Available online at http://www.tjpr.org http://dx.doi.org/10.4314/tjpr.v23i10.7

### **Original Research Article**

# Toxicological evaluation of hydro-alcohol root extract of *Rauwolfia vomitoria* Afzel (Apocynaceae)

Valentine O Adegoke<sup>1\*</sup>, Emmar E Okpakpor<sup>2</sup>, Dickson O Uwaya<sup>2</sup>, Oluwakanyinsola A Salawu<sup>3</sup>, Raymond I Ozolua<sup>4</sup>

<sup>1</sup>National Institute for Pharmaceutical Research and Development, Idu, Abuja, <sup>2</sup>Department of Science Laboratory Technology, Faculty of Life Sciences, University of Benin, Benin City 300001, <sup>3</sup>Department of Pharmacology & Toxicology, Faculty of Pharmaceutical Sciences, Gombe State University, Gombe, <sup>4</sup>Department of Pharmacology & Toxicology, Faculty of Pharmacy, University of Benin, Benin City 300001, Nigeria

\*For correspondence: Email: valadegoke@gmail.com; Tel: +2347038930984

Sent for review: 10 May 2024 Revised accepted: 5 October 2024

#### **Abstract**

**Purpose:** To investigate the phytochemical content and toxicity profile of Rauwolfia vomitoria Afze (Apocynaceae).

**Method:** Qualitative phytochemical screening of hydro-alcohol extract (HAE) of R. vomitoria roots was carried out followed by acute toxicity evaluation in nine Swiss albino Wistar rats using two phases of Lorke's method. In the 14 days subacute study, thirty-two Swiss albino rats were randomly divided into 4 groups of 8 rats each. Group A received 0.5 mL Tween 80 daily and served as control, while groups B, C and D was administered 125, 250 and 500 mg/kg/day of HAE respectively. All the rats were monitored daily for signs of toxicity. Selected haematological and biochemical parameters were assessed at the termination of experiment.

**Results:** Alkaloids, cardiac glycosides, flavonoids and saponins were present in HAE. The estimated oral median lethal dose ( $LD_{50}$ ) was greater than 5000 mg/kg. Daily administration of HAE for 14 days resulted in 15.63 % mortality of animals across the groups by day 14 with significant loss in total body weight in group D animals on day 8 (p < 0.05), and in group C animals on day 11 (p < 0.05) and day 15 (p < 0.001). The dose of 500 mg/kg/day significantly (p < 0.05) increased haemoglobin concentration but reduced (p < 0.05) white blood cell count, while the dose of 250 mg/kg/day significantly (p < 0.05) reduced alkaline phosphatase levels but increased serum albumin levels (p < 0.05).

**Conclusion:** Rauwolfia vomitoria may be safe on acute basis but repeated administration of high doses over many days needs to be done with caution.

Keywords: Rauwolfia vomitoria, Sickle cell, Alkaloids, Flavonoids, Saponins, Toxicity

This is an Open Access article that uses a funding model which does not charge readers or their institutions for access and distributed under the terms of the Creative Commons Attribution License (http://creativecommons.org/licenses/by/4.0) and the Budapest Open Access Initiative (http://www.budapestopenaccessinitiative.org/read), which permit unrestricted use, distribution, and reproduction in any medium, provided the original work is properly credited.

Tropical Journal of Pharmaceutical Research is indexed by Science Citation Index (SciSearch), Scopus, Web of Science, Chemical Abstracts, Embase, Index Copernicus, EBSCO, African Index Medicus, JournalSeek, Journal Citation Reports/Science Edition, Directory of Open Access Journals (DOAJ), African Journal Online, Bioline International, Open-J-Gate and Pharmacy Abstracts

#### INTRODUCTION

Rauwolfia vomitoria Afzel, (Apocynaceae) is known by many names such as asofeyeje by the Yorubas in western Nigeria, akkemta by the Igbos, wada by the Hausas, mmoneba by the Efiks, and utoenyin by the Ibibios [1]. It is an

evergreen perennial shrub found mainly in the wild in Africa with height reaching up to 15 m, with oblong or oval shiny leaves, depending on the location [2]. The constituents of *Rauwolfia vomitoria* is reported to possess antipsychotic, antihypertensive, antidiarrheal [2] and antisickling effects [1,3] amongst other useful

pharmacological activities, while the safety of *R. vomitoria* has attracted global concern.

