/mL penicillin and 100 U/mL streptomycin at 37 °C in 5 % CO₂ and 95 % O₂.

MTT assay

The cytotoxic activity of TCSM against PANC-1 cells was evaluated by MTT assay. PANC-1 cells were seeded on 96-well culture plates with DMEM medium. After 24 h of incubation, cells were treated with TCSM at concentrations of 0 (for control), 5, 10, 15, 20, 25 and 30 μ M. After 48 h, MTT was added into every well, and cells were cultured at 37 °C in 5 % CO₂ and 95 % O₂ for another 3 h. Then the optical density (OD) was measured at 570 nm with a micro plate reader (Thermo, USA). The inhibition rate of TCSM against PANC-1 cells could be determined by the following equation: inhibition

Cytotoxic activity of TCSM against PANC-1 cells

The results of MTT assay indicated that the inhibition rates of TCSM in PANC-1 cells were between 9.28 and 83.23 %, and the IC₅₀ value was 19.16 μ M. Namely, TCSM showed good cytotoxic activity against PANC-1 cells. The results are shown in Figure 2.

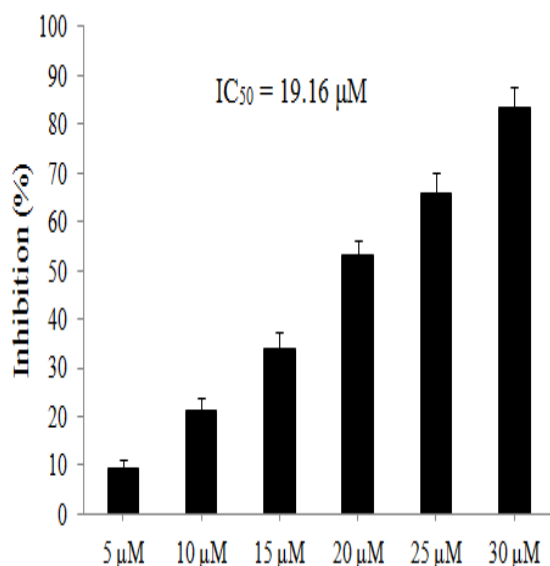


Figure 2: Cytotoxic activity of TCSM against PANC-1 cells

TCSM induces apoptosis in PANC-1 cells

Flow cytometry was used to study whether the cytotoxic activity of TCSM against PANC-1 cells was related to apoptosis. The results of flow cytometry suggested that the cytotoxic activity of TCSM was related to apoptosis. As shown in Figure 3, the apoptosis rates were 6.12, 24.45, 35.26 and 54.16 % after treatment with TCSM at concentrations of 0 (for control), 5, 10 and 20 μ M, and compared with the control group, the apoptosis rates of TCSM groups were significantly ($p < 0.01$) increased.

Effects of TCSM on the expressions of mitochondria-mediated apoptotic proteins

Western blot was used to study the pro-apoptotic mechanism of TCSM in PANC-1 cells. As shown in Figures 4 and 5, after PANC-1 cells were treated with TCSM at concentrations of 5, 10 and 20 μ M, the expressions of anti-apoptotic proteins (Bcl-2 and Survivin) were significantly ($p < 0.01$) down-regulated, the expressions of pro-apoptotic proteins (Bax, Smac, c-caspase-3 and c-caspase-9) were significantly ($p < 0.01$) up-regulated, and the release of cytochrome c from the mitochondria to the cytoplasm was significantly ($p < 0.01$) promoted, compared with the control group.