The reported anti-sickling property of a plant recipe containing *R. vomitoria* root and leaf extract [1,3] necessitated further evaluation of the safety profile of its hydro-alcohol extract in the present study since management or prophylaxis of sickle cell disease require that doses of selected therapeutic agent are safe both for short time and chronic use across all age groups.

#### **EXPERIMENTAL**

#### Plant material

Roots of *R. vomitoria* were collected between August and September 2015 from the Botanical Garden of University of Ibadan, Ibadan, Oyo State, Nigeria. The specimen was authenticated by Dr Grace Ugbabe of the Department Medicinal Plant Research and Traditional Medicine, National Institute for Pharmaceutical Research and Development, (NIPRD), Idu, Abuja. The specimen was assigned a voucher no. NIPRD/H/6715 and kept in the Herbarium of NIPRD, at Idu, Abuja. The roots were carefully cleaned, air-dried under shade and ground to coarse powder using mortar and pestle before further pulverisation.

## Preparation of hydro-alcohol extract (HAE) of R. vomitoria

The method of Egunyomi *et al* [3] with slight modification was used. The powdered roots of *R. vomitoria* (800 g) was soaked in a mixture of solvents (3 L of distilled water and 3.6 L of 95 % ethanol) inside a 5 L sealed flat bottom flask for five days with intermittent stirring using a spatula. The extract was thereafter filtered with Whatman filter paper and the filtrate concentrated to dryness on a water bath at 70 °C to obtain the extract which was subsequently stored in a labelled sample bottle at 4 °C.

#### Animals

Swiss albino Wistar rats, weighing 167.69 ± 29.57 g (mean ± SD), of equal sexes were obtained from the Animal House of the Faculty of Pharmacy, University of Benin, Benin-City, Nigeria. They were kept in standard plastic cages and had free access to water and feeds (Livestock Feeds Ltd, Ibadan, Nigeria). All the animals were exposed to natural lighting, maintained at room temperature (30 °C) and handled according to international protocols for use of laboratory animals in experiments [4].

Ethical approval (EC/FP/021/01) was obtained from the Ethical Committee, Faculty of Pharmacy, University of Benin, Benin City, Nigeria. Chemicals were sourced from reputable manufacturers such as Sigma-Aldrich and reagents were freshly prepared before use.

#### Preliminary phytochemical screening

Preliminary phytochemical screening to identify the class of secondary metabolites in HAE was done using the methods described in literature [5].

#### Oral acute toxicity test

The oral median lethal dose (LD50) was estimated using the method of Lorke [6]. In the first phase, nine (09) rats of both sexes were randomly assigned to three groups (n = 3)labelled A, B, and C. With the aid of an orogastric tube, animals in groups A, B and C were administered 10, 100 and 1000 mg/kg of HAE, respectively. The animals were observed continuously for the first four hours after dosing, after 24 h and subsequently, after 72 h for signs of toxicity such as changes in behaviour (pawlicking, salivation, stretching, mood, motor activity and gnawing), posture, nature and frequency of stooling, and mortality. The absence of death in this phase necessitated the next. For the second phase, three rats were randomly assigned to three groups (n = 1). With the aid of an orogastric tube, doses of 1600, 2900 and 5000 mg/kg of HAE were administered to each group, respectively. The animals were observed as was done in phase 1 above. The LD50 value was calculated using Eq 1.

$$LD_{50} = \sqrt{(D_0 \times D_{100})}$$
 .....(1)

Where:  $D_0$  = Highest dose that gave no mortality and  $D_{100}$  = Lowest dose that produced mortality

#### Oral sub-acute toxicity test

Sub-acute toxicity testing was carried out using the method of Ozolua *et al* [9] with slight modification. Thirty-two (32) rats were randomly allotted into 4 groups of 8 rats each (4 males and 4 females). The rats in group A were administered 0.5 mL of 10 % Tween 80 daily while rats in groups B, C and D received 125, 250 and 500 mg/kg/day of HAE, respectively for fourteen days orally via orogastric tube. All the rats were monitored daily for signs of toxicity such as depression and death. Rats that survived were fasted overnight prior to sacrificing on day 15. Animals were anaesthetized in an

airtight glass chamber saturated with chloroform and a portion of blood (5 mL) was drawn from the abdominal aorta of each rat, from which 1 mL was collected into ethylene diamine-tetra-acetic acid (EDTA)-containing bottle for haematological analysis. The remaining portion of blood (4 mL) was collected into a plain bottle and used for biochemical analysis.

#### Haematological analysis

Automatic analyser (ERMA PCE-210, Japan) was calibrated and programmed to analyse the blood samples in EDTA bottles for the following parameters: red blood cell count (RBC), haemoglobin concentration (Hgb), haematocrit (HCT), white blood cell count (WBC), platelets count (PLT); lymphocytes count (LY), monocytes count (MO) and mean platelet volume (MPV).

#### **Biochemical analysis**

The blood samples in the plain bottles were allowed to clot at room temperature for 4 h and the resulting serum (1 mL) was analysed using automatic clinical chemistry analyzer (Architect c4000, Abbott Diagnostics, Japan) for alkaline phosphatase (ALP), aspartate aminotransferase (AST), alanine aminotransferase (ALT), total bilirubin (TBil), conjugated bilirubin (CBil), total proteins (TP), albumin (Alb), creatinine and urea levels. Automatic analyser (Selectra Pro S. ELITech. USA) was used to determine the concentration sodium. of potassium. bicarbonates and chloride ions in each serum sample.

#### Statistical analyses

All results were expressed as mean  $\pm$  standard error of mean (SEM). Comparisons were made between groups by use of one-way analysis of variance (ANOVA) followed by Tukey's *post hoc* test. All data were analysed using GraphPad Prism software version 6 (GraphPad Prism, USA). P < 0.05 indicates statistically significant difference.

#### **RESULTS**

#### Phytochemical and extraction yield

Phytochemical screening of hydro-alcohol extract of *R. vomitoria* root (HAE) showed that it contained alkaloids, cardiac glycosides, flavonoids and saponins (Table 1). The hydro-alcohol extraction yield was 5.62 % w/w.

**Table 1:** Classes of secondary plant metabolites identified in the hydro-alcohol extract of *R. vomitoria* roots

Phytochemical Test	Qualitative
Alkaloid	+
Cardiac glycosides	+
Flavonoids	+
Saponins	+
Tannins	-
Glycosides	-
Anthraquinones	-
Phlobotannins	-

Note: - = absent, + = present

#### **Oral acute toxicity**

There was no lethality at acute oral doses of 10, 100 and 1000 mg/kg in rats given HAE (Table 2). The rats did not exhibit any significant sign of toxicity such as writhing, diarrhoea, hyper-motility and aggression. Mild depressant activity was observed in the rats given 1000 mg/kg but they recovered within 48 h. The rats manifested toxicological signs of restlessness (writhing) after administration of extract at 1600, 2900 and 5000 mg/kg, but became calm 2 h afterwards. After 24 h, all the rats manifested signs of depression (hypo-activity) as evident from their shut eyes, orbital red patches, immobility and lack of interest in the environment. The absence of death at 5000 mg/kg after 48 h indicated that the LD<sub>50</sub> of HAE was greater than 5000 mg/kg.

#### **Oral sub-acute toxicity**

The signs of depression such as loss of corneal reflex/partial eye closure were more observable in rats given 500 mg/kg/day than those given 250 mg/kg/day after one week of HAE administration. During the experiment, mortality was recorded in all treated groups, made up of two rats each in 250 and 500 mg/kg/day groups on days 9 and 11, respectively and one rat in 125 mg/kg/day group on day 13. No death was observed among the rats in control group (Table 3).

#### Effects on haematological indices

There was a significant (p < 0.05) increase in haemoglobin concentration in the group of animals that received 500 mg/kg/day of HAE compared to the animals in control group. Other red blood cell (RBC) parameters such as RBC count, haematocrit levels and mean corpuscular haemoglobin concentration in all treated groups were not significantly different from those of control (Table 4 a). The decrease in white blood cell counts within the 500 mg/kg/day group was significant (p < 0.05) compared to control.

Table 2: Oral acute toxicity effects of the hydro-alcohol extract of R. vomitoria roots in Wistar rats

Phase	Dose (mg/kg)	Writhing	Depression	Diarrhoea	Tremor	Death	Others
I	10	0/3	0/3	0/3	0/3	0/3	0/3
	100	0/3	0/3	0/3	0/3	0/3	0/3
	1000	0/3	3/3	0/3	0/3	0/3	0/3
II	1600	1/1	1/1	0/1	0/1	0/1	0/1
	2900	1/1	1/1	0/1	0/1	0/1	0/1
	5000	1/1	1/1	0/1	0/1	0/1	0/1

Numerator = number of animals affected. Denominator = number of animals in the group.  $LD_{50} > 5000$  mg/kg (p.o.)