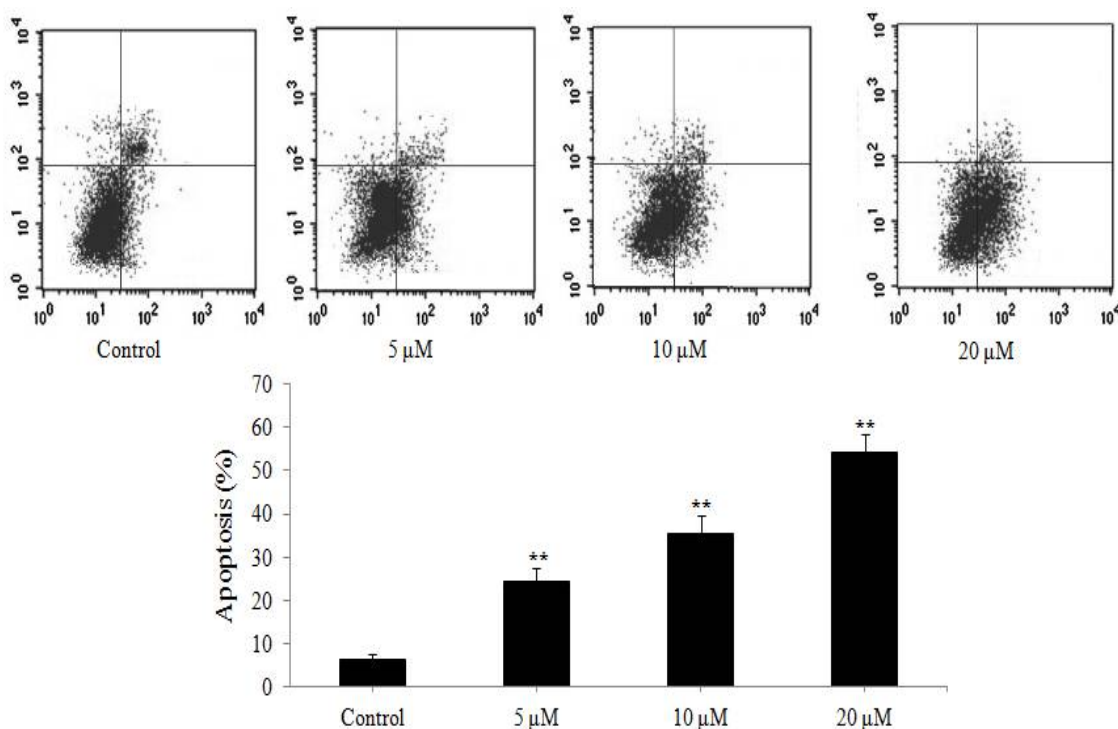


Figure 3: Effect of TCSM on apoptosis in PANC-1 cells; ** $p < 0.01$, compared with the control group