Table 3: Toxicological effects following oral daily doses (x14 days) of HAE of R. vomitoria in Wistar rats

Group		Morta	lity	Symptoms		
	Number	(%)	Latency (Days)	•		
Control	0	0		None		
125 mg/kg/day	2	25	9, 11	None		
250 mg/kg/day	1	13	13	Anorexia, hypoactivity, loss of corneal reflex		
500 mg/kg/day	2	25	9, 11	Anorexia, hypoactivity, loss of corneal reflex		

#### Effects on haematological indices

There was a significant (p < 0.05) increase in haemoglobin concentration in the group of animals that received 500 mg/kg/day of HAE compared to the animals in control group. Other red blood cell (RBC) parameters such as RBC count, haematocrit levels and mean corpuscular haemoglobin concentration in all treated groups were not significantly different from those of control (Table 4). The decrease in white blood cell counts within the 500 mg/kg/day group was significant (p < 0.05) compared to control. The mean platelet volume (MPV) and lymphocytes count were significantly decreased (p < 0.001) in the groups that received 250 and 500 mg/kg/day compared to control, although there was no significant (p > 0.05) change in platelet and monocytes counts. The decrease in lymphocyte count among treated groups was observed to be dose-dependent (Table 4).

#### Effect on enzymes and proteins

The level of alkaline phosphatase (ALP) was significantly (p < 0.05) lower in the group of rats that received 250 mg/kg/day of HAE but levels of other serum enzymes (aspartate aminotransferase and alanine amino-transferase) in all treated groups were not significantly (p > p)0.05) different from those of control (Table 5). Also, albumin levels were significantly (p < 0.05) higher in 250 mg/kg/day group when compared to control. Other serum proteins such as total bilirubin, conjugated bilirubin and total protein were not significantly (p > 0.05) different compared to control (Table 6).

#### Effect on electrolytes, urea and creatinine

The levels of sodium ions (Na<sup>+</sup>) in the group that received 500 mg/kg/day were significantly elevated compared to control ((p < 0.05; Table 6 a) but there were no significant changes in the levels of other electrolytes, urea and creatinine in all other treatment groups in comparison with control group ((p > 0.05; Table 7).

#### DISCUSSION

The hydro-alcohol extract (HAE) of *R. vomitoria* root contains alkaloids, saponins, cardiac glycosides and flavonoids. The constituents identified in this study were also detected by Abere *et al* [1]. However, the area of divergence was likely due to the number and differences in test protocols used for qualitative analysis.

The calculated oral LD $_{50}$  obtained for HAE of R. vomitoria roots in this present study was greater than 5000 mg/kg and is consistent with the results obtained from Abere et al which evaluated the acute toxicity of R. vomitoria leaves in mice [1]. The result is also similar to the findings of Ebuehi et al that estimated LD $_{50}$  of aqueous extract of R. vomitoria root in rats using the Lorke method [8]. However, the LD $_{50}$  obtained for the root extract in this study is at variance with the value estimated by Ebuehi et al for the ethanol extract of the R. vomitoria root using the Probit curve analysis technique [8].

Table 4: Effects of oral daily doses (x14 days) of HAE of *R. vomitoria* on blood parameters

Group	RBC (x10 <sup>6</sup> /μL)	HgB (g/dL)	HCT (%)	MCH (pg)	PLT (10³/ul)	WBC (x103/ul	LY (%	MO (%	MPV (fl)
Control	6.02±0.23	13.89±0.50	38.81±1.73	23.08±0.45	375.40±64.	13.08±2.0	7.16±0.52	1.78±0.48	3.74±0.10
125 mg/kg/day	5.71±0.89	14.44±1.37	35.76±5.36	31.80±8.23	548.90±83.	9.11±0.97	5.69±0.82	0.91±0.25	3.53±0.16
250 mg/kg/day	6.98±0.46	16.10±0.78	41.95±2.78	23.15±0.50	474.70±30.	7.18±0.97	3.70±0.64	0.88±0.31	3.12±0.04
500 mg/kg/day	7.57±0.26	17.50±0.67*	50.75±1.51	23.05±0.19	650.00±86.	4.25±1.54	2.83±1.39	0.40±0.07	3.15±0.03