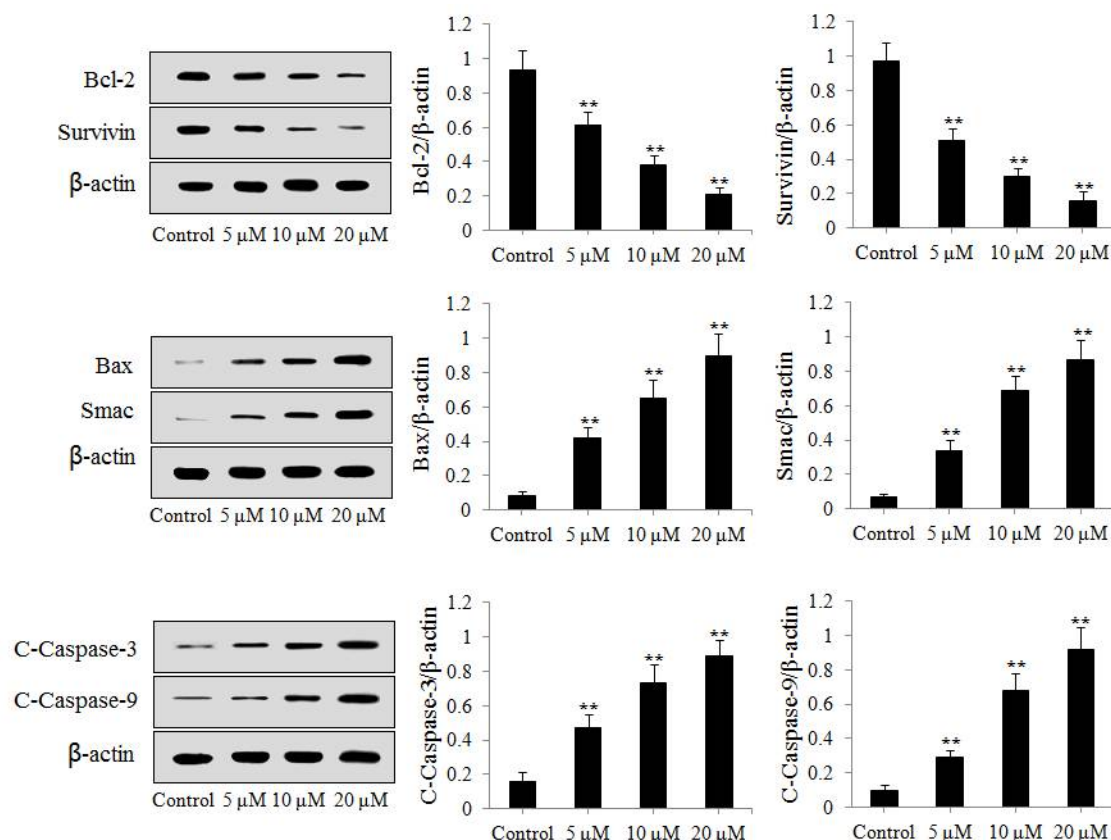


Figure 4: Effect of TCSM on anti-apoptotic proteins (Bcl-2 and Survivin) and pro-apoptotic proteins (Bax, Smac, c-caspase-3 and c-caspase-9) *in vitro*; ** $p < 0.01$, compared with the control group

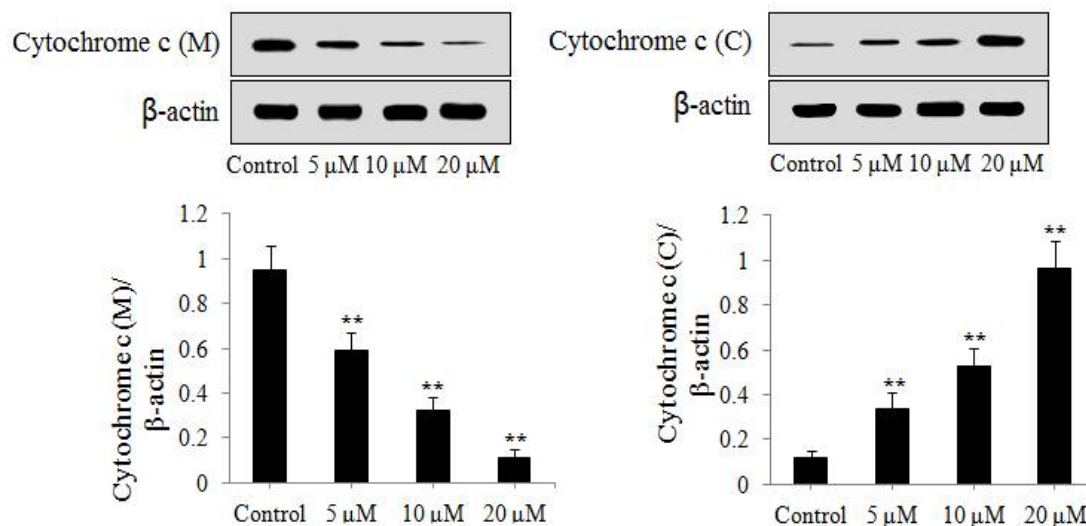


Figure 5: Effect of TCSM on the release of cytochrome c from the mitochondria to the cytoplasm; Cytochrome c (M): cytochrome c in mitochondria; Cytochrome c (C): cytochrome c in cytoplasm; ** $p < 0.01$, compared with the control group

Effects of TCSM on PANC-1 cell-induced xenograft model

PANC-1 cell-induced xenograft model was used to confirm whether TCSM could inhibit the growth of PANC-1 cell-induced tumor and regulate the

expressions of apoptotic proteins *in vivo*. The results of xenograft assay indicated that TCSM (20 mg/kg once a day for 20 day) significantly ($p < 0.05$ or 0.01) inhibited the growth of PANC-1 cell-induced tumor without any effect on the body weight of the nude mice during the 20 days'

observation period, compared with the control group. Moreover, the results of western blot of tumor tissues indicated that TCSM significantly ($p < 0.01$) down-regulated the expressions of anti-apoptotic proteins (Bcl-2 and Survivin) and up-

regulated the expressions of pro-apoptotic proteins (Bax, Smac, c-caspase-3 and c-caspase-9) *in vivo*, compared with the control group. The results are shown in Figures 6 and 7.