<sup>\*</sup>P < 0.05 vs Control. RBC: red blood count; HgB: haemoglobin; HCT: haematocrit; MCH: mean cell haemoglobin, PLT: platelet; WBC: white blood count; LY: lymphocyte; MO: monocyte

Table 5: Effects of oral daily doses (x14 days) of HAE of R. vomitoria on some serum enzymes

Enzyme	Control	125 mg/kg/day	250 mg/kg/day	500 mg/kg/day
ALP (iU/L)	691.9±64.5	479.6±79.5	407.0±61.25*	456.3±23.4
AST (iU/L)	42.3±4.2	45.2±3.5	48.0±4.0	62.7±10.1
ALT (iU/L)	80.6±10.0	73.7±15.3	82.4±4.9	60.5±20.7

<sup>\*</sup>P < 0.05 vs Control. ALP: Alkaline phosphatase, AST: Aspartate aminotransferase, ALT: Alanine amino-transferase

Table 6: Effects of oral daily doses (x14 days) of HAE of R. vomitoria on serum proteins

Parameter	Control	125 mg/kg/day	250 mg/kg/day	500 mg/kg/day
Total bilirubin (µm/L)	0.33±0.05	0.34±0.06	0.34±0.04	0.37±0.09
Conjugated bilirubin (µm/L)	0.15±0.02	0.14±0.03	0.16±0.02	0.13±0.03
Total protein (g/dL)	0.61±0.02	0.62±0.04	0.62±0.03	0.60±0.04
Albumin (g/dL)	3.94±0.11	4.36±0.11	4.54±0.16*	4.30±0.25

<sup>\*</sup>P < 0.05 vs Control

Table 7: Effects of oral daily dose (x14 days) of HAE of R. vomitoria on serum electrolytes

Group	Na+ (mol/L)	K+ (mol/L)	HCO₃ (mol/L)	CI · (mol/L)	Creatinine (µmol/L)	Urea (mol/L)
Control	143.80±0.56	6.79±0.20	28.00±1.40	108.50±0.68	0.61±0.02	38.00±1.75
125 mg/kg/day	145.80±1.60	6.84±0.33	26.40±0.51	108.00±0.95	0.62±0.04	36.00±1.34
250 mg/kg/day	146.00±1.18	6.93±0.21	26.00±0.63	108.80±1.66	0.62±0.04	44.67±5.24
500 mg/kg/day	151.50±3.70*	7.63±0.42	28.75±0.75	112.30±2.39	0.60±0.04	37.50±2.39

<sup>\*</sup>P < 0.05 vs Control

Based on OECD guidelines and the Hodge and Sterner scale classification [9], the HAE of R. vomitoria may be regarded as practically safe for consumption. In the oral sub-acute toxicity test results, the signs of depressant activity such as sedation and loss of corneal reflex observed in animals treated with 250 and 500 mg/kg/day of HAE nine days after the commencement of experiment suggest the involvement of the central nervous system (CNS) in the extract's toxic effect and the ability of the active phytochemicals to cross the blood-brain barrier. These symptoms and mortality recorded in all the treatment groups were not observed in the findings of Abere et al [1] but are consistent with that of Ebuehi et al [8] which may probably be due to differences in the test protocol. Doses used in the sub-acute test (one-tenth, onetwentieth, and one-fortieth of the maximum dose of 5000 mg/kg used for the LD50 evaluation), appeared toxic as indicated by mortality.

The significant increase in hemoglobin concentration and the lack of significant difference in other hematological indices such as red blood cell count, hematocrit level and mean cell hemoglobin in all treated groups when compared to control as observed in this study is at variance with the findings of Isaiah et al [10] in which 150 mg/kg of the root bark extract exerted significant increase in hemoglobin concentration, red blood cell count and hematocrit level. The decrease in white blood cell count observed in this study may be due to toxic effect of HAE of R. vomitoria root on the immune system and is consistent with the findings of Isaiah et al [10] who reported a decrease in white blood cell count, lymphocyte count and mean platelet volume following administration of 150 and 300 mg/kg of ethanol extract of R. vomitoria bark to Albino Wistar rats. However, the findings differ from that of Bonheur et al [11] which reported elevated white blood cell count, lymphocyte and platelet counts following administration of 900 mg/kg of aqueous extract of R. vomitoria stem bark to male Wistar rats. The difference in dosage of administration, as well as part of the plant used, could be the reason for the different observations.