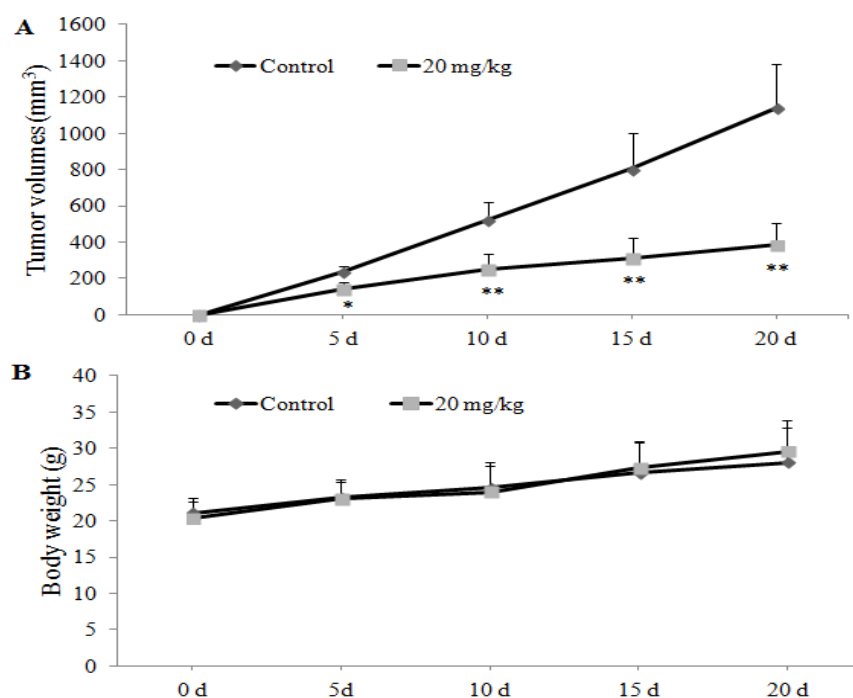


Figure 6: Effect of TCSM on tumor growth and body weight of nude mice bearing PANC-1 cells xenograft; * $p < 0.05$, ** $p < 0.01$, compared with the control group

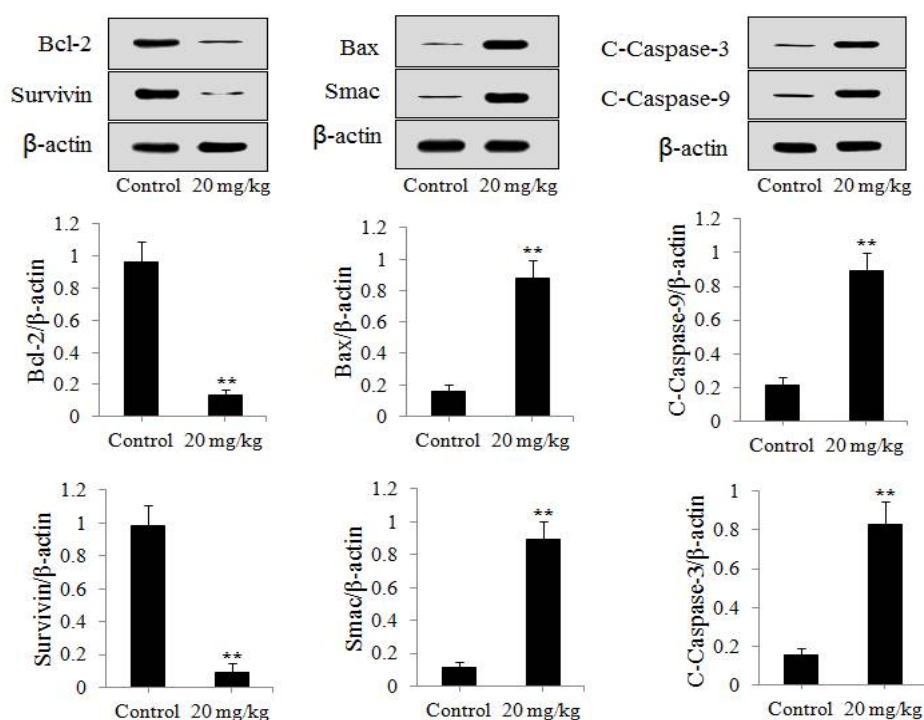


Figure 7: Effect of TCSM on anti-apoptotic proteins (Bcl-2 and Survivin) and pro-apoptotic proteins (Bax, Smac, c-caspase-3 and c-caspase-9) *in vivo*; ** $p < 0.01$, compared with the control group

DISCUSSION

In the present study, we investigated the cytotoxic activity of TCSM against PANC-1 cells and possible mechanisms of action for the first time by MTT, flow cytometry, Western blot and xenograft assays.