Although higher levels of ALT and AST are often diagnostic of underlying cellular injuries, this was not the case in this study as the levels of the two enzymes - ALT and AST were not significantly different between treated and control groups [7]. The absence of a significant change in the levels of ALT and AST in all treated groups could imply the safety of HAE of *R. vomitoria* in the hepatic system. The safety of HAE at the dose

investigated is further confirmed by the absence of significant changes in the levels of total bilirubin, conjugated bilirubin and total protein. Although the levels of albumin were elevated in animals that received 250 mg/kg of HAE, this increase was not dose-dependent. However, increased albumin levels have been associated with improved liver health [2]. Research evidence also revealed that elevated albumin levels could be a result of dehydration, infections, congenital disorders, liver injury, malnutrition, chronic inflammatory disease, and diminished protein intake among other physiological biochemical factors [12]. The reduction in the levels of ALP as obtained in this study is at variance with the findings of Eteng et al [2] but agrees with the observations of Ezejindu et al [13] possibly due to differences in the dose of ethanol extract of R. vomitoria root administered.

Furthermore, the estimated serum concentration of sodium, potassium, chloride and bicarbonate electrolytes obtained in this study are all within the reference values of 141 - 150 mmol/L, 5.2 - 7.8 mmol/L, 99 - 114 mmol/L, and 24 - 31 mmol/L, respectively [14]. Although the serum concentration of sodium ion was significantly elevated in animals that received 500 mg/kg of HAE, suggestive of the potential ability of HAE to increase blood pressure, the change in serum concentrations of other electrolytes, especially potassium, was not significant, thus implying that HAE may possess cardio-protective activities [15].

The concentration of serum creatinine in healthy rats is usually in the range of 0.4 – 0.8 mg/dL [16] while the reference values for serum urea concentrations in rats vary, depending on the dietary protein content and dietary intake of protein [17], but usually within the range of 15 -26 mg/dL [16]. The unremarkable variations in serum urea concentration of all treated animals may be due to their attempts to adapt to the effects of HAE which was not achieved within the 14 days of experiment [18]. Although the levels of serum urea in this study are higher than reference values in all treated animals and controls, the change in creatinine and urea levels in treated groups was not significant compared to controls, thus implying that HAE did not exert any deleterious effect on the renal function.

#### CONCLUSION

The hydro-alcohol extract (HAE) of R. vomitoria root contains alkaloids, cardiac glycosides, flavonoids and saponins. The oral LD<sub>50</sub> > 5000

mg/kg shows that the extract appears safe on acute basis but sub-acute doses used in the present study all appear toxic, thus indicating the need for monitoring of serum electrolytes, protein and liver function.

#### **DECLARATIONS**

#### **Acknowledgements**

The authors wish to express gratitude to Dr Grace Ugbabe for the taxonomic identification of sample and Mr. Ibe of Pharmacology and Toxicology Department for taking care of the animals.

#### **Funding**

None provided.

#### Ethical approval

Ethical approval (EC/FP/021/01) was obtained from the Ethical Committee, Faculty of Pharmacy, University of Benin, Benin City, Nigeria.

#### Availability of data and materials

The datasets used and/or analyzed during the current study are available from the corresponding author on reasonable request.

#### Conflict of Interest

No conflict of interest associated with this work.

#### **Contribution of Authors**

We declare that this work was done by the authors named in this article and all liabilities pertaining to claims relating to the content of this article will be borne by the authors. Valentine Adegoke designed the study. Valentine Adegoke, Emmar Okpakpor and Dickson Uwaya carried out the experiments and collection of data. Valentine Adegoke and Raymond Ozolua analysed and interpreted the data. Valentine Adegoke drafted the manuscript. Raymond Ozolua and Oluwakanyinsola Salawu supervised the experiments, data collection, analysis, interpretation of data and discussion of findings. All authors read and approved the manuscript for publication.

#### **Open Access**

This is an Open Access article that uses a funding model which does not charge readers or their institutions for access and distributed under the terms of the Creative Commons Attribution License (http://creativecommons.org/licenses/by/4.0) and the Budapest Open Access Initiative (http://www.budapestopenaccessinitiative.org/read), which permit unrestricted use, distribution, and reproduction in any medium, provided the original work is properly credited.

#### **REFERENCES**

- Abere TA, Ojogwu OK, Agoreyo FO, Eze GI. Antisickling and toxicological evaluation of the leaves of Rauwolfia vomitoria Afzel (Apocynaceae). J Sci Pract Pharm 2014; 1(1): 11-15.
- Eteng MU, Ibekwe HA, Abolaji AO, Okoi AI, Onwuka FC, Osuchukwu NC. Effect of Rauwolfia vomitoria Afzel (Apocynaceae) extract on serum aminotransferase and alkaline phosphatase activities and selected indices of liver and kidney functions. Afr J Biotechnol 2009; 8: 4604-4607.
- Egunyomi AJ, Moody O, Eletu O. Anti-sickling activities of two ethnomedicinal plant recipes used for the management of sickle cell anemia in Ibadan, Nigeria. Afr J Biotechnol 2009; 8(1):20-25.
- National Institute of Health. Public Health Service Policy on Humane Care and Use of Laboratory Animals, USA 2002.
- AOAC. Association of Official Analytical Chemists: Official Methods of Analysis. 1990; 16th Edition., Washington, DC, 1: 600-792.
- 6. Lorke D. A new approach to practical acute toxicity testing. Arch Toxicol 1983; 54(4): 275–287.
- 7. Ozolua RI, Uwaya DO. Laboratory-based safety assessment tests of some Nigerian commercial herbal products. J Pharmacovigilance S 2013; 1: 2.
- 8. Ebuehi OAT, Asoro I, Igwo-Ezikpe MN, Imaga NO, Erukainure OL, Lawal RA, Adenekan S, Duncan U, Micah C. Comparative toxicity studies of Rauwolfia vomitoria leaf and root extracts in wistar rats. IJBMR 2018; 9(2): 6357-6362.
- Schlede E, Genschow E, Spielmann H, Stropp G, Kayser
   D. Oral acute toxic class method: A successful
   alternative to the oral LD50 test. Regul Toxicol
   Pharmacol 2005; 42(1): 15-23.
- Isaiah AM, Olawale O, Effiong EE, Idongesit NJ, Fidelis UA, Friday UU. Vitamin E supplementation with Rauwolfia vomitoria root bark extract improves hematological indices. N Am J Med Sci 2012; 4(2): 86-86
- 11. Bonheur YDD, Désiré DDP, Antoine KS, David F, Théophile D. Acute and sub-acute toxicity of the aqueous extract of the stem bark of Rauwolfia vomitoria

- (Apocynaceae) in Wistar rats. WJARR 2020; 8(3): 373-385
- 12. Al-Attar AM, Elnaggar MH, Almalki EA. Physiological study on the influence of some plant oils in rats exposed to a sub-lethal concentration of diazinon. Saudi J Biol Sci 2018; 25(4): 786-796
- 13. Ezejindu DN, Chinweife KC, Uloneme GC. The effect of Rauwolfia vomitoria extract on liver enzymes of potassium-induced hepatotoxicity in adult Wistar rats. Intl J Biomed Adv Res 2013; 04(12)
- 14. Abubakar S, Sule M. Effect of oral administration of aqueous extract of cassia occidentalis I. seeds on serum electrolytes concentration in rats. BAJOPAS 2010; 3(1)
- 15. Akpanabiatu MI, Umoh IB, Edet EE, Ekanem T, Ukaffia S, Ndem JI. Effects of interaction of vitamin A and Rauwolfia vomitoria root bark extract on marker enzymes of cardiac diseases. Indian J Clin Biochem 2009; 24(3): 241-244.
- 16. Thammitiyagodage MG, De Silva NR, Rathnayake C, Karunakaran R, Wgss K, Gunatillka MM, Ekanayaka N, Galhena BP, Thabrew MI. Biochemical and histopathological changes in Wistar rats after consumption of boiled and un-boiled water from high and low disease prevalent areas for chronic kidney disease of unknown etiology (CKDu) in North Central Province (NCP) and its comparison with low disease prevalent Colombo, Sri Lanka. BMC Nephrol 2020; 21(1): 1-12.
- 17. Matsuzawa T, Nomura M, Unno T. Clinical pathology reference ranges of laboratory animals. J Vet Med Sci 1993; 55(3): 351-362.
- Yakubu MT, Bilbis LS, Lawal M, Akanji MA. Evaluation of selected parameters of rat liver and kidney function following repeated administration of yohimbine. Biochem 2003; 15(2): 50-56

Trop J Pharm Res, October 2024; 23(10): 1646