MTT assay is a commonly used method to study the cytotoxic activity of compound against cells in vitro [12]. Flow cytometry is a commonly used method to investigate whether the cytotoxic activity of compound is related to apoptosis [13]. The results of MTT assay (Figure 2) indicated that TCSM showed good cytotoxic activity against PANC-1 cells with an IC₅₀ of 19.16 μ M. The results of flow cytometry assay (Figure 3) confirmed that the cytotoxic activity of TCSM against PANC-1 cells was related to apoptosis.

Western blot is a commonly used method to explore anticancer mechanism at the protein level, and the effect of compound on tumor growth and body weight of nude mice are two important indices in vivo [14,15]. Proteins including Bcl-2, Survivin, Bax, Smac, cytochrome c, c-caspase-3 and c-caspase-9 are very important markers in mitochondria-mediated apoptotic pathway [16]. The relations among these apoptotic proteins are rather complex and described as follows [17-21]. When cells suffer from apoptotic stimuli, pro-apoptotic proteins (Smac and cytochrome c) are released from the mitochondria to the cytoplasm, but their release can be inhibited by anti-apoptotic protein (Bcl-2). However, the function of Bcl-2 can be inhibited by pro-apoptotic protein (Bax).

Cytochrome c along with apoptotic protease activating factor-1 (Apaf-1), caspase-9 and ATP can form apoptosome in the cytoplasm to activate caspase-9 (c-caspase-9), and then c-caspase-9 can promote the activation of caspase-3 (c-caspase-3), which can lead to apoptosis of cells. The results of western blot assay (Figures 4 and 5) suggested that TCSM down-regulated the expressions of anti-apoptotic proteins (Bcl-2 and Survivin), up-regulated the expressions of pro-apoptotic proteins (Bax, Smac, c-caspase-3 and c-caspase-9) and promoted the release of cytochrome c from the mitochondria to the cytoplasm in PANC-1 cells.

The results of the xenograft assay (Figures 6 and 7) in nude mice indicated that TCSM inhibited the growth of PANC-1 cell-induced tumor without any effect on the body weight of the nude mice by regulating the expressions of mitochondria-mediated apoptotic proteins in vivo as described above.

CONCLUSION

TCSM exhibits cytotoxic activity against PANC-1 cells *in vitro* and *in vivo* through mitochondria-mediated apoptotic pathway. Further investigation into the mechanism of action of TCSM is required.

CONFLICT OF INTEREST

The authors declare no conflict of interest associated with this work.

CONTRIBUTION OF AUTHORS

We declare that this work was done by the authors named in this article and all liabilities pertaining to claims relating to the content of this article will be borne by the authors. Jian-Bo Zhou, Gang Peng, Jun Li and Yu-Cheng Jia conceived, designed and carried out the study and written the manuscript. Jia Wang, Shu Nie and Qiu-Ying Zhang collected and analyzed the data.

REFERENCES

1. Li DH, Xie KP, Wolff R, Abbruzzese JL. Pancreatic cancer. *The Lancet* 2004; 363: 1049-1057.
2. Li CW, Heidt DG, Dalerba P, Burant CF, Zhang LJ, Adsay V, Wicha M, Clarke MF, Simeone DM. Identification of pancreatic cancer stem cells. *Cancer Res* 2007; 67(3): 1030-1037.
3. Lau ST, Lin ZX, Liao YH, Zhao M, Cheng CHK, Leung PS. Brucein D induces apoptosis in pancreatic adenocarcinoma cell line PANC-1 through the activation of p38-mitogen activated protein kinase. *Cancer Lett* 2009; 281(1): 42-52.
4. Wang XL, Zheng Z, Hong ZY, Fan GR. Advancements on chemical components and quality control of *Rhizoma Corydalis*. *Lishizhen Med Mater Med Res* 2011; 22: 227-229.
5. Gao JL, He TC, Li YB, Wang YT. A traditional Chinese medicine formulation consisting of *Rhizoma corydalis* and *Rhizoma Curcumae* exerts synergistic anti-tumor activity. *Oncol Rep* 2009; 22(5): 1077-1083.
6. Zhang GD, Xie L, Yu LX, Hu WJ, Qian XP, Liu BR. Antiproliferation effect of total alkaloids fraction of *Yahusuo* on six human gastric cancer cell lines *in vitro*. *Chin J Integr Trad West Med Dig* 2009; 17: 81-85.
7. Piyanuch R, Sukthanthar M, Wandee G, Baek, SJ. Berberine, a natural isoquinoline alkaloids, induces NAG-1 and ATF3 expression in human colorectal cancer cells. *Cancer Lett* 2007; 258(2): 230-240.
8. Gao JL, Shi JM, Lee SMY, Zhang QW, Wang YT. Angiogenic pathway inhibition of *corydalis yanhusuo* and berberine in human umbilical vein endothelial cells. *Oncol Res* 2009; 17(11-12): 519-526.

9. The National Research Council of The National Academy of Sciences. *Guide for the Care and Use of Laboratory Animals: Eight Edition*. Washington, D.C.: The National Academies Press; 2010.
10. Zhao YL, Zhao LJ, Luo YX, Li X, Zhang YF, Liu XD, Luo YG, Zhong LL. Synergistic effect of radiation and traditional Chinese medicine *Rhizoma Typhonii* ethanol extract depends on p53 expression in treatment of lewis mouse lung cancer cells. *Afr J Tradit Complement Altern Med* 2015; 12(1): 109-114.
11. Wang HS, Yang HR. The alkaloids of *Corydalis dasystern maxim*. *Nat Prod Res Dev* 1997; 9(1): 37-39.
12. Yang QH, Yang J, Liu GZ, Wang L, Zhu TC, Gao HL, Kou XG. Study on in vitro anti-tumor activity of *Bidens bipinnata* L. extract. *Afr J Tradit Complement Altern Med* 2013; 10(3): 543-549.
13. Yuan JL, Wang SM, Lan T, Liu JZ, Wang GH, Sun QS, Chen H. Studies on anti-hepatoma effect of Gan-Ai-Xiao decoction. *Trop J Pharm Res* 2015; 14(7): 1249-1255.
14. Liu HY, Zhu YJ, Zhang T, Zhao ZG, Zhao Y, Cheng P, Li H, Gao H, Su XM. Anti-tumor effects of atractylenolide I isolated from *Atractylodes macrocephala* in human lung carcinoma cell lines. *Molecules* 2013; 18(11): 13357-13368.
15. Yang XK, Xu MY, Xu GS, Zhang YL, Xu ZX. In vitro and in vivo antitumor activity of scutebarbatine A on human lung carcinoma A549 cell lines. *Molecules* 2014; 19(7): 8740-8751.
16. Shi YG. A structural view of mitochondria-mediated apoptosis. *Nat Struct Biol* 2001; 8(5): 394-401.
17. Deng YB, Lin YH, Wu XW. TRAIL-induced apoptosis requires Bax-dependent mitochondrial release of Smac/DIABLO. *Gene Dev* 2002; 16(1): 33-45.
18. Ambrosini G, Adida C, Altieri DC. A novel anti-apoptosis gene, surviving, expressed in cancer and lymphoma. *Nat Med* 1997; 3(8): 917-921.
19. Du CY, Fang M, Li YC, Li LL, Wang XD. Smac, a mitochondrial protein that promotes cytochrome c-dependent caspase activation by eliminating IAP inhibition. *Cell* 2000; 102(1): 33-42.
20. Kuida K, Haydar TF, Kuan CY, Gu Y, Taya C, Karasuyama H, Su MS, Rakic P, Flavell RA. Reduced apoptosis and cytochrome c-mediated caspase activation in mice lacking caspase 9. *Cell* 1998; 94(3): 325-337.
21. Lakhani SA, Masud A, Kuida K, Porter Jr GA, Booth CJ, Mehal WZ, Inayat I, Flavell RA. Caspase 3 and 7: key mediators of mitochondrial events of apoptosis. *Science* 2006; 311(5762): 847-851.